1	Selection of aprild prey by a generalist predator: Do prey chemical defences matter?
2	Nesbit, C.M.*a, Wilby, Aa., Roberts, M.R.a & Menéndez, R.a
3	<sup>a</sup> Lancaster University, Lancaster, Lancashire, UK, LA1.
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	*corresponding author: chrisnesbit.ecol@gmail.com
16	Tel: 07837 256697
17	
18	
19	Running Title: Do chemical defences affect prey selection?

### 20 Abstract

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

- For predators, prey selection should maximize nutrition and minimize fitness costs. Here, we investigate whether a generalist predator (Chrysoperla carnea lacewing larvae) rejected harmful, chemically-defended prey (Brevicoryne brassicae aphids) when non-defended prey (Myzus persicae aphids) were available. We tested: (1) the effect of consuming different prey species on predator mortality; (2) whether naïve predators reject chemically-defended prey during foraging when non-defended prey are available; (3) whether the relative abundance of each prey affects the predator's prey choice; and (4) whether predators learn to avoid consuming chemically-defended prey following exposure to both prey species. Consumption of B. brassicae yielded greater C. carnea mortality than M. persicae consumption, but naïve C. carnea did not reject B. brassicae in favour of M. persicae during foraging. When presented at unequal abundances, naïve predators generally consumed each aphid species according to their initial relative abundance, although, predation of non-defended prey was less than expected where defended prey were initially more abundant, indicating high consumption of B. brassicae impeded M. persicae consumption. With experience, C. carnea maintained predation of both aphid species but consumed more M. persicae than B. brassicae, indicating a change in behaviour. Although prey choice by C. carnea may change with experience of available prey, prey chemical defences do not appear to influence prey choice by naïve predators. This inability to avoid harmful prey could facilitate wider, indirect interactions. M. persicae may benefit where high consumption of B. brassicae hinders predators in the short term, and in the long term, increases predator mortality.
- 41 Keywords: Apparent commensalism, associational resistance, Brassicacae, predator
- behaviour, trait-mediated indirect interaction

#### INTRODUCTION

Predators can strongly affect the composition and persistence of their prey communities, and where prey are herbivorous, predators may indirectly affect plant diversity and biomass (Schmitz et al. 2000, Schmitz 2003, 2006). Through this mechanism (a 'trophic cascade'), predatory invertebrates deliver biological pest control, where predation of herbivorous crop pests improves or maintains crop plant production (Symondson et al. 2002, Snyder and Wise 2001). In many agricultural and natural communities, prey species co-occur in diverse assemblages, thus, to develop successful biological control schemes, it is necessary to predict how predatory invertebrates respond to the availability of multiple prey species. i.e., are predators selective in their prey choice? For predators, selecting the 'best quality available prey' means consuming prey that offer maximum nutrition and/or minimal costs such as greater risk of wounding if prey are structurally-defended, or poisoning if prey possess chemical-defences, acquired from their food plants (Forbes 1989, Nishida 2002, Hayward and Kerley 2005, Magalhães et al. 2005, Opitz and Muller 2009).

Predators may choose between available prey species when locating prey habitats, locating prey within a habitat and when accepting located prey (Hoy and Smilanick 1981, Vinson 1976), using a variety of different cues. When plants are attacked by herbivores, they release volatile olfactory cues as a signal to attract the herbivore's natural enemies (Zhu et al. 1999, Dicke and van Loon 2000, Glinwood et al. 2011). The blend of volatiles varies depending on the identity of the herbivore attacking the plant, thus, predators may use plant volatiles to locate prey habitats and potentially, to differentiate between plants hosting different prey species (although this is more common for parasitoids than predators, Hatano et al. 2008). Generalist predators more commonly locate and select their prey using cues associated with prey themselves. For example, the scent of aphid honeydew or aphid alarm pheromone ((E)-β-farnesene) can be attractive to predators such as *Episyrphus balteatus* 

(DeGeer) (Diptera: Syrphidae) hoverfly larvae and *Adalia bipunctata* (Linnaeus) (Coleoptera: Coccinellidae) ladybird larvae (Francis et al 2004, 2005), yet isothiocyanates released by *Brevicoryne brassicae* (Linnaeus) (Hempitera: Aphididae) aphids, as an indicator of chemical defences, can be repellent to *A. bipunctata* (Francis et al. 2005). Should predators successfully locate prey within a habitat, predators may register olfactory, or other cues (such as aposematic coloration) associated with chemically-defended prey as a warning signal against attack (Francis et al. 2004, Mappes et al. 2005). If cues are absent or not innately recognized and harmful prey is accepted, then assuming harmful effects are sub-lethal, predators may learn from experience to avoid the harmful prey when next foraging.

It is widely assumed that selectivity exhibited by generalist predators is a learned behaviour (Dukas 2008). Coccinelid beetles for example can learn to recognise suitable prey habitats and to select the most nutritious available prey. *Coccinella septempunctata* (Linnaeus) (Coleoptera: Coccinellidae) larvae were shown to associatively learn the odours of barley cultivars upon which it had previously located and fed on aphids (Glinwood et al. 2011), while *Coleomagilla maculata lengi* (Timberlake) (Coleoptera; Coccinellidae) have been shown to reject poorer-quality, parasitized *Pieris rapae* (Linnaeus) (Lepidoptera: Pieridae) eggs in favour of consuming more nutritious, unparasitized eggs at greater frequency with greater experience (Boivin et al. 2008). In both studies however, the behaviours were shown to be at least partially forgotten after forty-eight hours (Boivin et al. 2008; Glinwood et al. 2011). As prey that vary in levels of chemical defences can occur in the same communities as non-defended prey (Kalule and Wright 2002b, Staley et al. 2010, van Veen et al. 2009), we assess here whether a widespread predatory invertebrate rejects, or learns to reject a harmful, chemically-defended aphid species if a better quality, non-defended aphid species is available.

