- 1 Alpine meadow could be better restored before moderately degradation using
- 2 inorganic nitrogen on the Tibetan Plateau
- Jianbo Sun^{12*}, Chimin Lai^{3*}, Fei Peng¹⁴, Junzhou⁵, Mengting Hu⁶, Xingzhi Xu¹², Xiaowei
- 4 Gou⁷, Huakun Zhou⁸, Carly Stevens⁹
- ⁵ Key Laboratory of Ecological Safety and Sustainable Development in Arid Lands, Northwest Institute of
- 6 Eco-Environment and Resources, Chinese Academy of Sciences, Lanzhou, China
- 7 ² University of Chinese Academy of Sciences
- 8 ³ School of Geography and Environment, Jiangxi Normal University, Nanchang, China
- 9 ⁴ Beiluhe Observation and Research Station on Frozen Soil Engineering and Environment in Qinghai-
- 10 Tibet Plateau
- 11 ⁵Key Laboratory of Mountain Surface Processes and Ecological Regulation, Institute of Mountain Hazards
- 12 and Environment, Chinese Academy of Sciences, Chengdu, China
- 13 ⁶ Anhui Normal University, Wuhu, China, 241002
- ⁷ Wildfire Research Center, National Institute of Natural Hazards, Beijing 100085, China
- 15 8 Institute of Tibetan Biology Research, Chinese Academy of Sciences, Xining, China
- 16 ⁹ Lancaster Environment Centre, Lancaster University, Lancaster, UK
- 17 *Equally contribute to the paper
- 18 Correspondence to Fei Peng, pengfei@lzb.ac.cn

19 Abstract

- 20 Nitrogen (N) fertilization is considered as a powerful alternative approach to restore
- 21 degraded alpine meadows. However, whether form of available N matters and their
- 22 efficiency along a degradation gradient remains largely unexplored. A four-year N addition
- experiment with different available N forms (NH₄⁺-N, NO₃⁻-N and Glycine) was carried out
- on the undegraded, moderately (MD) and severely degraded (SD) alpine meadows on the
- 25 Tibetan Plateau. Plant aboveground productivity was greatly increased in undegraded and
- 26 MD alpine meadows but belowground productivity was enhanced in SD alpine meadow,
- especially in the two inorganic N treatments. When inorganic N was added, plants

accumulated more N compared to the control in undegraded alpine meadow. Plants accumulated more N in the MD alpine meadow than in undegraded alpine meadow in all forms of N addition treatments as the plant N uptake rate was almost doubled in MD alpine meadow when N was added. In SD alpine meadow, most of the added N moved to the microbial N pool with the largest increase observed in the NO₃-N treatment. Leaching remained almost unchanged in NH₄+N and Glycine in undegraded and MD alpine meadows but increased in SD alpine meadow, especially in NO₃-N treatment. However, leaching was low, accounting for less than 5% of the added N even in the SD alpine meadow. Our results indicate 1) the added N were mostly up taken and retained in plants or microorganisms regardless of N forms, 2) inorganic N is more efficient in increasing plant productivity for MD but not for SD alpine meadow, 3) fertilization using inorganic N to restore the degraded alpine meadow is efficient for MD but not for SD alpine meadows. **Keywords:** Nitrogen addition, degradation, nitrogen partitioning, alpine meadow, Tibetan

Introduction

plateau

Alpine meadows on the Qinghai-Tibetan Plateau (QTP), are of great importance for biodiversity conservation, soil and water conservation, carbon sequestration and supporting the livelihood of local people (Liu et al., 2022; Wang et al., 2022) but 70 % of it is severely degraded (Bardgett et al., 2021). Although there have been restoration efforts (Dong et al., 2024), sustainable restoration still requires further understanding of ecological processes and mechanisms involving alpine meadows' degradation (Bardgett et al., 2021). In nitrogen (N) limited ecosystems such as these (Du et al., 2020), manipulative N addition

experiments have revealed an increase in plant biomass, litter decomposition rate, and soil microbial activity, thereby increasing soil organic carbon (SOC) and possibly being conducive to restoring the degraded ecosystem (Liu et al., 2019; Tang et al., 2023; Tian et al., 2019). Although N from atmospheric deposition has increased twofold in last two decades on the QTP (Chen et al., 2023), it is still much lower than the plant N requirements (Fu & Shen, 2016). Thus, N fertilizer application is practiced in many restoration actions on the QTP (Gu et al., 2023).

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

The form in which the N is applied (ammonia nitrogen: NH₄⁺-N, nitrate nitrogen: NO₃⁻ -N, and smaller molecular organic nitrogen like urea and amino acids) matters for two reasons: firstly, plant preferences for specific available N forms may determine the efficiency of them for plant growth performance in a short term (Xu et al., 2012); secondly, differences in mobility and associations with soil minerals differ between N forms, together with stoichiometry of plants, soils and microorganisms affecting the capability of them to retain the added N in the system and consequently long-term potential for restoration (Yang et al., 2022; Yang et al., 2021). All added N will be fiercely competed for by plants and microorganisms and is likely to be fully retained in the ecosystem regardless of the form in a N limited ecosystem (Choudhary et al., 2016). Plants can assimilate both inorganic N and smaller molecular compounds of organic N but they generally prefer inorganic N (Ashton et al., 2010; Harrison et al., 2007; Liu et al., 2017). Recent studies have indicated that plants in undegraded alpine meadows on the QTP mainly assimilate NH₄+-N most effectively ((Lai et al., 2024; Lai et al., 2022). Any form of added N may first enter into the microbial N pool due to their competitive advantage resulting from their large volume to

surface area ratio (Zhang et al., 2024) but more NH₄+-N will be immobilized in plant organs or microbes because of its lower energy requirement for being synthesized into protein in contrast to NO₃--N (L'Espérance et al., 2024). As they assimilate additional N, soil microorganisms would need to decompose more soil organic matter in order to maintain a stoichiometric homeostasis (Li et al., 2021; Zhu et al., 2021), thereby increasing the net mineralization rate and providing N for plants. In undegraded alpine meadows, NH₄+-N is the most abundant form of available N in the soil. Even when NO₃--N or organic forms or N such as glycine are added, NH₄+-N is still the most abundant available form (Lai et al., 2024; Lai et al., 2022).

