

Lixisenatide Attenuates the Detrimental Effects of Amyloid β Protein on Spatial Working Memory and Hippocampal Neurons in Rats

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Abstract

Type 2 diabetes mellitus (T2DM) is a risk factor of Alzheimer's disease (AD), which is most likely linked to impairments of insulin signaling in the brain. Hence, drugs enhancing insulin signaling may have therapeutic potential for AD. Lixisenatide, a novel long-lasting glucagon-like peptide 1 (GLP-1) analogue, facilitates insulin signaling and has neuroprotective properties. We previously reported the protective effects of lixisenatide on memory formation and synaptic plasticity. Here, we describe additional key neuroprotective properties of lixisenatide and its possible molecular and cellular mechanisms against AD-related impairments in rats. The results show that lixisenatide effectively alleviated amyloid β protein (A β) 25-35-induced working memory impairment, reversed A β 25-35-triggered cytotoxicity on hippocampal cell cultures, and prevented against A β 25-35-induced suppression of the Akt-MEK1/2 signaling pathway. Lixisenatide also reduced the A β 25-35 acute application induced intracellular calcium overload, which was abolished by U0126, a specific MEK1/2 inhibitor. These results further confirmed the neuroprotective and cytoprotective action of lixisenatide against A β -induced impairments, suggesting that the protective effects of lixisenatide may involve the activation of the Akt-MEK1/2 signaling pathway and the regulation of intracellular calcium homeostasis.

Key words: Lixisenatide; Amyloid β protein; Working memory; Cell viability; Akt-MEK1/2 signal pathway; Intracellular calcium concentration

1. Introduction

Alzheimer's disease (AD) is a progressive neurodegenerative disorder, afflicting more than 40 million people all around the world (World Alzheimer Report, 2015). It is characterized by memory loss and cognitive decline (Selkoe, 2001). One of the prominent events in the pathogenesis of AD is the formation of abundant deposits of senile plaques composed of amyloid β protein (A β) (Arnold et al., 1991; Thal et al., 2002). The toxicity of A β has been widely reported. For example, prolonged infusion of synthetic A β into the brain impaired learning and memory in rats, including working memory and place learning in the eight-arm radial maze, Y-maze and water maze (Nitta et al., 1997; Stepanichev et al., 2005); A β destabilized neuronal calcium homeostasis, altered cellular ionic activity, and rendered neurons more vulnerable to toxic stimuli in human cerebral cortical cell cultures (Fraser et al., 1997; LaFerla, 2002); A β acutely impaired synaptic functions when added to hippocampal slices or slice cultures (Li et al., 2009). Not only full length A β but also its fragments, such as A β ₂₅₋₃₅ and A β ₃₁₋₃₅, could induce apoptosis in cultured cortical neurons (Yan et al., 1999), in vivo LTP (Gault and Holscher, 2008), and cognitive deficits in rats (Alkam et al., 2007; Yuan et al., 2016). Unfortunately, effective neuroprotective strategies against A β neurotoxicity are still lacking.

Epidemiological studies found a clear correlation between type 2 diabetes mellitus (T2DM) and AD, in which T2DM has been identified as a risk factor for AD and the insulin resistance in the brain might initiate or accelerate the development of AD (Luchsinger et al., 2004; Ohara et al., 2011; Talbot et al., 2012). Based on these observations, it might be a promising strategy to normalize insulin signaling in the brain for the prevention and treatment of AD. Glucagon-like peptide 1 (GLP-1), an incretin hormone, and GLP-1 analogues that are protease resistant, have been reported to be able to cross the blood-brain barrier (BBB) and it facilitates insulin signaling (Gengler et al., 2012; Holscher, 2014a; Hunter and Holscher, 2012a). Furthermore, it has been found that GLP-1 and the GLP-1 receptor (GLP-1R) are expressed in the rodent central nervous system (Cork et al., 2015; Merchenthaler et al., 1999), especially the memory-related brain regions, such as the hippocampus (Cork et al., 2015; During et al., 2003; Hamilton and Holscher, 2009). More