Among *Brassica* crops, predatory insects, such as ladybird, lacewing and hoverfly larvae can feed on two widespread pest aphid species that can occur together on the same plants, or in the same communities (Kalule and Wright 2002b, Snyder et al. 2006, 2008, Staley et al. 2010). *Myzus persicae* (Sulzer) (Hempitera: Aphididae) is a non-defended generalist aphid (Bridges et al. 2002), while *B. brassicae* is a specialist aphid that co-opts the glucosinolate defence chemicals of its host plant as a defence against predators (Bridges et al. 2002, Kazana et al. 2007). Continued consumption of *B. brassicae* can be lethal to, or inhibit the growth rates of generalist predators (Francis et al. 2001, Kos et al. 2011, 2012, Pratt et al. 2008); effects that can be stronger when *B. brassicae* feed on more chemically-defended crop plant varieties (Chaplin-Kramer et al. 2011, Kos et al. 2011).

The aim of this study was to assess whether the generalist predator *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae) lacewing larvae innately avoids, or learns to avoid consuming chemically-defended *B. brassicae* where *M. persicae* are available, and whether any selectivity behaviour varies depending on the variety of plant hosting the prey species. We predict that *C. carnea* will learn to reject *B. brassicae* in favour of *M. persicae* and for the strength of selectivity to vary depending on the aphid's host plant. First, we tested the assumption that *B. brassicae* are more harmful for *C. carnea* to consume than *M. persicae*. Second, we assessed the ability of naïve *C. carnea* to select between plants hosting different prey species (*B. brassicae* or *M. persicae*). Thirdly, we assessed whether innate prey choice of *C. carnea* was determined by the relative abundance of the two prey species, and finally, we tested whether *C. carnea* learn to avoid consuming *B. brassicae* through experience.

### **METHODS**

The study system included two aphid species, chemically-defended *Brevicoryne* brassicae and non-defended *Myzus persicae*; and the shared predator *Chrysoperla carnea* larvae. Two widely studied *Brassica oleracea* (Linnaeus) (Brassicales: Brassicaceae) cabbage cultivars, Derby Day and f<sub>1</sub> Minicole, were used as aphid host plants. Derby Day is considered an herbivory-susceptible cabbage cultivar (Ellis et al. 1996, Verkirk et al. 1998, Staley et al. 2009) and Minicole to possess some herbivory-resistance (Verkirk et al. 1998, Schuler and van Emden 2000, Hariprasad and van Emden 2010). Populations of both aphid species have been reported to be lower on Minicole compared to Derby Day plants in lab and field experiments (Kalule and Wright 2002a, 2002b; Nesbit 2013), suggesting a difference in antibiosis between cultivars.

Derby Day (Nicky's Nursery Ltd., UK) and Minicole (E.W. King & Co. Ltd., UK) seeds were sown individually in John Innes N°.2 compost in 15 cell seed trays (each cell 65 x 65 mm and 60 mm deep) and grown in a glasshouse with daily watering. *M. persicae* and *B. brassicae* were maintained in monocultures in a controlled environment room (hereafter, CE room) at Lancaster University (day/night temp (°C): 22/17, 15L:9D photoperiod) on plants of both cultivars. *C. carnea* larvae (2<sup>nd</sup> instar, supplied by Fargro Ltd., UK) were stored on arrival in buckwheat seed husks in a refrigerator at 4°C, for 3 days before each experiment began. Twenty-four hours prior to testing, *C. carnea* larvae were weighed (mg), transferred to numbered Petri dishes (5 cm diameter, containing a small filter paper saturated with water and one *Rhopalosiphim padi* aphid (from a culture maintained on wheat) to prevent death from starvation) and assigned to treatments.

## Predator survival on diets of different prey

To test the assumption that B. brassicae are more harmful for C. carnea to consume than M. persicae, diet assays were employed. Survival was measured for C. carnea larvae that were fed diets of M. persicae reared on Derby Day (MpDD), B. brassicae reared on Derby Day (BbDD, to test against MpDD for species effects), or B. brassicae reared on Minicole plants (BbM, to test against BbDD for B. brassicae-mediated host plant effects). Regrettably, our stock of M. persicae on Minicole was contaminated with Aphidius ervi (Haliday) (Hymenoptera: Braconidae) parasitoids at the time of this experiment, precluding the possibility of a fully factorial experiment (although this treatment was not strictly necessary to test our prediction). Ninety C. carnea were transferred individually to numbered plastic pots (4.5 cm diameter, 4 cm depth, with perforated lids) containing a filter paper saturated with water. Each C. carnea larva was fed in excess, with 20 mixed-age wingless aphids according to their diet treatment (30 replicates per treatment). Aphids were replaced every day, at which point the remains of dead aphids and all surviving aphids were removed. Survival of C. carnea was recorded at the end of the experimental period of 7 days. The experiment was repeated, giving 60 replicates per treatment in total. Predators were not used more than once.

156

157

158

159

160

161

162

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

## Innate predator selectivity between different aphid host plants

To test whether naïve *C. carnea* select between aphid species while locating aphid host plants, predators were given free choice to visit a plant hosting *B. brassicae* and/or a plant hosting *M. persicae*. Each assay exposed one *C. carnea* larva to two plants, one host to *B. brassicae*, the other to *M. persicae*. Assays alternated in using Derby Day or Minicole as host plants. Six assays were conducted per day over three days for two weeks, giving a total

of 36 assays, 18 per plant cultivar. A new predator and new aphid host plants were used in each assay.

Twenty-four hours before each day of assays, plants and insects were pre-treated as follows: Six plants (6 week old) per cultivar were re-potted (10 cm diameter, 9 cm depth pots) and transferred to the CE room. Measurements of leaf number and height (mm, from base of the stem to the tip of the budding leaf) were used to assign plants to assays, so the two aphid host plants were of approximately equal size. Each plant was transferred to an individual insect rearing cage (30 cm diameter, approx. 60 cm high). A Petri dish (5 cm diameter) containing 20 mixed-age wingless aphids of the assigned aphid species was left at the base of the stem overnight for aphids to colonize.

The assay arena was a Perspex observation box (30 x 30 x 30 cm Bugdorm1, MegaView Science Co. Ltd, Taiwan, modified to have one transparent side to facilitate observation) adapted from Wilby et al. (2013). Inside were two plants of the same cultivar, one hosting *B. brassicae*, the other *M. persicae*, connected by a 15 cm Perspex bridge in contact with the base of their stems. A single *C. carnea* larva was placed in the centre of the bridge, which could visit one, both or neither of the plants. *C. carnea* were then observed for 45 minutes. Time and location were recorded when *C. carnea* visited a plant (defined as having made physical contact with the plant), left a plant, foraged (defined as actively seeking prey as opposed to resting immobile on the plant), captured an aphid or discarded a consumed aphid. The bridge was rinsed with tap water between assays and plant configuration (whether the plant with *B. brassicae* was on the left or right) alternated between assays to control for any directional effects.