The fate of different forms of additional N, whether it is transformed into NO₃-N, quickly up taken by plants, lost by leaching, or immobilization or mobilization by microbes, depends on the soil environment. With degradation of alpine meadows, changes in the soil environment such as an increase in soil pH, improvement in aeration, and reduction in soil moisture (Peng et al., 2016, 2018) favoring different microbial processes. The added NH₄⁺-N and smaller molecular N may be transferred into oxidized forms like NO₃-N and N₂O (Che et al., 2019; Liu et al., 2016). In conditions that result in more NO₃-N in soil, the high mobility of NO₃- anion may lead to a higher plant N uptake rates and thus retention of a large proportion of added N in plants (K. E. Chen et al., 2020); or on the other hand, less N may be retained in the ecosystem because of the high inclination of NO₃- anion to be leached (Di & Cameron, 2002). Although the soil conditions in degraded alpine meadows facilitate nitrification and make NO₃-N the most abundant available N in the soil, degradation also promotes the loss of N via leaching due to a lack of clay particles to

adsorb NO₃-N. This means that degraded soils are even more N limited. In severely N limited ecosystems such as arctic tundra, soil microbes retain 39 and 70% of the additional N in mineral and organic layers, acting as an effective buffer for leaching (Choudhary et al., 2016). How the forms of N that are added impacts partitioning among active N pools in plants, soil and microorganisms along degradation gradients remain unexplored. We conducted a four-year field experiment with NH₄⁺-N, NO₃⁻-N and Glycine addition in undegraded (IN), moderately (MD) and severely degraded (SD) alpine meadows in the inland of QTP to investigate the fate of N after addition, and the efficiency of different N forms in restoring the degraded alpine meadow. We hypothesized that 1) NH₄+N addition will result in the highest plant N uptake rate, the largest increase in aboveground net primary productivity (ANPP), and the greatest retention of added N within the plant N pool compared to NO₃--N or glycine in undegraded meadows. 2) Microbial biomass nitrogen (MBN) will increase disproportionately relative to plant N uptake in degraded (meadows, especially for NO₃-N and glycine additions, leading to a higher microbial C:N ratio. 3) Inorganic N fertilization (particularly NH₄⁺-N) will be most effective at increasing overall ecosystem N retention and plant productivity (ANPP & BNPP) in moderately degraded (MD) meadows, but will have diminished efficacy in SD meadows where microbial immobilization dominate N fate.

Materials and methods

Site description

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

Alpine meadows which were undegraded, moderately degraded (MD) and severely degraded (SD) were set up near Beiluhe Permafrost Engineering and Environment

Comprehensive Observation and Research Station (34° 49' N, 92° 55' E) in a permafrost region in the hinterland of the QTP (Fig. S1). The meadows were classified according to (Xue et al., 2009). In the classification framework, plant coverage, plant community composition, and the aeolian activities are considered. The study site is in an alluvial and diluvial high plain with relatively flat terrain. The altitude in the study site is 4620–4640 m. Annual average temperature and precipitation was -3.8 °C and 290 mm (2009–2024). More than 90% of the precipitation occurs in May-September when plants grow, and the freezing period is from October to April. The permafrost in the study area is 30–70 m deep, and the depth of active layer is 1.5–3.5 m (Li et al., 2021a). The plant community of the undegraded alpine meadow in the study area is dominated by sedges, namely *Kobresia humilis* and *Kobresia pygmaea*. With degradation, forb species like *Leontopodium nanum* and *Saussurea pulchra* typically increase (Li et al., 2024).

N fertilization experiment

An experimental area covering the undegraded, MD and SD meadows (100 × 100 m) was set up near the research station. Twelve 2 × 2 m plots were randomly established in each alpine meadow, and the spacing between each pair of plots was about 5 m.

Starting in 2019 N fertilizer was added as NH₄Cl, KNO₃ and Glycine with an equivalent N amount of 3.1 g m⁻² yr⁻¹ in July each year, 20 times the average N deposition in the eastern QTP (Liu et al., 2015). All N forms were first dissolved in water and sprayed manually. The unfertilized control plot received the same amount of water. All the N addition and control treatments had three replicates in each meadow.

Plant community, productivity and C, N concentration

A 30 × 30 cm quadrat was set in each plot to study plant community characteristics. All vegetation was clipped at ground level to determine the above-ground net primary productivity (ANPP) in mid-August 2020, 2021, and 2023. The below-ground net primary productivity (BNPP) was determined using the in-growth method. In September 2019, a PVC collar with 13 cm in diameter and 30 cm in depth was inserted into the center of each plot. The PVC was refilled with local soil after removing root debris with 2 mm sieve. In September 2020, 2021, and 2023, roots in each PCV were collected and washed. The oven-dried roots was considered BNPP (Lai et al., 2021). Following collection, above-ground and root biomass were dried, ground, and analyzed for C and N. The foliar and root C and N concentrations were measured with an elemental analyzer (Elementar Vario EL, Hanau, Germany).

Soil sampling and measurement of N concentration

Soil samples were taken in each plot using the five-point sampling method. Soil cores (5 cm in diameter) were collected at 0–10, 10–20, 20–30 cm depths, and sub-samples of the same depth were mixed to form a composite sample. After the visible roots and other debris were picked out, each composite soil sample was sifted through a 2 mm sieve and then divided into two sub-samples. One was stored in a refrigerator at 4 $^{\circ}$ C for the measurement of NH₄+-N, NO₃-N, soil microbial biomass carbon (MBC) and nitrogen (MBN) and water content, and the other part is naturally air-dried for the determination of soil pH, SOC and total nitrogen (TN).

Soil MBC and MBN were determined using the chloroform fumigation-potassium sulfate extraction method (Brookes et al., 1985). Soil pH was tested using a acidimeter

(FE28K, Mettler Toledo, Shanghai). The NH₄+-N, NO₃--N in soil were determined by flow autoanalyzer (SEAL Analytical AutoAnalyzer 3, Northern Ireland, UK). Soil inorganic N was the sum of NH₄+-N and NO₃--N.