importantly, GLP-1 possesses neurotrophic properties (Holscher, 2014b; Perry et al., 2002b), and could protect neurons against glutamate-induced apoptosis and even decrease endogenous A β accumulation in cellular and animal models of AD (Li et al., 2010; McClean et al., 2011b). However, endogenous GLP-1 is broken down quickly and lasts for only several minutes in blood plasma, which greatly limits its application in clinical practice (Deacon et al., 1995). Currently, several GLP-1 mimetics are on the market (e.g. exendin-4 (Byetta), liraglutide (Victoza) and lixisenatide (Lyxumia)). They have been engineered and developed to resist protease degradation (Christensen et al., 2011; Holst, 2004; Madsbad et al., 2008; Vilsboll, 2009). These GLP-1R agonists enhance cognition and reduce blood glucose levels in T2DM models (Gumuslu et al., 2016), but do not affect blood glucose levels in non-diabetic animals or people (McClean and Holscher, 2014). It has been reported that exendin-4 enhanced nerve growth factor (NGF)-induced cell differentiation into neurons (Perry et al., 2002a), and showed pronounced neuroprotective and anti-inflammatory effects in middle-aged diabetic animals (Darsalia et al., 2012), reversed T2DM-induced neuronal pathology in the piriform cortex of the rat (Lietzau et al., 2016), and antagonized A β 1-42-induced suppression of hippocampal long-term potentiation in rats (Wang et al., 2015). Liraglutide also enhanced synaptic plasticity in the hippocampus of the APP/PS1 AD mouse model (McClean et al., 2010), reduced the chronic inflammation response (McClean et al., 2011a; Parthasarathy and Holscher, 2013), and ameliorated working memory impairment in the A β -induced AD mouse model (Qi et al., 2016). Lixisenatide, a novel long-lasting GLP-1 analogue, has a higher permeability across the BBB and greater biological activity than exendin-4 and liraglutide (Hunter and Holscher, 2012b; Liu et al., 2015). Our previous studies (Cai et al., 2014) showed that lixisenatide prevented A β 25-35 -induced spatial memory deficits and hippocampal LTP suppression in rats. In the present study, we first observed the effects of lixisenatide on A β -induced deficits in spatial working memory of rats and cultured cell viability by using Y maze test and primary hippocampal neuronal cultures. Furthermore, the molecular and cellular mechanisms underlying the neuroprotective role of lixisenatide were investigated by using the Western blotting technique and intracellular calcium imaging.

2. Methods

2.1 Animals and drugs

Adult male Sprague Dawley (SD) rats (200–230 g) were provided by the Animal Research Center of Shanxi Medical University. All animal handling and procedures accorded with the guidelines of the Shanxi Animal Research Ethics Committee. During experiments, rats were kept at controlled room temperature (20–24°C) and humidity (60%–80%). A β 25–35 and lixisenatide (Sigma, St. Louis, MO, USA) were stored in dry form and dissolved in saline (5 nmol/ μ l) before experiments.

2.2 Intrahippocampal injection

Intrahippocampal injection was performed as previously described (Ryu and McLarnon, 2006, 2008). In brief, SD rats were anesthetized (chloral hydrate, 0.3 g/kg, i.p.) and placed in a stereotaxic apparatus (Narishige, Tokyo, Japan). Lixisenatide (5 nmol/ μ l) or vehicle (0.9% NaCl) and A β 25–35 (5 nmol/ μ l) or saline (0.9% NaCl) were bilaterally injected into the hippocampi (anterior-posterior: -3.0 mm; medial-lateral: \pm 2.2 mm; dorso-ventral: -3.0 mm, from bregma), with an injection rate of 0.2 μ l/min under the control of a micropump (KD Scientific, Inc, KDS310 Plus, USA). In co-application group, lixisenatide was injected 15 min before A β 25–35 application. To make sure the drugs were fully dispersed into the hippocampus, a 5 min-retention of the injection syringe in the brain was given after every injection.

2.3 Y maze test

Spontaneous alternation of Y maze was performed to examine the spatial working memory of rats. Rats were examined 10 days after drug administration. The Y maze has three radial arms (A, B, C) with the same angle (120°) between arms. Each arm is 45 cm long and 12 cm high. Rats were put into the junction of three arms and allowed to move freely for 8 min session. The entries of rats into each arm were recorded and every entry different from last two entries was considered as a successful alternation. The alternation

percentage was calculated according to the following: [(number of alternations)/(total number of arm entries - 2)] × 100(%) (Iwai et al., 2014).

2.4 Primary hippocampal neuronal cell culture

Primary hippocampal formation cultures were prepared from 24 hours postnatal SD rats. Briefly, pups were anesthetized with ether, and sterilized in 75% ethanol. Rat brains were quickly removed into ice-cold dissection solution. The hippocampi were stripped and cut into small pieces (each cube <1 mm³). The tissue pieces were incubated with 0.125% trypsin at 37°C for 15-20 min, and then complete culture medium was added to stop enzymatic reaction. Single-cell suspensions were obtained by mechanical dissociation using a Pasteur pipette with a fire-polished tip in complete culture medium, and then filtered through a 200 mesh nylon screen, centrifuged for 5 min at 1000 rpm. The cells at the bottom were resuspended with fresh complete culture medium and plated on poly-D-lysine coated 96-well plates and 35 mm culture dishes (Corning Inc) at a density of 5×10⁵ cells/ml. Cultures were maintained in 5% CO₂ at 37°C in complete culture medium for 6 h. The culture media were then changed to serum-free B27/neurobasal medium. Afterward, half of the culture medium was replaced with fresh serum-free B27/neurobasal medium every 3 days. 7-10 days after plating, the mature cells were used for further experimental observation. The experiment was all carried out under sterile condition.