To test whether innate prey choice of *C. carnea* was determined by the relative abundance of the two prey species, predators were exposed to mixed populations of *B. brassicae* and *M. persicae* at different relative abundances. *C. carnea* were randomly assigned to ten treatment combinations of plant cultivar (Derby Day or Minicole) crossed against aphid abundance ratio (number of *B. brassicae*: number of *M. persicae*: 0:20; 5:15; 10:10; 15:5; 20:0). Each treatment was replicated 3 times per day and the experiment was repeated over six days, giving 18 replicates per treatment in total.

On the day of testing, aphids were transferred to Petri dishes (5 cm diameter) according to the appropriate abundance ratio. Each Petri dish also contained a small water-saturated filter paper, and approximately 2 cm<sup>2</sup> of fresh, randomly excised leaf material from 8 week old plants of the treatment cultivar. *C. carnea* were transferred to the aphid Petri dishes, one per dish, and left for 5 hours, after which time the number of surviving aphids was counted.

### Predator associative learning

To test whether *C. carnea* learn to avoid consuming *B. brassicae* through experience, predators were repeatedly exposed to mixed populations of *B. brassicae* and *M. persicae* (at equal proportion). Thirty *C. carnea* larvae were exposed individually to mixed groups of 10 *B. brassicae* and 10 *M. persicae* in a Petri dish (5 cm diameter) for five hours, after which time the number of surviving aphids was counted. The same *C. carnea* larvae were then exposed to new, mixed-aphid groups, for the same five hour period, over four further consecutive days, thus, predators were re-used in this experiment. The experimental protocol was the same as described in previous section (involving a Petri dish, plant material and filter

paper), however, there were only two treatments of ten of each Derby Day-reared aphid species, or ten of each Minicole-reared aphid species (15 replicates per treatment).

## Statistical Analysis

Unless stated, data from the first and second experiments were analysed using generalised linear mixed effects models (GLMMs) fit to a binomial distribution with a logit link function. The significance of fixed effects was assessed by contrasting the deviance between models with and without the fixed effect using chi-squared test statistics (hereafter: analysis of deviance) (Crawley 2007, Zuur et al. 2009).

For the first experiment, survival of *C. carnea* (yes or no) fed MpDD and BbDD was contrasted, with aphid species as the fixed effect. Survival of *C. carnea* fed BbDD and BbM was contrasted with plant cultivar as the fixed effect (Crawley 2007; Zuur et al. 2009). Temporal block was included as a random effect (factor: 1 or 2).

For the second experiment, to test whether *C. carnea* discriminated between plants hosting different aphid species, we analysed whether a plant hosting one aphid species was consistently visited first in an assay over the other (yes or no). To test whether *C. carnea* registered any subsequent cues while foraging on plants, three further responses were analysed: (a) plant fidelity - whether the first visit was exclusive (yes or no) or if *C. carnea* also visited the other plant during the assay; (b) the time *C. carnea* spent foraging on a plant (minutes spent on the plant minus time spent feeding minus time spent immobile), and (c) whether predation occurred (yes or no). The fixed effects of all maximal models were aphid species, plant cultivar and the interaction term. The random effects of all models were the time of day (factor: 1:6), nested within the day (factor: Wednesday/Thursday/Friday), nested within the week the assay was conducted (factor: week 1 or 2). Foraging time (mins) was

analysed using linear mixed effects (LME) models and analysis of deviance under maximum likelihood (ML) parameter estimation.

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

For the third experiment, we assumed that if C. carnea were unselective they would be expected to encounter and consume aphids of each species in proportion to their initial abundance in mixed-prey populations, weighted by their respective feeding rates on the two aphid species when presented alone to account for differences between prey species in handling time or satiation. Differences in handling time or satiation between aphid species were analysed using single-species treatment data. Aphid count was analysed using GLMMs with Poisson errors and log-link function. Fixed effects in the maximal model included aphid species, plant cultivar and the interaction term. The random effect was predator weight (mg). The significance of fixed effects was assessed through analysis of deviance (Zuur et al. 2009). The mean final count of each aphid species per single-species treatment was obtained using the parameter estimates from the minimum adequate model, and was used to obtain an expected final count in the mixed-aphid treatments for each aphid species: Expected final aphid count (mixed-species treatment) = [final count (single-species treatment)/initial count (single-species treatment)] \* initial count (mixed-species treatment) Data from mixed-aphid treatments were used to test whether counts of each aphid species when presented together deviated from expected counts. The response was: Deviation from expected count = ln(observed final aphid count/expected final aphid count) Fixed effects in the maximal LME model included aphid species, plant cultivar, aphid abundance ratio (initial B. brassicae: M. persicae: 5:15; 10:10; 15:5) and all interaction

terms. Random effects included predator identification number, to account for aphid counts

from the same test, and predator weight. The significance of fixed effects was assessed by analysis of deviance under ML parameter estimation (Zuur et al. 2009).

For the fourth experiment, data were split to test for (a) innate prey selection, using only data from the first day when predators were naïve, and (b) selection resulting from experience, using data from predators having been previously exposed to the aphids (1 to 4 exposures). All final count response data were analysed using GLMMs fit to a Poisson distribution with a log-link function. Fixed effects for the innate prey selection analysis included aphid species, plant cultivar and the interaction term, with random effects of predator identification number (factor 1:30) to account for aphid counts from the same test, and the starting predator weight. For the analysis of prey selection of experienced predators, fixed effects included aphid species, plant cultivar, previous exposures (1 to 4) and all interaction terms. The random effects included initial predator weight and a grouping term to account for repeated measurements from the same predator (predator identification number) across successive exposures.

All analyses used the 'lme4' package (Bates et al. 2012) for 'R.v.2.15.2' (R Core Development Team 2012). Adequacy of best-fitting LME models and GLMMs fit to a poisson distribution was assessed by inspecting the residuals against the fitted values, fixed effects and random effects (Zuur et al. 2009).