Nitrogen partitioning among soil, plant and soil microorganisms

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

A randomized block design was adopted, with three identical plots for each treatment. All measurements were conducted during two major field surveys: the initial sampling in June 2022 and the final sampling in September 2023. This period covered the important growing season while taking into account practical limitations such as the impact of the COVID-19 pandemic and worker availability. Plant aboveground parts were collected at the peak of plant growth (mid-August). ANPP and BNPP were multiplied by the N concentrations of leaves and roots respectively, and then added to obtain the plant N pool. By comparing the increase in the plant N pool between the fertilized plots and the unfertilized plots during the two sampling dates, the amount of additional N accumulated by the plants due to N addition measures was determined. The determination of soil and microbial N content involved collecting soil cores up to 30 centimeters deep, which were divided into three depth layers (0 - 10 cm, 10 - 20 cm, 20 -30 cm). For each layer, we calculated the N storage, taking into account the soil bulk density. Three types of soil N were measured: MBN, inorganic nitrogen, and DON. The calculation of the additional N stored in the microorganisms and the N retained in the soil was the same as that for plant N: by comparing the changes in these N reserves between

the fertilized plots and the control plots during the period from June 2022 to September

2023.

From June 2022 to September 2023, in situ anion-cation exchange resin bags were buried in PVC pipes with a diameter of 5 centimeters, placed at depths of 10 centimeters and 20 centimeters respectively. To assess the N loss due to leaching. These resins were extracted with 1 M KCl solution and the concentrations of NH₄⁺-N and NO₃⁻-N in the resins were detected by a flow automatic analyzer. The concentration of inorganic N (the sum of NH₄⁺-N and NO₃⁻-N concentrations, mg kg⁻¹) at each depth (10 cm and 20 cm) was divided by the area of the PVC tube. The inorganic N concentrations were then converted into unit area N leaching (Karhu et al., 2021).

¹⁵N labeling experiments were conducted to explain the N partitioning between plant and soil microorganisms. The ¹⁵N labeling experiment was conducted in August 2021. We used three types of ¹⁵N-labeled chemicals: ¹⁵NH₄+ (98 atom% ¹⁵N), ¹⁵NO₃- (99 atom% ¹⁵N), and ¹⁵N-Glycine (98atom% ¹⁵N). The specific labeling methods, sampling and calculations refer to our previous article (Lai et al., (Lai et al., 2024; Lai et al., 2022).

Statistical analysis

Analyses of observational and experimental data were performed using SPSS 27.0 (SPSS, Chicago, IL, USA) software. Separate one-way analyses of variance (ANOVA) were conducted for each degradation level (undegraded/IN, moderately degraded/MD, severely degraded/SD) to assess the impact of nitrogen addition treatment (Control, NH_4^+ - N, NO_3^- -N, Glycine) on individual response variables (e.g., pH, water content, ANPP, BNPP, plant C and N concentration, soil available N, N absorption rates, N pools, leaching). Where the ANOVA indicated a significant treatment effect (* p^* < 0.05), Tukey's Honestly

Significant Difference (HSD) post-hoc test was used to identify significant differences between individual treatment means within that degradation level. Homogeneity of variances was verified using Levene's test and normality of residuals was assessed using the Shapiro-Wilk test; data met ANOVA assumptions. Structural equation modeling (SEM) was employed to investigate the specific mechanisms and pathways through which N addition affects ecosystem productivity. The model was developed based on established ecological theory and estimated using the lavaan package in R 4.5.1. N addition was specified as an exogenous variable directly affecting key soil properties (including soil pH, soil moisture, soil available N, and soil C/N) and plant C/N, which were hypothesized to subsequently influence ANPP and BNPP(Sun et al., 2020; Zhang et al., 2026). After N addition (especially NH_4^+), the nitrification process ($NH_4^+ > NO_3^-$) releases H^+ ions, which is the main cause of soil acidification (Wang et al., 2023). The effect of N addition on soil moisture was based on the fact that N addition could promote plant growth (especially aboveground parts), thereby increasing transpiration water loss, which may lead to a decrease in soil moisture(Liang et al., 2020). The effect of N addition on soil C/N was based on the ecological processes that exogenous N input dilutes the relative concentration of soil organic matter, usually resulting in a decrease in the soil C/N (Li et al., 2023; Wang et al., 2020; Xu et al., 2022). When N is abundant, microorganisms accelerate the decomposition of organic matter (priming effect), which may also release CO2 and indirectly affect the C/N(Qin et al., 2024; Widdig et al., 2020). Correlations among mediating variables were permitted in the model specification. All figures present the data as mean \pm standard error (SE). The significance level was set at p = 0.05.

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

Results

Changes in ANPP and BNPP after the N addition

In the control treatment, NPP (the sum of ANPP and BNPP) decreased with the severity of alpine meadow degradation. The responses of ANPP and BNPP varied annually, with degradation severity and different forms of added reactive N (Fig.1). In 2020 and 2021, ANPP was significantly increased by all N additions, with a relatively higher increase under inorganic N addition both in undegraded and MD alpine meadows (Figs.1a and b). In 2020, the addition of NH₄⁺-N and NO₃⁻-N increased ANPP by 29.6% and 37.3% in undegraded, and by 57.3% and 61.5% in MD alpine meadow, respectively. In 2021, the addition of NH₄⁺-N and NO₃⁻-N increased ANPP by 34.6% and 8.0% in undegraded, and by 81.2 % and 92.3 % in MD alpine meadow, respectively. But in SD alpine meadow, ANPP showed no change under any of the N addition treatments in 2021 (Fig.1b). In 2023, ANPP only increased in NH₄⁺-N and Glycine treatments in MD alpine meadow but enhanced in all three N additions in SD alpine meadow (Fig.1c). BNPP showed no change or a slight decrease in all the three alpine meadows in 2020 and 2021, but it dramatically increased in the undegraded and MD alpine meadows under all N addition treatments in 2023 (Fig.1).