2.5 Cell viability assay

To assess the rate of cell viability, cell counting kit-8 (CCK-8) and lactate dehydrogenase (LDH) were used. Briefly, cells were inoculated in 96-well plates. After 7-10 days, cells were incubated with Aβ₂₅₋₃₅ (10 μM) for 24 hours. In co-application group, Lixisenatide (100 μM) were applied 24 hours before Aβ₂₅₋₃₅ treatment. Six wells were used for each sample, and each experiment was repeated three times. CCK-8 solution (10 μl) was added to each well, then cells were incubated at 37°C for 2 h. The absorbance of samples was measured by a microplate reader (SpectraMAX 190, Molecular Devices,

Sunnyvale, CA) at 450 nm. Supernatant was collected to measure the LDH (a stable cytosolic enzyme, which is released upon cell lysis) release according to the manufacturer's instructions.

2.6 Western blotting

Cell lysate containing 5 µg of protein was separated on 4–12% gel and electrophoresed in running buffer at 200 mV for 35 min, followed by transfer to polyvinylidenedifluoride membrane. The membrane was washed in 1X TBST and blocked in 5% skimmed milk for 2 h at 25 °C. The membrane was then incubated with anti-pAkt (Ser473) (1:400) and anti-phospho-MEK1/2 (Ser217/221) (1:1000) antibodies at 4 °C overnight. All the primary antibodies used were generated in rabbit. β-actin was used as loading control. The membrane was incubated with 1:400 horseradish peroxidase-conjugated anti-rabbit IgG for 2 h at 25 °C. Relative density of bands was calculated.

2.7 Confocal calcium imaging

Intracellular free Ca²⁺ was measured with fluorescent dye Fluo-3/AM utilizing confocal laser scanning microscope (CLSM) (Olympus, FV-1000). Briefly, cultures were washed three times with balanced salt solution. Then, cells were loaded with the same solution containing 5 µM fluorescent Ca²⁺ indicator dye Fluo-3/AM for 1 h in a dark place at 37 °C. To remove the extracellular Fluo-3/AM, the loaded cells were rinsed three times with the balanced salt solution. Live video calcium images of selected neurons under microscopic field were recorded with a CLSM. Excitation wavelength of 488 nm and emission wave length of 526 nm were selected for the Fluo-3/AM fluorescence. Images were typically collected at 10-sec interval and recorded at least 20 min. The fluorescence intensities associated with a given cell were determined from digitized images and the data were processed using Olympus image analysis software. The experiment was carried out in the dark.

2.8 Data analysis

All values in the experiments were expressed as means \pm standard errors (SEM). The SPSS 13.0 and SigmaPlot 11.0 statistical packages were used for statistical analyses. The data from Y maze test, cell viability assay and western blotting were examined by a two-way repeated measures analysis of variance (ANOVA). The data from confocal calcium imaging was performed by using one-way ANOVA and Student-t test. The statistical significance level was defined as $p < 0.05$.

3. Results

3.1 Lixisenatide reversed the A β 25-35-induced impairment of spatial working memory

Two-way ANOVA showed A β 25-35 injection and lixisenatide pretreatment had significant main effects on the spontaneous alternation of rats (A β 25-35: $F(1,36) = 139.156$, $P < 0.001$; lixisenatide: $F(1,36) = 137.718$, $P < 0.001$; A β 25-35 by lixisenatide: $F(1,36) = 163.483$, $P < 0.001$). As shown in the Figure 1A, Tukey's post hoc tests showed that A β 25-35 significantly reduced the percentage of right turns of rats ($P < 0.001$, $n = 10$) and lixisenatide reversed this detrimental effect ($P < 0.001$, $n = 10$). At the same time, the total arm entries of rats did not show any significant difference among these groups ($P > 0.05$; Figure 1B).