#### RESULTS

Predator survival on diets of different prey

Diet assays were used to test whether *B. brassicae* are more harmful for *C. carnea* to consume than *M. persicae*. Survival of *C. carnea* larvae was significantly greater for those fed *M. persicae* (mean survival: 95%,  $\pm$  SEM: 90-97%) compared to those fed chemically-

defended *B. brassicae* (mean survival: 83%,  $\pm$  SEM: 77-88%) from Derby Day plants ( $\chi^2_1$  = 4.435, p = 0.035). No significant difference was observed in survival of *C. carnea* given diets of *B. brassicae* reared on Minicole (mean survival: 80%,  $\pm$  SEM: 71-87%) or Derby Day (mean survival: 83%,  $\pm$  SEM: 77-88%) cabbage cultivars ( $\chi^2_1$  = 0.226, p = 0.635).

## Innate predator selectivity between different aphid host plants

To test whether naïve *C. carnea* select between aphid species while locating aphid host plants, predators were given free choice to visit a plant host to *B. brassicae* and/or a plant host to *M. persicae*. The first plant visited in an assay by *C. carnea* (Table 1a) was not significantly affected by aphid species, by plant cultivar or by the interaction term between the two factors. Additionally, there was no significant effect of aphid species or any other fixed factor on: plant fidelity (Table 1b); the time *C. carnea* spent foraging on host plants (Table 1c) or the occurrence of predation on plants (Table 1d). Of the 28 tests where predators visited an aphid host plant, 10 tests yielded predation. The most number of attacks observed in one test was six, with three kills and three aphids evading predation by dropping from the plant; the least was one attack, with feeding lasting beyond the forty-five minute experimental duration. Of the complete feeding times observed, the duration ranged from two to thirty-one minutes.

## Innate prey selection in response to different prey abundances

C. carnea were exposed to mixed populations of B. brassicae and M. persicae at different relative abundances to test whether their innate prey choice of was determined by prey relative abundance. After 5 hours exposure to C. carnea, the counts of M. persicae were

significantly lower than the counts of *B. brassicae* in the single species treatments ( $\chi^2_1$  = 5.083, p = 0.024). The back-transformed parameter estimates for mean *B. brassicae* count was  $18.6 \pm 0.7$  aphids (a 7% reduction from the initial population size) and the mean *M. persicae* count was  $16.4 \pm 0.9$  aphids (a reduction of approximately 18% from the initial population size). These parameter estimates were used to calculate the expected counts.

The interaction between aphid abundance ratio and aphid species was significant ( $\chi^2_2$  = 6.884, p = 0.032), as M. persicae counts were higher than expected when the aphid abundance ratio was 15:5 in favour of B. brassicae (Fig. 1). However, aphid abundance ratio did not significantly affect the degree of deviation from the expected counts of each aphid species in mixed-species treatments ( $\chi^2_2 = 1.019$ , p = 0.601) and the effect of aphid species on degree of deviation was only marginally significant ( $\chi^2_1 = 3.685$ , p = 0.055).

## Predator associative learning

To test whether *C. carnea* learn to avoid consuming *B. brassicae* through experience, predators were repeatedly exposed to mixed populations of *B. brassicae* and *M. persicae* (at equal proportion). After the first 5 hour exposure of naive *C. carnea* to mixed-species aphid populations, aphid counts were not significantly affected by aphid species, plant cultivar or by the interaction term (Table 3a, Fig. 2a). In all subsequent exposures, when predators had previous experience with the aphid species, *M. persicae* counts were lower than *B. brassicae* counts and the counts of both species declined with progressive exposures (Table 3b, Fig. 2b).

### **DISCUSSION**

The aim of this study was to assess whether a generalist predatory invertebrate (Chrysoperla carnea, lacewing larvae) innately avoids, or learns to avoid consuming harmful, chemically-defended prey (Brevicoryne brassicae) where non-defended prey (Myzus persicae) are also available, and whether any selectivity that predators exhibit varies depending on the variety of plant hosting the prey species. Our experiments found no evidence that naïve C. carnea preferentially select M. persicae aphids when both prey species are available, and even with experience of both prey species, C. carnea did not reject B. brassicae entirely in favour of M. persicae. Continued consumption of B. brassicae by C. carnea may influence how C. carnea affects the wider community, by increasing C. carnea mortality rates (when consumed in isolation), and potentially, by reducing the strength of predation experienced by other prey species.

Our results supported the assumption that *B. brassicae* are more harmful for *C. carnea* to consume than *M. persicae*, as a diet of *B. brassicae* reduced survival of *C. carnea* by approximately 12% compared to a diet of *M. persicae*. This is consistent with other studies showing that *B. brassicae* chemical defences (Bridges et al. 2002, Kazana et al. 2007) can increase mortality, or reduce the growth rates of generalist predatory invertebrates, including *Adalia bipunctata* (Francis et al. 2001; Kazana et al. 2007; Pratt et al. 2008), *Episyrphus balteatus* and *C. carnea* (Chaplin-Kramer et al. 2011, Kos et al. 2011, 2012). Additionally, although the strength of *B. brassicae* chemical defences can vary depending on the host plant's chemical defences (as Chaplin-Kramer et al. 2011, Kos et al. 2011), we found no difference in survival of *C. carnea* fed *B. brassicae* from the two cabbage cultivars used in this study. This lack of host plant effects may be due to host plants affecting biological parameters of the predator we did not measure (such as predator growth rates or fecundity, Chaplin-Kramer et al. 2011, Kos et al. 2011), or due to similar expression of chemical defences by the two cultivars. However, as *C. carnea* survival was unaffected by the cultivar

hosting *B. brassicae*, this may help explain why *C. carnea* behaviour in the subsequent experiments was unaffected by plant cultivar. If there was a difference in mortality between *C. carnea* consuming *B. brassicae* from one cultivar compared to the other, then theoretically, consuming the more harmful *B. brassicae* may provide a stronger stimulus for deterrence or learning than consumption of the less harmful *B. brassicae*. As *C. carnea* survival was only affected by differences in aphid species (chemically-defended or non-defended), only a difference in species, not the additional difference in *B. brassicae* host plant, is likely to provide a strong enough stimulus for discrimination or learning.