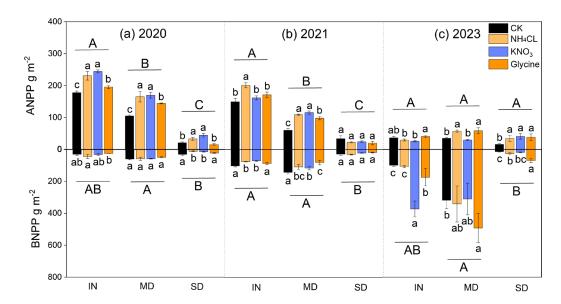


Fig. 1 The effect of different forms of available nitrogen (N) addition on ANPP and BNPP in undegraded, moderately degraded and severely degraded alpine meadows during 2020-2023. IN: undegraded; MD: moderately degraded; SD: severely degraded alpine meadows, respectively; CK: the control. Values are means ± SE (n=3)

Soil physical-chemical properties

The average pH value at the 0–30cm soil in three alpine meadows was above 7 regardless of treatment (Table 1). It decreased in the undegraded meadow but increased in MD and SD alpine meadows with the N addition. TN increased in all the N addition treatments in undegraded and MD alpine meadows, with the largest increase in NH_4 ⁺-N

Table 1 The effects of N addition (NH₄Cl, KNO₃ and Glycine treatment) on soil pH, water content and inorganic N in alpine meadows at different degradation stages

| | IN | | | | MD | | | | SD | | | |
|------------------------|---------------------|---------------------|----------------------|----------------------|---------------------|---------------------|---------------------|----------------------|---------------------|---------------------|---------------------|---------------------|
| | СК | NH ₄ CI | KNO ₃ | Glycine | CK | NH ₄ CI | KNO ₃ | Glycine | СК | NH ₄ CI | KNO ₃ | Glycine |
| рН | 8.16 | 8.06 | 8.12 | 8.13 | 8.34 | 8.38 | 8.56 | 8.58 | 8.85 | 8.79 | 8.82 | 8.84 |
| | (0.00) ^a | (0.02) ^b | (0.01) ^{ab} | (0.02) ^{ab} | (0.01) ^b | (0.01) ^b | (0.00) ^a | (0.01) ^a | (0.01) ^a | (0.02) ^a | (0.00) ^a | (0.01) ^a |
| TN | 0.77 | 1.17 | 0.97 | 0.93 | 0.28 | 0.40 | 0.32 | 0.31 | 0.48 | 0.46 | 0.41 | 0.50 |
| (g kg ⁻¹) | (0.03) ^c | (0.06) ^a | (0.05) ^b | (0.02) ^b | (0.01) ^b | (0.02) ^a | (0.01) ^b | (0.01) ^b | (0.05) ^a | (0.03) ^a | (0.03) ^a | (0.07) ^a |
| Soil C:N | 15.93 | 16.04 | 15.38 | 15.56 | 12.11 | 13.13 | 12.34 | 12.28 | 7.59 | 8.55 | 8.34 | 9.44 |
| | (0.21) ^a | (0.08) ^a | (0.10) ^c | (0.02) ^b | (0.27) ^b | (0.14) ^a | (0.11) ^b | (0.17) ^b | (0.20) ^c | (0.10) ^b | (0.11) ^b | (0.37) ^a |
| Water | 14.60 | 18.62 | 17.95 | 15.54 | 7.60 | 9.01 | 8.35 | 8.58 | 10.81 | 10.93 | 11.66 | 10.07 |
| content (%) | (0.18) ^c | (0.31) ^b | (0.55) ^b | (0.44) ^a | (0.14) ^c | (0.17) ^a | (0.04) ^b | (0.24) ^{ab} | (0.06) ^b | (0.22) ^b | (0.31) ^a | (0.49) ^c |
| Inorganic N | 38.08 | 43.16 | 43.06 | 41.98 | 20.74 | 34.08 | 39.13 | 36.17 | 14.28 | 22.54 | 24.38 | 28.25 |
| (mg kg ⁻¹) | (0.68) ^b | (0.47) ^a | (0.62) ^a | (1.34) ^a | (0.71) ^b | (3.49) ^a | (1.68) ^a | (1.92) ^a | (0.29) ^c | (1.88) ^b | (1.02) ^b | (0.90) ^a |

and no difference between NO₃-N and glycine treatments (Table S1). The soil C:N was significantly increased in MD and SD but showed no significant response in undegraded alpine meadows in all N addition treatments. On average, the relative increase in C:N was larger in SD than in MD alpine meadows (Table S1). The soil inorganic N significantly increased in all meadows. On average, the increase of inorganic N was much higher in MD and SD than in undegraded alpine meadows (Table S1).

Changes in leaf and microbial carbon and nitrogen content and stoichiometry

The community leaf C content showed no significant response to any N addition treatments (Fig. 2a) in undegraded alpine meadow. It slightly increased in the Glycine (5.4%) and NH₄+-N (4.2%) application in MD and SD alpine meadows, respectively. The community leaf N content was only slightly enhanced in Glycine and NH₄+-N treatments in undegraded and MD alpine meadows and it was largely promoted in SD alpine meadow (Fig. 2c). Leaf C:N generally reduced under all the N addition treatments in the three alpine meadows, except the 9.2% increase of Glycine treatment in the MD alpine meadow (Fig. 2e).

The MBC was only increased significantly (by 8.2%) in the NO₃-N addition but MBN increased by 94.7% and 116.2% in NH₄+-N and NO₃-N addition in undegraded alpine meadow (Fig.2b). The NO₃-N and Glycine addition reduced both the MBC and MBN in MD alpine meadow (Figs. 2b and d). By contrast, NH₄+-N addition increased them although was only statistically significant for MBC (Fig. 2b). The addition of inorganic N largely reduced the MBC and MBN, whereas the Glycine had opposite effect on MBC and MBN in SD alpine meadow (Figs. 2b and d). The microbial C:N responses to N addition greatly

varied with different N forms and degradation status. It declined in all three N addition treatments in undegraded alpine meadow but increased in MD alpine meadow (Table S1). In undegraded alpine meadow the greatest reduction of microbial C:N occurred in NO₃-N treatment (58.7%) while the greatest increase in MD alpine meadow was in NH₄+-N treatment (161.7%) (Fig. 2f).