3.2 Lixisenatide inhibited the A β 25-35-induced cytotoxicity on cultured hippocampal cells

Two-way ANOVA showed A β 25-35 injection and lixisenatide pretreatment had significant main effects on the viability of cultured hippocampal formation by CCK-8 assay (A β 25-35: $F(1,36) = 382.892$, $P < 0.001$; lixisenatide: $F(1,36) = 180.696$, $P < 0.001$; A β 25-35 by lixisenatide: $F(1,36) = 203.032$, $P < 0.001$). As shown in the Figure 2A, Tukey's post hoc tests showed that A β 25-35 significantly decreased the viability of hippocampal formation cultures ($P < 0.001$, $n = 10$) and lixisenatide reversed this toxicity

effect ($P < 0.001$, $n = 10$). Two-way ANOVA showed A β 25-35 injection and lixisenatide pretreatment had significant toxic effects on the viability of cultured hippocampal formation by LDH assay (A β 25-35: $F(1,36) = 330.942$, $P < 0.001$; lixisenatide: $F(1,36) = 183.66$, $P < 0.001$; A β 25-35 by lixisenatide: $F(1,36) = 111.299$, $P < 0.001$). As shown in the Figure 2B, Tukey's post hoc tests showed that A β 25-35 significantly induced the death of cultured hippocampal formation ($P < 0.001$, $n = 10$) and lixisenatide weakened this bad effect ($P < 0.001$, $n = 10$).

3.3 Lixisenatide relieved the A β 25-35-induced suppression of the Akt-MEK1/2 signaling pathway

The neuroprotection of lixisenatide might be related to the Akt-MEK1/2 signal pathway. Two-way ANOVA showed A β 25-35 injection and lixisenatide pretreatment had significant main effects on the level of pAkt (A β 25-35: $F(1,36) = 6.984$, $P < 0.05$; lixisenatide: $F(1,36) = 8.492$, $P < 0.05$; A β 25-35 by lixisenatide: $F(1,36) = 9.398$, $P < 0.05$). As shown in the Figure 3A, Tukey's post hoc tests showed that A β 25-35 significantly decreased the level of pAkt ($P < 0.05$, $n = 10$) and lixisenatide reversed this decrease ($P < 0.05$, $n = 10$). Two-way ANOVA showed A β 25-35 injection and lixisenatide pretreatment had significant main effects on the level of pMEK1/2 (A β 25-35: $F(1,36) = 7.429$, $P < 0.05$; lixisenatide: $F(1,36) = 4.499$, $P < 0.05$; A β 25-35 by lixisenatide: $F(1,36) = 5.409$, $P < 0.05$). As shown in the Figure 3B, Tukey's post hoc tests showed that A β 25-35 induced a significant decrease in the level of MEK1/2 ($P < 0.05$, $n = 10$) and this decrease was reversed by lixisenatide ($P < 0.05$, $n = 10$).

3.4 Lixisenatide significantly protected against the A β 25-35-induced elevation of $[Ca^{2+}]_i$, which was abolished by a MEK1/2 inhibitor

The cell mechanism of A β -induced neurotoxicity involves in the perturbation of Ca^{2+} homeostasis. Firstly, we investigated the change of $[Ca^{2+}]_i$ level in rat primary cultured hippocampal neuron by applying 10 μ M A β 25-35. As shown in the Figure 4B, the relative fluorescent intensity at resting condition in the control group ($n = 20$) was very stable, nearly

showing as a straight horizontal line. After application of A β 25-35 (n=20), the fluorescent intensity in most of neurons gradually and persistently increased during all of recording time. Figure 4C showed the relative fluorescent intensity values of [Ca²⁺]_i in different experimental groups at 20 min after application of A β 25-35. Obviously, A β 25-35 increased the relative fluorescent intensity of [Ca²⁺]_i, being 185.34 \pm 3.2%, significantly higher than the value in the control group ($P<0.01$). This result indicated that A β 25-35 could increase [Ca²⁺]_i, which might contribute to the neurotoxicity of A β seen in cultured hippocampal neurons. Further, we investigated the pretreatment effect of lixisenatide on A β 25-35 induced [Ca²⁺]_i elevation. As shown in the Figure 4C, A β 25-35-induced elevation of [Ca²⁺]_i level was mostly reversed by lixisenatide (100 μ M) and the relative fluorescent intensity decreased to 134.01 \pm 2.7% (n=20, $P<0.01$). These results indicated that lixisenatide could protect against A β 25-35-induced intracellular calcium overloading.

To investigate the molecular mechanism of the protection of lixisenatide against A β 25-35, we also observed the effect of lixisenatide on A β 25-35-induced [Ca²⁺]_i elevation in the presence of U0126, a MEK1/2 inhibitor. The result showed that treatment with U0126 for 20 min, essentially blocked the protective effect of lixisenatide against A β 25-35 induced [Ca²⁺]_i elevation. Compared with co-application of lixisenatide and A β 25-35, the relative fluorescent intensity in co-application of U0126 (1 mM), lixisenatide (100 μ M) plus A β 25-35 group increased to 166.18 \pm 3.1% (n=20, $P<0.01$). These results indicated that the protective role of lixisenatide against A β 25-35 induced [Ca²⁺]_i elevation might be closely associated with the activation of MEK1/2.