Given *B. brassicae* are more harmful to consume than *M. persicae*, it may be expected that *C. carnea* would choose to feed on *M. persicae* over *B. brassicae* where available. However, naïve *C. carnea* larvae did not display any innate preference for *M. persicae* over *B. brassicae* during any stage of foraging: when locating occupied plants; when locating prey within plants and when accepting prey (Table 1) (Hoy and Smilanick 1981, Vinson 1976). Naïve *C. carnea* therefore were not innately deterred by any cues associated with *B. brassicae* (Francis et al. 2005, Mappes et al. 2005). The results suggest that naïve *C. carnea* consume the aphids they encounter first while foraging and support the widely held assumption that any selectivity that generalist predatory invertebrates exhibit is a learned behaviour (Boivin et al. 2010, Dukas 2008).

A lack of selectivity by naïve *C. carnea* was also shown in the third experiment, as *C. carnea* generally predated according to the relative abundance of each aphid species (Fig. 1). Interestingly however, when *B. brassicae* were more abundant than *M. persicae* (15:5 *B. brassicae*: *M. persicae* treatment), there was some evidence that the high consumption of *B. brassicae* negatively impacted on *M. persicae* consumption. The final count of *B. brassicae*, although lower than expected, was not significantly different to the expected count, however, counts of *M. persicae* were significantly higher than expected (Fig. 1). In a similar study,

Eisner et al. (2000) observed that larvae of the green lacewing *Ceraeochrysa cubana* (Hagen) were deterred from feeding on moth eggs if only a few of those present in a cluster were identified as being chemically-defended. Therefore, upon encountering and consuming *B. brassicae* at a high frequency, *C. carnea* may have been deterred from feeding (especially as other glucosinolate-sequestering herbivores can be unpalatable to their predators, Müller et al. 2002, Vlieger et al. 2004), rejecting all available prey and releasing *M. persicae* from predation.

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

Given repeated exposure to harmful and non-harmful prey, it may be expected that predators would learn to select the non-harmful prey species (Boivin et al. 2010, Dukas 2008). After the first exposure, there was an initial change in response to the two aphid species, where M. persicae counts were lower than B. brassicae by  $0.91 \pm 0.04$  aphids in each successive exposure. However, the difference did not become more marked over time. Thus, although there was a change in behaviour, C. carnea did not learn from experience to avoid B. brassicae consumption altogether. Welch & Lundgren (2014) recently assessed the ability of three arthropod predators to learn to avoid chemically-defended western corn rootworm Diabrotica virgifera virgifera LeConte (Coleoptera: Chrysomelidae). Like B. brassicae, D. virgifera does not display aposematic colouration to indicate chemical defences, and harms predators upon attack. The rootworm's haemolymph can be unpalatable or can impede feeding by coagulating around the predator's mouthparts. Interestingly, the predators did not learn to avoid predating on the rootworm, rather Gryllus pennsylvanicus Burmeister (Orthoptera: Gryllidae) crickets attacked rootworm as much as palatable maggots of the house fly Musca domestica Linnaeus (Diptera: Muscidae), stone centipedes (Chilopoda: Lithobiidae) showed little impediment resulting from the rootworms defences and Lasius neoniger Emery (Hymenoptera: Formicidae) ants learned to attack successfully in groups, rather than individually (Welch & Lundgren 2014). Here, the underlying cause of the

behavioural change requires further investigation, yet like the predators of the rootworm, it may be possible that *C. carnea* bypassed or tolerated *B. brassicae* chemical defences. By consuming more *M. persicae* than *B. brassicae*, *C. carnea* may have maintained an optimal, mixed diet; by which the concentration of ingested defence chemicals from *B. brassicae* was diluted by feeding on *M. persicae*, yielding a high nutritional payoff and facilitating the increase in aphid consumption as the predator grows (as Fig. 1; Cruz-Rivera and Hay 2003; Glendinning 2007).

The results suggest that prey chemical defences do not influence the prey choice of naïve *C. carnea*. Rather, naïve *C. carnea* locate plants hosting prey at random and feed on aphids they encounter first. If multiple species are encountered (for example, if they share a host plant), then prey choice may be determined by the relative abundance of each prey species (Fig. 2). However, prey choice may be affected *in situ* by how prey use a shared host plant and by how the predator uses the plant to forage (Schmitz et al. 2004). For example, in the plant selection assays, *C. carnea* used the stem to access the lower leaves and consumed aphids they encountered first. If one aphid species colonises the stem or lower leaves more than the other, then predators are likely to encounter and consume that species first at higher frequency (*M. persicae* reportedly use the lower leaves of *Brassica* plants and *B. brassicae* use leaves higher up the stem, Trumble 1980). Furthermore, if parts of the plant are inaccessible and one prey species uses them as a refuge, this may affect prey encounter and consumption rates as well (Eigenbrode et al. 1999, Fordyce and Agrawal 2001, Northfield et al. 2012).

The lack of naïve preference between toxic and non-toxic prey, at least in the short term, could lead to wider associational/apparent interactions. Associational interactions, typically, involve chemically-defended species that reduce predation of non-defended species (associational resistance) or are consumed incidentally due to the presence of palatable

species (associational susceptibility) (Wahl and Hay 1995; Barbosa et al. 2009). These indirect interactions have mostly been observed between plant species (Barbosa et al. 2009, Kostenko et al. 2012, Castagneyrol et al. 2013) and in aquatic systems (Hay 1986, Wahl and Hay 1995), yet, although examples of 'associational' interactions in terrestrial systems are sparse (Barbosa et al. 2009), indirect 'apparent' interactions have been widely reported in terrestrial invertebrate communities (van Veen et al. 2006, Chailleux et al. 2014). In this body of literature, apparent commensalism, where one species benefits from the presence of another, through indirect interactions with a natural enemy, is analogous to associational resistance (van Veen et al. 2005, 2009) and apparent amensalism, where one species suffers from the presence of another, through indirect interactions with a natural enemy, is analogous to associational susceptibility (Chaneton & Bonsall 2000). The mechanisms that determine associational resistance or susceptibility and the effects of these apparent interactions in applied agricultural systems however remain unclear (Barbosa et al. 2009, Chailleux et al. 2014). Here we report that continued consumption of B. brassicae may increase C. carnea mortality and potentially reduce predator numbers – a density mediated indirect interaction that could benefit M. persicae. In the short term, the effects of B. brassicae chemical defences may potentially reduce predation of other herbivores (Fig. 1) – a trait-mediated indirect interaction from which M. persicae benefit.