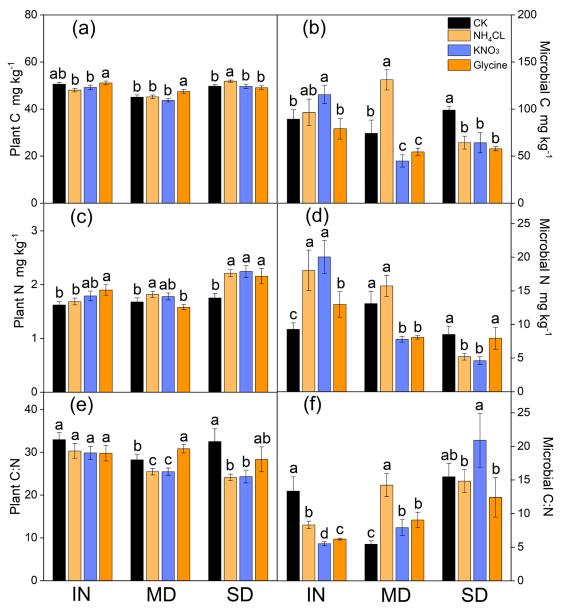


Fig. 2 The effect of different forms of available N addition on leaf C (a), soil microbial C (b), plant N (c), microbial N (d), plant C:N ratio (e) and microbial C:N ratio (f) in alpine meadow at different degradation stages. IN: undegraded; MD: moderately degraded; SD: severely degraded alpine meadows; CK: the controls. Values are means \pm SE (n=3)

Changes in various N pools after different available N forms addition

The plant N pool increased under all N treatments (Fig. 3a). The increase was higher in inorganic N treatments than that of Glycine treatment (Fig. 3a). NH₄*-N increased the plant N pool by 35.2%, 80.3% and 33.6% in undegraded, MD and SD alpine meadows, respectively. NO₃*-N treatment increased them by 52.4%, 77.7% and 65.5%, respectively (Fig. 3a). The plant N uptake rate was only increased in the NH₄*-N treatment (84.8%) in undegraded alpine meadow, but it was increased in all N addition treatments in MD and SD alpine meadows. The increase in plant N uptake rate was not significantly different among the three N additions in either MD or SD alpine meadows (Fig. S2a).

Accumulation of soil inorganic N after one year was much lower in N addition treatments than control with 75.7%, 85.3% and 50.4% less N accumulated in NH₄⁺-N, NO₃⁻-N and Glycine treatments relative to the control in undegraded meadow (Fig. 3b). In MD and SD alpine meadows, the soil inorganic N was decreased in N addition treatments contrast to the accumulation of it in control. In undegraded alpine meadow, the microbial N pool was decreased in the inorganic N additions but increased in the Glycine treatment (Fig. 3c). By contrast, microbial N pool only increased in Glycine treatment in MD and increased in all N addition treatments in SD alpine meadow. The microbial N uptake rate only slightly increased in Glycine treatment in MD (13.8%) and decreased in SD alpine meadows (56.0%) (Fig. S2).

In the undegraded meadow, leaching was only increased in the NO₃-N treatment. But it increased in the Glycine treatment in MD and in both NO₃-N and Glycine treatment in SD alpine meadows (Fig. 3d).

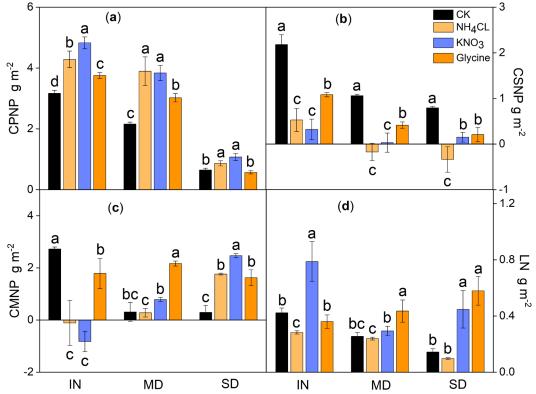


Fig. 3 The effect of different forms of available N addition on plant N pool changes in aboveground and belowground plants in the growing season of 2021 (a), soil available N pool between June 2021 and 2022 (b), microbial N pool (c) and LN (potential N loss due to leaching at the same period) (d) in alpine meadow at different degradation stages. See Figure 1 for abbreviations. Values are means \pm SE (n=3)

Partitioning of N into different active N pools

In undegraded alpine meadow, plants retained 1.11 and 1.66 g N m⁻² of added NH₄⁺-N and NO₃⁻N, respectively. The soil inorganic N and microbial N pool shrank, and leaching showed minimal change (Fig. 4a, b, c). The sum of the four N pools showed a net loss in NH₄⁺-N but a minor gain in NO₃⁻-N and Glycine treatments.

The N retained in plants was greater in any N treatment in MD compared to undegraded alpine meadows. Like the undegraded, the soil inorganic N pool declined in all N addition treatments in MD, yet microbial N pool increased, especially in the Glycine treatment. This contrasted with the reduction observed in the undegraded alpine meadow where the four N pools showed a net gain after N addition, with the greatest gain in NO₃-

The soil inorganic N pool also declined in the SD alpine meadow with N addition but the microbial N pool was greatly enhanced in all N treatments compared to the undegraded and MD alpine meadows. The N addition resulted in a net gain in SD but all of which was lower than in the MD alpine meadow (Figs. 4 g, h, i).

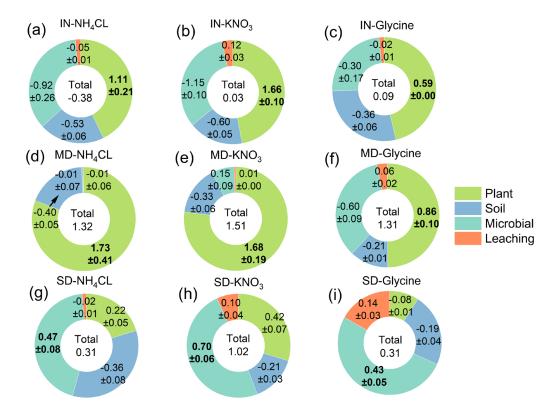


Fig. 4 The partitioning of added N (mg m⁻²) into active N pools and the net balance among them in responding to addition of different available N forms relative to the control (simple mass balance method). Although leaching is a flux not a N pool, the accumulated N in the anion-cation resin could be treated as potential loss N during a period. IN: undegraded; MD: moderately degraded; SD: severely degraded alpine meadows, respectively. Values are means ± SE (n=3)

Discussion

Impact of different forms N addition on NPP of alpine meadows with different degradation statuses

The observation of the largest increase in ANPP in NH₄+-N treatment in the undegraded alpine meadow in 2021 (Fig. 1) supported our first hypothesis, which is in agreement with recent findings that plants use the most abundant available N in grassland ecosystem (Liu et al., 2024). The significant increase in plant N uptake rate in NH₄+-N also could contribute to the largest increase of ANPP in undegraded alpine meadow (Fig. S2). But this phenomenon was not consistent throughout the three years. There was no difference in N addition induced change in ANPP between the N addition treatments in 2020 but in 2021 ANPP increase was much higher in NH₄+-N than in other two forms of N addition. This implies that the impact of different forms of N addition on ANPP may take time to happen. In 2023, the measurement of ANPP and BNPP was conducted close to the senescence period, which may account for the lack of treatment response. Further observations are needed to confirm the impact of different N forms on ANPP.