4. Discussion

AD is a biologically complex neurodegenerative syndrome. Nearly 20 years ago, with the combination of observations from biochemistry, neuropathology and genetics, a compelling hypothesis known as the amyloid cascade hypothesis was formulated. Although growing amounts of data are inconsistent with the basically linear causal chain of events of this hypothesis (Herrup, 2015), it has been recognized that A β plays a key part in the early

development of the pathogenesis of AD. The amyloid plaque load does not correlate with disease progression at a later stage of disease (Edison et al., 2008). The neurotoxicity of different A β fragments such as A β 1-42 and A β 1-40 was observed in cell culture first. A β peptide- induced cell apoptosis and death in cultured cortical neurons is a common observation (Deshpande et al., 2006). The sequence 25-35 of A β is further recognized as an active site of A β , as this fragment shows similar neurotoxic properties to the full length of A β (Holscher et al., 2007). A β 25-35 can induce cell apoptosis and necrosis in cultured hippocampal neurons (Sendrowski et al., 2015). Therefore, the present study used A β 25-35 in primary cell culture and behavioral experiments. For the cell viability assay, mature neurons were exposed to A β 25–35 for 24 hours. For behavioral memory testing, a Y maze test was performed. The injected solution includes soluble monomers, dimers, and oligomers. It has been proposed that soluble oligomeric forms of A β were the chief mediators of cytotoxicity in AD (Ashe and Aguzzi, 2013; Selkoe, 2002; Wisniewski and Goni, 2015). Other configurations of amyloid also can induce neuronal death (Orellana et al., 2011). GLP-1 is an incretin hormone which effectively re-sensitizes insulin signaling and physiological activity by increased expression of the insulin receptor gene (Doyle and Egan, 2007). Another benefit for using GLP-1 in AD treatment is that GLP-1 does not affect blood glucose level in normoglycemic people (Gallwitz, 2006; Vella et al., 2002). However, natural GLP-1 lasts only several minutes in blood plasma because of DPP4-induced degradation (Deacon et al., 1995). Fortunately, many GLP-1 analogues with longer half-life, such as lixisenatide used in the present study, have been developed and even applied to clinical treatment (Elkinson and Keating, 2013; Holst, 2004; Shyangdan et al., 2011).

Memory dysfunction is a prominent symptom of patients with AD (Webster et al., 2014). Spontaneous alternation test of rats with Y maze is a hippocampal dependent short-term spatial working memory (Stuchlik et al., 2014). In the present study, our behavioral results show that bilateral intrahippocampal injection of A β 25-35 significantly decrease the percentage of alternation of rats, and lixisenatide reversed this detrimental effect. At the same time, the total arm entries of rats did not show any significant

differences among these groups, suggesting the difference in spontaneous alternation among these groups was due to the change in spatial working memory, but not impairing general locomotor activity of rats.

In cell culture and cell viability experiments, we found that the cell viability of hippocampal cell cultures significantly decreased in A β 25-35 alone group, which is consistent with the behavioral results in the Y maze test. Interestingly, lixisenatide inhibited A β 25-35-induced cytotoxicity, with a significant viability recovery in the lixisenatide plus A β 25-35 group, compared with A β 25-35 only group. The Akt-MEK1/2 signaling pathway is important for cell survival (Wang et al., 2010). The suppression of the Akt-MEK1/2 signaling pathway would activate the downstream apoptosis pathway and promote cell apoptosis (Cervellati et al., 2014; Nunomura et al., 2001). Accumulation of A β , which induces cytotoxicity and behavior impairments, may lead to a decrease of the level of Akt and of MEK1/2 activity (Kashour et al., 2003; Ryder et al., 2004). Our experiment shows that lixisenatide prevents the A β 25-35 induced decrease of pAkt and pMEK1/2 levels, indicating that activation of the Akt-MEK1/2 signaling pathway might be involved in the protective action of lixisenatide against A β toxicity.