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

The occurrence of associational/apparent interactions may further change if *C. carnea* can learn from cues not measured in this investigation. Our experiments did not account for the ability of *C. carnea* to learn olfactory cues from plants host to different prey species (Glinwood et al. 2011), or their ability to learn to select between prey that are at different densities. Further studies could usefully explore these aspects of learning, along with the duration over which any discriminatory behaviours can be retained (Boivin et al. 2008; Glinwood et al. 2011). If predators fail to discriminate between toxic and non-toxic prey or if

451 selectivity is forgotten, associational/apparent interactions may be prevalent and furthermore, 452 may affect the predator's impacts on prey communities. 453 **ACKNOWLEDGEMENTS** 454 This work was supported by the Biotechnology and Biological Sciences Research Council 455 (BBSRC), by studentship funding to C.M. Nesbit. Thanks to Dr. Annabel Rice and Dr. Mark 456 Mainwaring for advice with statistics and manuscript preparation. The authors wish to thank 457 two anonymous referees for their helpful comments in improving the manuscript. 458 459 460 **REFERENCES** 461 Barbosa P, Hines J, Kaplan I, Martinson H, Szczepaniec A, Szendrei Z. 2009. Associational 462 Resistance and Associational Susceptibility: Having Right or Wrong Neighbors. Annu. Rev. 463 Entomol., 40, 1-20. 464 465 Bates B, Maechler M, Bolker B. 2012. lme4: Linear mixed-effects models using S4 classes. 466 R package version 0.999999-0. http://CRAN.R-project.org/package=lme4 467 468 Boivin G, Roger C, Coderre D, Wajnberg E. 2008. Learning affects prey selection in larvae 469 470 of a generalist coccinellid predator. Entomol. Exp. Appl., 135, 48-55.

- 472 Bridges M, Jones AME, Bones AM, Hodgson C, Cole R, Bartlet E, Wallsgrove R, Karapapa
- VK, Watts N, Rossiter J.T. 2002. Spatial organization of the glucosinolate-myrosinase system
- in brassica specialist aphids is similar to that of host plants. *Proc. R. Soc. B.*, 269, 187-191.

- Castagneyrol B, Giffard B, Péré C, Jactel H. 2013. Plant apparency, an overlooked driver of
- associational resistance to insect herbivory. *J. Ecol.*, 101, 418-429.

478

- Chailleux A, Mohl EK, Alves MT, Messelink GJ, Desneux N. 2014. Natural enemy-mediated
- 480 indirect interactions among prey species: potential for enhancing biocontrol services in
- 481 agroecosystems. *Pest Manag. Sci.*, 70, 1769-1779.

482

- Chaplin-Kramer R, Kliebenstein DJ, Chiem A, Morrill E, Mills NJ, Kremen C. 2011.
- 484 Chemically mediated tritrophic interactions: opposing effects of glucosinolates on a specialist
- herbivore and its predators. J. Appl. Ecol., 48, 880-887.

486

- Charnov EL. 1976. Optimal foraging, the Marginal Value Theorem. Theor. Popul. Biol., 9,
- 488 129-136.

489

490 Crawley M. 2007. The R Book. John Wiley & Sons, Ltd, England.

- 492 Cruz-Rivera E, Hay ME. 2003. Prey Nutritional Quality Interacts with Chemical Defenses to
- 493 Affect Consumer Feeding and Fitness. *Ecol. Monogr.*, 73, 483-506.

- Dejean A, Gibernau M, Lauga J, Orivel J. 2003. Coccinellid Learning During Capture of
- 496 Alternative Prey. J. Insect Behav., 16, 859-864.

497

- Dicke M, van Loon JJA. 2000. Multitrophic effects of herbivore-induced plant volatiles in an
- 499 evolutionary context. *Entomol. Exp. Appl.*, 97, 237-249.

500

501 Dukas R. 2008. Evolutionary Biology of Insect Learning. *Annu. Rev. Entomol.*, 53, 145-160.

502

- 503 Eigenbrode SD, Kabalo NN, Stoner KA. 1999. Predation, behaviour, and attachment by
- 504 Chrysoperla plorabunda larvae on Brassica oleracea with different surface waxblooms.
- 505 Entomol. Exp. Appl., 90, 225-235.

506

- 507 Eisner T, Eisner M, Rossini C, Iyengar VK, Roach BL, Benedikt E, Meinwald J. 2000.
- 508 Chemical defences against predation in an insect egg. P. Natl. Acad. Sci. USA, 97, 1634-
- 509 1639.

- 511 Ellis PR, Singh R, Pink DAC, Lynn JR, Saw PL. 1996. Resistance to Brevicoryne brassicae
- in horticultural brassicas. *Euphytica*, 88, 85-96.

Forbes LS. 1989. Prey Defences and Predator Handling Behaviour: The Dangerous Prey

515 Hypothesis. *Oikos*, 55, 155-158.

516

518

517 Fordyce JA, Agrawal AA. 2001. The role of plant trichomes and caterpillar group size on

growth and defence of the pipevine swallowtail Battus philenor. J. Anim. Ecol., 70, 997-

519 1005.

520

521 Francis F, Lognay G, Wathelet J-P, Haubruge E. 2001. Effects of allelochemicals from first

(Brassicaceae) and second (Myzus persicae and Brevicoryne brassicae) trophic levels on

523 *Adalia bipunctata. J. Chem. Ecol.*, 27, 243-256.

524

522

525 Francis F, Lognay G, Haubruge E. 2004. Olfactory responses to aphid and host plant volatile

releases: (E)- $\beta$ -farnesene an effective kairomone for the predator Adalia bipunctata. J. Chem.

527 Ecol., 30, 741-755.

528

526

529 Francis F, Martin T, Lognay G, Haubruge E. 2005. Role of (E)-β-farnesene in systematic

aphid prey location by Episyrphus balteatus larvae (Diptera: Syrphidae). Eur. J. Entomol.,

531 102, 431-436.

532

- Glendinning JI. 2007. How Do Predators Cope With Chemically Defended Foods? *The Biol.*
- 534 Bull., 213, 252-266.

- 536 Glinwood R, Ahmed E, Qvarfordt E, Ninkovic V. 2011. Olfactory learning of plant
- genotypes by a polyphagous insect predator. *Oecologia*, 166, 637-647.

538

Griffiths D. 1980. Foraging costs and relative prey size. *Am. Nat.*, 116, 743-752.