Plant, microbial and soil inorganic N and C:N change with different forms N addition

Ecosystem retention of N depends largely on the size of plant and soil C pools and their C:N stoichiometric ratio (Lovett & Goodale, 2011; S. Yang et al., 2022). In an N limited ecosystem with high plant and soil C:N, plants and microbes will fiercely compete for the added N (Zhang et al., 2023). Under these circumstances, soil microbes will quickly absorb N to induce a priming effect on soil organic matter decomposition (Chen et al., 2020; Riggs et al., 2015), thereby improving plant N uptake and alleviating N limitation and declining the plant C:N (Lu et al., 2010). In our study, the higher microbial C:N in the control treatment of the undegraded relative to MD alpine meadows (Fig. 2f) suggests a relatively severe N limitation, thus a greater increase of microbial N and decrease in microbial C:N when N

was added (Figs. 2d and f). The significant increase in plant and microbial N contents but no change in C contents (Figs. 2a-d) could contribute to the decline in their C:N in almost all treatments at the three alpine meadows. Microbial C:N generally declined more than plant C:N with added N (Figs. 2e, f) as MBN increased more than leaf N content (Figs. 2c, d), which might be due to superiority of microorganisms in competing for available N (Ouyang et al., 2016). Although the plant N uptake rate was greatly improved in MD (Fig. S2a) and the plant N pool increase was much larger in undegraded and MD than in SD alpine meadows (Figs. 3, 4), the largest increase in plant leaf N content increase occurred in SD (Fig. 2c), which is probably due to growth dilution of assimilated N with the much larger increase in plant productivity in undegraded and MD alpine meadows (Fig. 1).

N rention in active N pools with different forms of additional N in alpine meadows with different degradation statuses

In our study, the difference in N rention among different N forms addition in each alpine meadow is much lower than the difference among degradation statuses (Fig. 3 and 4). The addition of N will significantly increase the available N content in the soil. The available N in the soil will have a significant negative effect on plant C/N and soil C/N, thereby affecting the nANPP and BNPP, promoting plant growth, and leading to the accumulation of N in plants, litter, and soil (Fig. S3). In both the undegraded and MD alpine meadows, no matter what form of N was added, plants retained most of the added N, with the highest retention in inorganic treatments (Fig. 4). Although microbes are superior to plants in absorbing available N in the short term (Kuzyakov & Xu, 2013), in the long-term, plants will get the extra N when microbes mineralize organic N after its N limition is eased (Kuzyakov & Xu,

2013) with an increase in MBN and decline in C:N (Figs. 2d,f). In addition, the increased plant N uptake also could contribute to a higher retention of N in plants (Fig. S2). Although plants in arctic tundra typically prefer to ultilize N in the form of amino acids, our previous studies revealed that dominant species mainly uptake the most avaible form of N (Liu et al., 2024). In our study area that is inorganic N (Lai et al., 2022), which suports the greater increase of plant N uptake rate in the inorganic N addtion than the Glycine addition (Fig. S2).

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

The increase in plant N could be counteracted by the reduction in soil inorganic N pool . The sum of the changes in the active N pools in the undegraded alpine meadow are close to zero (Fig. 4a,b,c) suggesting an internal closed cycing of the added N and no N loss in terms of gaserous or leaching, which is also supported by a near zero ¹⁵N in plants and soil in undegraded alpine meadow (Li et al., 2021b). The net loss of added N in NH₄+-N treatment in the undegraded alpine meadow could be attributed to the larger increase in plant productivity (Fig. 1) as dominant plants of undegraded apline meadow prefer NH₄⁺-N (Lai et al., 2022). The larger increase in plant N will transfer the active N into orgnaic N pools which is evidenced by the greater increase of TN in the undegraded alpine meadow (Table 1). The higher net N gain in MD compared to the undegarded alpine meadow could be the result of a lower reduction in soil inorgnaic N pool and a higher increase in plant N pool (Fig. 3 and 4). The larger rention of N in plants in MD alpine meadow could be the result of the increase in plant N uptake rate and relatively low microbial N uptake rate (Fig. S2). When alpine meadows was degrade, easily decomposable plant litter with lower C:N increases (Lai et al., 2021). These changes would acclelerate the soil orangic matter decompostion and replenish the soil inorganic N pool, therefore result in less reduction of soil inorganic N in MD alpine meadow (Fig. 3 and 4). The smaller increase in TN in MD relative to the undegarded alpine meadow (Table 1) support this.

In SD alpine meadow, microbes retained most of the added N relative to plants (Fig. 4g,h,i) which partly support our second hypothesis as plants retained most the added in MD alpine meadow. The higher microbial C:N in SD than in undegraded and MD alpine meadows (Fig. 2f) indicates a relatively severe N limitation for microbes in SD alpine meadow. Thus, the added N would be mobilzied by microbes which is supported by the realtively greater increase of microbial N pool (Fig. 3c). Although microibal N rention is higher, the net N gain is lower than in MD alpine meadow as leaching is enhanced (Fig. 3 and 4). The changes in soil texture, in particular an increase in silt and sand with alpine meadow degradation (Peng et al., 2018) will increase leaching because of reduction in soil minerals and soil organic matter's adsoprition to added N. The negative charge of soil minerals and organic matter could result in a higher leaching in NO₃-N and Glycine but not in NH₄+N treatment in SD alpine meadow (Fig. 3d).