A number of studies have shown that perturbed cellular Ca²⁺ homeostasis may play a prominent role in neuronal death in AD (LaFerla, 2002). The elevation of intracellular calcium mediated the neurotoxicity induced by A β in primary cortical neurons (Fu et al., 2006) and hippocampal neurons (Kelly and Ferreira, 2006). In this study, we observed the effect of A β 25-35 on the intracellular Ca²⁺ level in cultured primary rat hippocampal neurons. Our results showed that A β 25-35 significantly increased the intracellular Ca²⁺ level, while lixisenatide pretreatment effectively suppressed A β 25-35-induced Ca²⁺ elevation, demonstrating the association between A β and intracellular calcium overload, and suggesting the protection of lixisenatide might be due to alleviating A β -induced intracellular calcium overload. We further explored the effect of U0126 (a specific MEK1/2 inhibitor, with the cell permeability) in the lixisenatide plus A β 25-35 group. Our results indicate that the effect of lixisenatide on calcium homeostasis was mostly blocked in the presence of U0126, which might suggest that the MEK1/2 signaling pathway is involved in

the process of calcium homeostasis. This result confirms our previous cell culture study that demonstrates a normalizing effect of liraglutide in SH-SY5Y cells via activating Akt and MEK1/2 (Sharma et al., 2013). Furthermore, the observed effects explain in part the Neuroprotective effects of lixisenatide in the APP/PS1 mouse model of AD that we have found previously (McClellan and Holscher, 2014).

In summary, the present study confirmed the previous reports of the neuroprotective and cytoprotective action of lixisenatide against $A\beta$, suggesting that the protection of lixisenatide may be involved in the activation of Akt-MEK1/2 signaling pathway and the regulation of intracellular calcium homeostasis.

Figure legends

Figure 1. Lixisenatide alleviated the detrimental effect induced by A β 25-35 in Y maze working memory test.

A, histograms showing the lower spontaneous alternation of rats after injection of A β 25-35 (** p <0.001, n=10) and a significant recovery in A β 25-35 plus lixisenatide group (** p <0.001, n=10). B, histograms showing no significant difference in total arm entries among different groups.

Figure 2. Lixisenatide inhibited the toxicity of A β 25-35 on cultured hippocampal neurons.

Cells were incubated with A β 25-35 (10 μ M) in the absence or presence of lixisenatide (100 μ M) for 24 h. The cell viability and cytotoxicity were tested with CCK-8 and LDH assay, respectively. A, Histograms showing the lower viability of cultured hippocampal neurons in A β 25-35 plus vehicle group (n=10) and a significant reverse in A β 25-35 plus lixisenatide group (n=10). B, Histograms showing the higher toxicity in A β 25-35 plus vehicle group (n=10) and a significant recovery in A β 25-35 plus lixisenatide group (n=10). *** p <0.001

Figure 3. Lixisenatide prevented A β 25-35 induced down regulation of pAKT and pMEK1/2.

A, histograms showing the lower level of pAKT in A β 25-35 plus vehicle group (n=10) and a significant restoration in A β 25-35 plus lixisenatide group (n=10). B, histograms showing the decreased level of pMEK1/2 in A β 25-35 plus vehicle group (n=10) and a significant reverse in A β 25-35 plus lixisenatide group (n=10). * p <0.05

Figure 4. Lixisenatide protected against A β 25-35-induced [Ca²⁺]_i elevation, which was abolished by U0126.

A, Representative fluorescent $[Ca^{2+}]_i$ imaging of cultured hippocampal neurons a, before A β 25-35 injection; b, 20 min after A β 25-35 injection. Scale length: 20 μ m. B, Plots showing representative fluorescent intensity traces in different groups. C, histograms showing the relative fluorescent intensity 20 min after A β 25-35 injection in different groups. ** $p < 0.01$.

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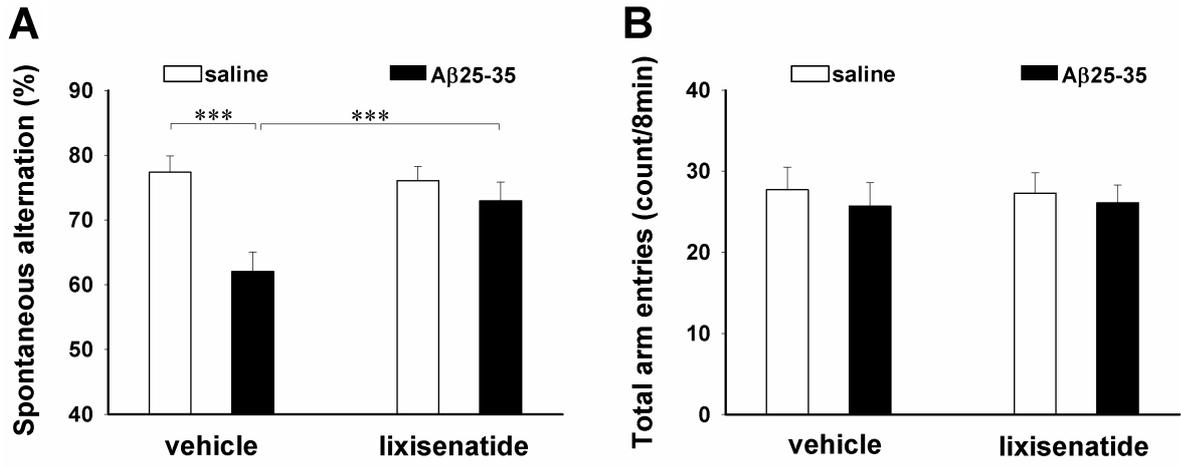