540

- 541 Hariprasad KV, van Emden HF. 2010. Mechanisms of partial plant resistance to
- diamondback moth (*Plutella xylostella*) in brassicas. *Int. J. Pest Manage.*, 56, 15-22.

543

- Hatano E, Kunert G, Michaud JP, Weisser WW. 2008. Chemical cues mediating aphid
- location by natural enemies. Eur. J. Entomol., 105, 97-806.

546

- Hay ME. 1986. Associational Plant Defenses and the Maintenance of Species Diversity:
- Turning Competitors into Accomplices. *Am. Nat.*, 128, 617-641.

549

- Hayward MW, Kerley GIH. 2005. Prey preferences of the lion (Panthera leo). Zool. Soc.
- 551 *Lond.*, 267, 309-322.

Hoy MA, Smilanick JM. 1981. Non-random prey location by the phytoseiid predator

554 Metaseiulus occidentalis: Differential responses to several spider mite species. Entomol. Exp.

555 Appl., 29, 241-253.

556

Kalule T, Wright DJ. 2002a. Effect of cabbage cultivars with varying levels of resistance to

aphids on the performance of the parasitoid, *Aphidius colemanii* (Hymenoptera: Braconidae).

559 Bull. Entomol. Res., 92, 53-59.

560

561

Kalule T, Wright DJ. 2002b. Tritrophic interactions between cabbage cultivars with different

resistance and fertilizer levels, cruciferous aphids and parasitoids under field conditions. *Bull.* 

563 Entomol. Res., 92, 61-69.

564

Kazana E, Pope TW, Tibbles L, Bridges M, Pickett JA, Bones AM, Powell G, Rossiter JT.

2007. The cabbage aphid: a walking mustard oil bomb. *Proc. Biol. Sci.*, 274, 2271-2277.

567

568

566

Kos M, Kabouw P, Noordam R, Hendriks K, Vet LEM, van Loon JJA, Dicke M. 2011. Prey-

mediated effects of glucosinolates on aphid predators. *Ecol. Entomol.*, 36, 377-388.

570

572

573

Kos M, Houshyani B, Achhami BB, Wietsma R, Gols R, Weldegergis BT, Kabouw P,

Bouwmeester HJ, Vet LEM, Dicke M, van Loon JJA. 2012. Herbivore-Mediated Effects of

Glucosinolates on Different Natural Enemies of a Specialist Aphid. J. Chem. Ecol., 38, 100-

574 115.

mediating the strength of predator-prey interactions (Doctoral Dissertation). Lancaster,

593

594

Lancashire: Lancaster University.

- Nishida R. 2002. Sequestration of Defensive Substances from Plants by Lepidoptera. *Annu*.
- 597 Rev. Entomol., 47, 57-92.

- Northfield TD, Snyder WE, Snyder GB, Eigenbrode SD. 2012. A simple plant mutation abets
- a predator-diversity cascade. *Ecology*, 93, 411-420.

601

- Opitz SEW, Müller C. 2009. Plant chemistry and insect sequestration. Chemoecology, 19,
- 603 117-154.

604

- Pratt C, Pope TW, Powell G, Rossiter JT. 2008. Accumulation of Glucosinolates by the
- 606 Cabbage Aphid Brevicoryne brassicae as a Defense Against Two Coccinellid Species. J.
- 607 Chem. Ecol., 34, 323-329.

608

- R Core Development Team. 2012. R: A language and environment for statistical computing.
- R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL
- 611 http://www.R-project.org/.

612

- 613 Schmitz OJ. 2003. Top predator control of plant biodiversity and productivity in an old-field
- 614 ecosystem. *Ecol. Lett.*, 6, 156-163.

- Schmitz OJ. 2006. Predators have large effects on ecosystem properties by changing plant
- diversity, not plant biomass. *Ecology*, 87, 1432-1437.

- 619 Schmitz OJ, Hambäck PA, Beckerman AP. 2000. Trophic Cascades in Terrestrial Systems: A
- Review of the Effects of Carnivore Removals on Plants. Am. Nat., 155, 141-153.

621

- 622 Schmitz OJ, Krivan V, Ovadia O. 2004. Trophic cascades: the primacy of trait-mediated
- indirect interactions. Ecol. Lett., 7, 153-163.

624

- Schuler TH, van Emden HF. 2000. Resistant cultivars change the susceptibility of *Plutella*
- 626 *xylostella* to *Bacillus thuringiensis*. *Agric. For. Entomol.*, 2, 33-38.

627

- 628 Snyder WE, Wise DH. 2001. Contrasting trophic cascades generated by a community of
- generalist predators. *Ecology*, 82, 1571-1583.

630

- 631 Snyder GB, Finke DL, Snyder WE. 2008. Predator biodiversity strengthens aphid
- suppression across single- and multiple-species prey communities. *Biol. Control*, 44, 52-60.

633

- 634 Snyder WE, Snyder GB, Finke DL, Straub CS. 2006. Predator biodiversity strengthens
- herbivore suppression. *Ecol. Lett.*, 9, 789-796.

- Staley JT, Stewart-Jones A, Poppy GM, Leather SR, Wright DJ. 2009. Fertilizer affects the
- behaviour and performance of *Plutella xylostella* on brassicas. *Agric. For. Entomol.*, 11, 275-
- 639 282.

- Staley JT, Stewart-Jones A, Pope TW, Wright DJ, Leather SR, Hadley P, Rossiter JT, van
- 642 Emden HF, Poppy GM. 2010. Varying responses of insect herbivores to altered plant
- chemistry under organic and conventional treatments. *Proc. Biol. Sci.*, 277, 779-786.

644

- 645 Symondson WOC, Sunderland KD, Greenstone MH. 2002. Can Generalist Predators be
- 646 Effective Biocontrol Agents? Annu. Rev. Entomol., 47, 561-594.

647

- Vandermoten S, Mescher MC, Francis F, Haubruge E, Verheggen FJ. 2012. Aphid alarm
- 649 pheromone: An overview of current knowledge on biosynthesis and functions. *Insect*
- 650 Biochem. Mol. Biol., 42, 155-163.

651

- van Veen FJF, van Holland PD, Godfray HCJ. 2005. Stable coexistence in insect
- communities due to density- and trait-mediated indirect interactions. *Ecology*, 86, 1382-1389.