Conclusion

Although leaching increased with alpine meadow degradation in NO₃⁻-N and Glycine treatments, it only accounted less than 5% of added N. The added N was mostly incorporated into plant biomass in undegraded and MD alpine meadows while it was retained primarily in microorganisms in SD alpine meadows. Plant productivity and plant N uptake rate was higher in the MD alpine meadow with a relatively larger increase in the inorganic N addition treatment. Thus, to restore the degraded alpine meadow, N fertilization

should be applied before the meadow reaches the stage of being moderatley degradaded 428 429 and use of inorganic N, epseicaly the NH₄⁺-N is more efficient. 430 Conflict of Interest: The authors declare that they have no conflict of interest. 431 **Acknowledgements** 432 The study was financially supported by the National Key Research and Development Program of China (Grants No. 2022YFF0801902), National Natural Science Foundation 433 of China (42471074, W2412147), Western Light Project of Chinese Academy of Sciences, 434 435 Grant/Award Number: xbzg-zdsys-202316. Speical thanks to Professor Ji Chen, the 436 director of the field station in supporting our field work. 437 References Ashton IW, Miller AE, Bowman WD, Suding KN (2010) Niche complementarity due to plasticity 438 in resource use: plant partitioning of chemical N forms. Ecology 91:3252-3260. 439 https://doi.org/10.1890/09-1849.1 440 441 Bardgett RD, Bullock JM, Lavorel S, Manning P, Schaffner U, Ostle N, Chomel M, Durigan G, 442 L Fry E, Johnson D, Lavallee JM, Le Provost G, Luo S, Png K, Sankaran M, Hou XY, Zhou HK, Ma L, Ren WB, Li XL, Ding Y, Li YH, Shi HX (2021) Combatting global 443 444 grassland degradation. Nat Rev Earth Env 2:720-735. https://doi.org/10.1038/s43017-445 021-00207-2 446 Brookes P, Landman A, Pruden G, Jenkinson, D (1985) Chloroform fumigation and the release 447 of soil nitrogen: a rapid derect extraction method to measure microbial biamass 448 nitrogen in soil. Soil Biol Biochem 17:837-842. https://doi.org/10.1016/0038-449 0717(85)90144-0 450 Che RX, Wang YF, Li KX, Xu ZH, Hu JM, Wang F, Rui YC, Li LF, Pang Z, Cui XY (2019) 451 Degraded patch formation significantly changed microbial community composition in 452 alpine meadow soils. Soil Till Res 195:104426. https://doi.org/10.1016/j.still.2019.104426 453 454 Chen QY, Niu B, Hu YL, Wang J, Lei TZ, Xu-Ri, Zhou JZ, Xi CW, Zhang GX (2020) Multilevel Nitrogen Additions Alter Chemical Composition and Turnover of the Labile Fraction Soil 455 Organic Matter via Effects on Vegetation and Microorganisms. J Geophys Res-Biogeo 456 125: e2019JG005316. https://doi.org/10.1029/2019jg005316 457 458 Chen KE, Chen HY, Tseng CS, Tsay YF (2020) Improving nitrogen use efficiency by 459 manipulating nitrate remobilization in plants. Nat Plants 6:1126-1135. 460 https://doi.org/10.1038/s41477-020-00758-0 Chen SL, Chen B, Wang SQ, Sun LG, Shi H, Liu ZH, Wang QY, Li H, Zhu TT, Li DH, Xia Y, 461 Zhao ZQ, Wang LC, Wang LZ (2023) Spatiotemporal variations of atmospheric nitrogen 462

- deposition in China during 2008–2020. Atmos Environ 315:120120. https://doi.org/10.1016/j.atmosenv.2023.120120
- Choudhary S, Blaud A, Osborn AM, Press MC, Phoenix GK (2016) Nitrogen accumulation and partitioning in a High Arctic tundra ecosystem from extreme atmospheric N deposition events. Sci Total Environ 554:303–310. https://doi.org/10.1016/j.scitotenv.2016.02.155
- Di HJ, Cameron KC (2002) Nitrate leaching in temperate agroecosystems: sources, factors and mitigating strategies. Nutr Cycl Agroecosys 46:237–256.

 https://doi.org/10.1023/A:1021471531188
- Dong SK, Xu YD, Li S, Shen H, Yang MY, Xiao JN (2024) Restoration actions associated with payment for ecosystem services promote the economic returns of alpine grasslands in China. J Clean Prod 458:142732. https://doi.org/10.1016/j.jclepro.2024.142439
- Du E, Terrer C, Pellegrini AFA., Ahlström A, van Lissa CJ, Zhao X, Xia N, Wu X, Jackson RB (2020) Global patterns of terrestrial nitrogen and phosphorus limitation. Nat Geosci 13:221–226. https://doi.org/10.1038/s41561-019-0530-4
- Fu G, Shen ZX (2016) Response of Alpine Plants to Nitrogen Addition on the Tibetan Plateau:
 A Meta-analysis. J Plant Growth Regul 35:974–979. https://doi.org/10.1007/s00344-479
 O16-9595-0
- Gu SS, Zhou XL, Yu H, Yan HL, Wang YC, Liu Y, Wang ZH, Feng K, Du XF, Lu GX, Deng Y (2023) Microbial and chemical fertilizers for restoring degraded alpine grassland. Biol Fertil Soils 59:911–926. https://doi.org/10.1007/s00374-023-01759-9
- Harrison KA, Bol R, Bardgett RD (2007) Preferences for different nitrogen forms by coexisting plant species and soil microbes. Ecology 88:989–999. https://doi.org/10.1890/06-1018
- Kuzyakov Y, Xu XL (2013) Competition between roots and microorganisms for nitrogen:
 mechanisms and ecological relevance. New Phytol 198:656–669.
 https://doi.org/10.1111/nph.12235
- L'Espérance E, Bouyoucef LS, Dozois JA, Yergeau E (2024) Tipping the plant-microbe competition for nitrogen in agricultural soils. iScience 27:110973. https://doi.org/10.1016/j.isci.2024.110973
- Lai CM, Li CY, Peng F, Xue X, You QG, Zhang WJ, Ma SX (2021) Plant community change mediated heterotrophic respiration increase explains soil organic carbon loss before moderate degradation of alpine meadow. Land Degrad Dev 32:5322–5333. https://doi.org/10.1002/ldr.4111
- Lai CM, Peng F, Sun JB, Zhou J, Li CY, Xu XL, Chen XJ, You QG, Sun HY, Sun J, Xue X, Lambers H (2022) Niche differentiation and higher uptake of available nitrogen maintained the productivity of alpine meadow at early degradation. Biol Fertil Soils 59:35–49. https://doi.org/10.1007/s00374-022-01679-0
- Li CY, Lai CM, Peng F, Xue X, You QG, Liu FY, Guo PL, Liao J, Wang T (2021a) Dominant Plant Functional Group Determine the Response of the Temporal Stability of Plant Community Biomass to 9-Year Warming on the Qinghai-Tibetan Plateau. Front Plant Sci 12:704138. https://doi.org/10.3389/fpls.2021.704138
- Li CY, Lai CM, Peng F, Xue X, You QG, Liu FY, Guo PL, Liao J, Wang T (2021b) Plant community changes determine the vegetation and soil δ13C and δ15N enrichment in degraded alpine grassland. Land Degrad Dev 32:2371–2382. https://doi.org/10.1002/ldr.3912