Fig. 1

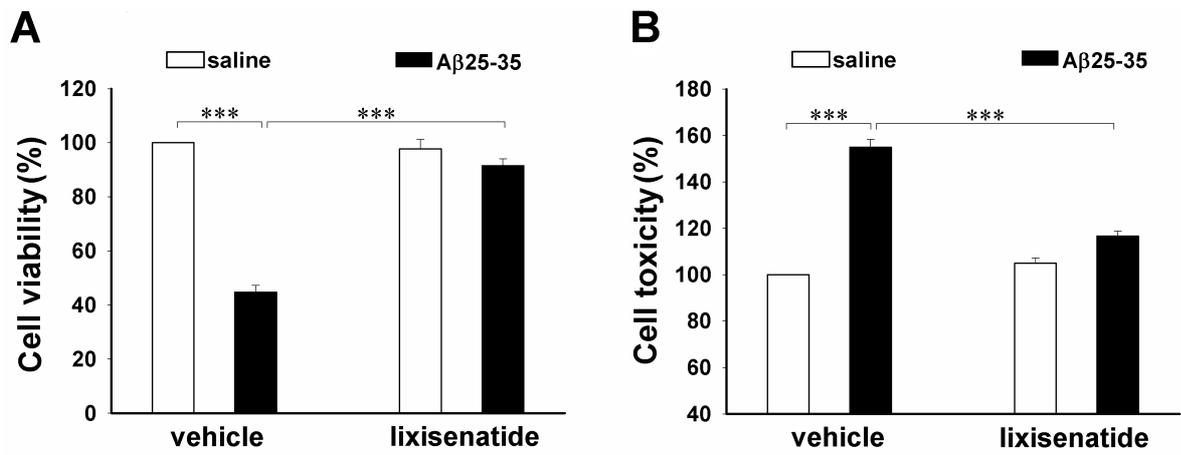


Fig. 2