- van Veen FJF, Morris RJ, Godfray HCJ. 2006. Apparent Competition, Quantitative Food
- 656 Webs, and the Structure of Phytophagous Insect Communities. Annual Review of
- 657 Entomology, 51, 187-208.

van Veen FJF, Brandon CE, Godfray HCJ. 2009. A positive trait-mediated indirect effect

involving the natural enemies of competing herbivores. *Oecologia*, 160, 195-205.

661

662

663

Verkirk RHJ, Neugebauer KR, Ellis PR, Wright DJ. 1998. Aphids on cabbage: tritrophic and

selective insecticide interactions. Bull. Entomol. Res., 88, 343-349.

664

665

Vinson SB. 1976. Host Selection by Insect Parasitoids. Annu. Rev. Entomol., 21, 109-133.

666

Vlieger L, Brakefield PM, Müller C. 2004. Effectiveness of the defence mechanism of the

668 turnip sawfly, Athalia rosae (Hymenoptera: Tenthredinidae), against predation by lizards.

669 Bull. Entomol. Res., 94, 283-289.

670

Wahl M, Hay ME. 1995. Associational resistance and shared doom: effects of epibiosis on

672 herbivory. *Oecologia*, 102, 329-340.

673

Wilby A, Anglin LA, Nesbit CM. 2013. Plant species composition alters the sign and strength

of an emergent multi-predator effect by modifying predator foraging behaviour. PLOSone.

676 8(8): e70258. doi:10.1371/journal.pone.0070258.

677

678	Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM. 2009. Mixed Effects Models and
679	Extensions in Ecology with R. Springer Science.
680	
681	Zehnder G, Gurr GM, Kühne S, Wade MR, Wratten SD, Wyss E. 2007. Arthropod Pest
682	Management in Organic Crops. Annu. Rev. Entomol., 52, 57-80.

## FIGURE LEGENDS

Figure 1: The expected and observed counts of *Brevicoryne brassicae* and *Myzus persicae* aphids alive after 5 hours exposure to predatory *Chrysoperla carnea* larvae, when presented at different aphid abundance ratios (number of *B. brassicae*: number of *M. persicae*). Error bars denote the standard errors of the means.

Figure 2: Counts of *Brevicoryne brassicae* and *Myzus persicae* aphids alive after 5 hours exposure to *Chrysoperla carnea* larvae, that were naïve (left of the dashed line at 0 previous exposures), or given previous exposure to their prey (right of the dashed line; 1-4 previous exposures). The starting population was 10 aphids of each species. Error bars denote the standard errors of the means.

### TABLES AND TABLE LEGENDS

Table 1: Results of deletion tests for GLMMs without a fixed effect, for different response variables associated with predator behaviour in aphid host plant choice tests: (a) of the plants visited; the number of plants visited first by predators (n = 37 plants); (b) of the first visited plants; the number of exclusive visits (as opposed to predators also visiting the other respective aphid host plant) (n = 28 plants); (c) the time predators spent foraging on the host plants (minutes) and (d) the number of plants where predation occurred (n = 37 plants). Fixed effects included aphid species (*Brevicoryne brassicae* or *Myzus persicae*), plant cultivar (Minicole or Derby Day). All fixed effects had one degree of freedom.

	Response variables							
	(2	1)	(b)		(c)		(d)	
	Plants visited		Plant fidelity		Foraging		Plants	
	first				Time (mins)		yielding	
							preda	ation
Fixed Effect	$\chi^2$	p	$\chi^2$	p	$\chi^2$	p	$\chi^2$	p
Aphid								
Species	1.549	0.213	0.681	0.409	1.807	0.179	2.341	0.126
Plant								
Cultivar	0.003	0.954	0.096	0.757	0.874	0.350	0.055	0.814
Species:								
Cultivar	0.478	0.489	1.449	0.229	0.151	0.697	0.554	0.457

Table 2: The significance of fixed effects on the number of aphids alive after 5 hours exposure to *Chrysoperla carnea* larvae, that were (a) naïve or (b) experienced of their prey. Fixed effects included aphid species (*Brevicoryne brassicae* or *Myzus persicae*), plant cultivar (Minicole or Derby Day) and the number of previous exposures predators had received to the aphids (1-4 previous exposures). All factors had one degree of freedom.

7	1	$^{\circ}$
1		$\mathcal{L}$
•	-	_

Predators:	(a) Naïve		(b) Experienced			
Fixed Effects	$\chi^2$	p	Fixed Effects	$\chi^2$	p	
Aphid Species	0.288	0.592	Aphid Species	4.285	0.038	
Plant Cultivar	0.000	1.000	Exposures	14.673	< 0.001	
Aphid:Plant	0.000	1.000	Plant Cultivar	0.002	0.963	
			Aphid:Exposures	0.016	0.897	
			Aphid:Plant	0.121	0.728	
			Exposures:Plant	0.071	0.790	
			Aphid:Exposure:Plant	0.571	0.450	

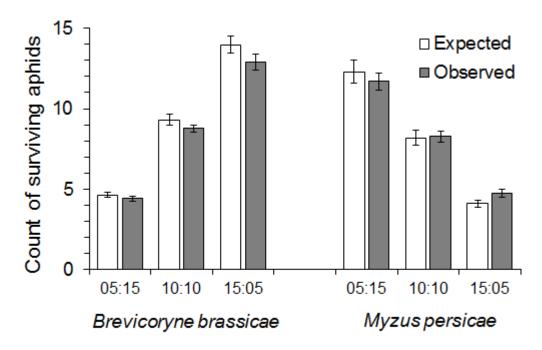
# 714 Figures

715

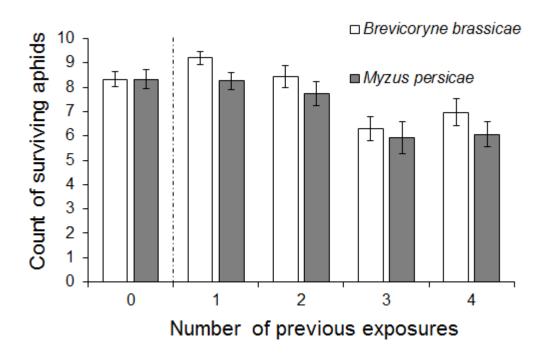
716

717

Figure 1.



Aphid abundance treatment (No. *B. brassicae* : No. *M. persicae*)



719 Figure 2.