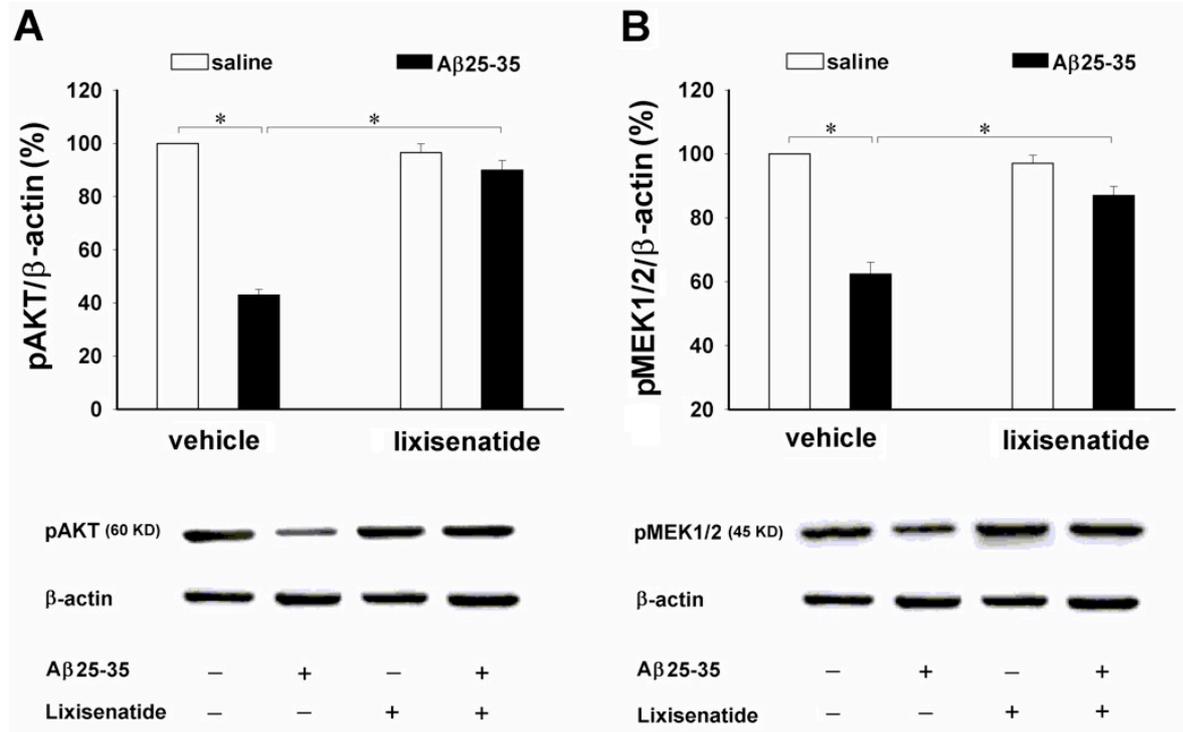


Fig. 3

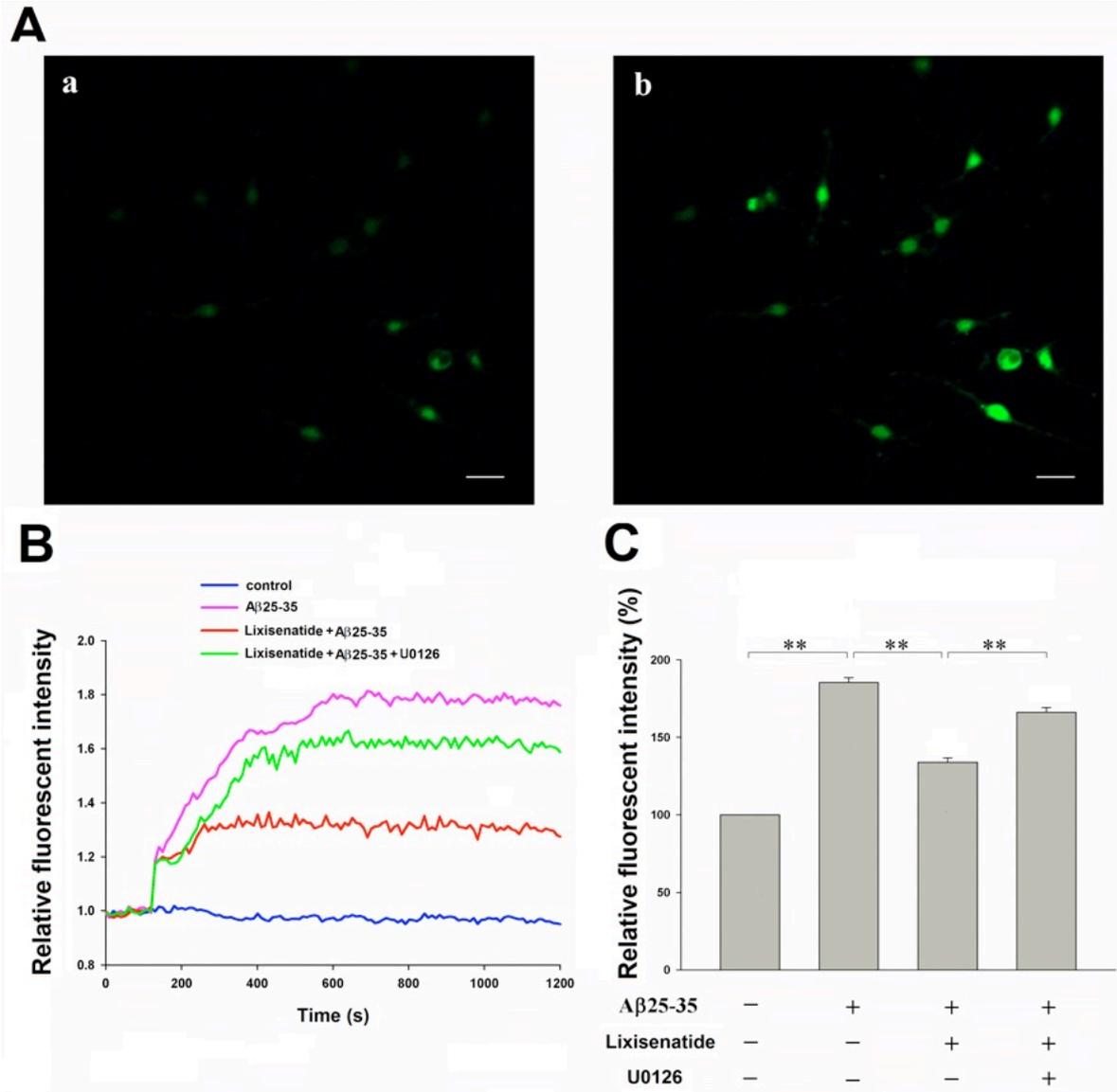


Fig 4