- Li J, Sang CP, Yang JY, Qu LR, Xia ZW, Sun H, Jiang P, Wang XG, He HB, Wang C (2021)
 Stoichiometric imbalance and microbial community regulate microbial elements use
 efficiencies under nitrogen addition. Soil Biol and Biochem 156:108207.
 https://doi.org/10.1016/j.soilbio.2021.108207
- Li CY, Lai CM, Peng F, Zhou J, Zhang W, Song XY, Luo SY, Sun JB, Chen XJ, Chen B, Chen J, Xue X (2024) Restoration of degraded alpine meadows from the perspective of plant–soil feedbacks. Biol Fertil Soils 60:941–953. https://doi.org/10.1007/s00374-024-514
- Li JH, Han YW, Ye LF, Deng HD, Gao XT, Soromotin AV, Kuzyakov Y, Knops JMH, Abbott LK (2023) Effects of nitrogen and phosphorus fertilization on soil organic matter priming and net carbon balance in alpine meadows. Land Degrad Dev 34:2681–2692. https://doi.org/10.1002/ldr.4642
- Liang X, Zhang T, Lu X, Ellsworth DS, BassiriRad H, You C, Wang D, He P, Deng Q, Liu H, Mo
 J, Ye Q (2020) Global response patterns of plant photosynthesis to nitrogen addition:
 A meta-analysis. Glob Chang Biol 26:3585–3600. https://doi.org/10.1111/gcb.15071
- Liu JS, Li XF, Ma QH, Zhang X, Chen Y, Isbell F, Wang DL (2019) Nitrogen addition reduced ecosystem stability regardless of its impacts on plant diversity. J Ecol 107:2427–2435. https://doi.org/10.1111/1365-2745.13187
- Liu M, Li CC, Xu XL, Wanek W, Jiang N, Wang HM, Yang XD (2017) Organic and inorganic nitrogen uptake by 21 dominant tree species in temperate and tropical forests. Tree Physiol 37:1515–1526. https://doi.org/10.1093/treephys/tpx046
- Liu M, Xu XL, Wanek W, Sun J, Bardgett RD, Tian YQ, Cui XY, Jiang LL, Ma ZQ, Kuzyakov Y,
 Ouyang H, Wang YF (2024) Nitrogen availability in soil controls uptake of different
 nitrogen forms by plants. New Phytol. https://doi.org/10.1111/nph.20335
- Liu SB, Schleuss PM, Kuzyakov Y (2016) Carbon and Nitrogen Losses from Soil Depend on Degradation of Tibetan Kobresia Pastures. Land Degrad Dev 28:1253–1262. https://doi.org/10.1002/ldr.2522
- Liu Y, Li SY, Shi JJ, Niu YL, Cui Z, Zhang ZC, Wang YL, Ma YS, López-Vicente M, Wu GL (2022)
 Effectiveness of mixed cultivated grasslands to reduce sediment concentration in runoff
 on hillslopes in the Qinghai-Tibetan Plateau. Geoderma 422:115933.

 https://doi.org/10.1016/j.geoderma.2022.115933
- Liu YW, Wang YS, Pan YP, Piao SL (2015) Wet deposition of atmospheric inorganic nitrogen at five remote sites in the Tibetan Plateau. Atmospheric Chem Phys 15:11683–11700. https://doi.org/10.5194/acp-15-11683-2015
- Lovett GM, Goodale CL (2011) A New Conceptual Model of Nitrogen Saturation Based on Experimental Nitrogen Addition to an Oak Forest. Ecosystems 14:615–631. https://doi.org/10.1007/s10021-011-9432-z
- Lu M, Yang YH, Luo YQ, Fang CM, Zhou XH, Chen JK, Yang X, Li B (2010) Responses of ecosystem nitrogen cycle to nitrogen addition: a meta-analysis. New Phytol 189:1040– 1050. https://doi.org/10.1111/j.1469-8137.2010.03563.x
- 547 Ouyang SN, Tian YQ, Liu QY, Zhang L, Wang RX, Xu XL (2016) Nitrogen competition between 548 three dominant plant species and microbes in a temperate grassland. Plant Soil 549 408:121–132. https://doi.org/10.1007/s11104-016-2904-3
- Peng F, Xue X, You QG, Huang CH, Dong SY, Liao J, Duan HC, Tsunekawa A, Wang T (2018)

- Changes of soil properties regulate the soil organic carbon loss with grassland degradation on the Qinghai-Tibet Plateau. Ecol Indic 93:572–580. https://doi.org/10.1016/j.ecolind.2018.05.047
- Qin W, Feng J, Zhang Q, Yuan X, Zhou H, Zhu B (2024) Nitrogen and phosphorus addition mediate soil priming effects via affecting microbial stoichiometric balance in an alpine meadow. Sci Total Environ 908:168350. https://doi.org/10.1016/j.scitotenv.2023.168350
- Riggs CE, Hobbie SE, Bach EM, Hofmockel KS, Kazanski CE (2015) Nitrogen addition changes grassland soil organic matter decomposition. Biogeochemistry 125:203–219. https://doi.org/10.1007/s10533-015-0123-2
- Sun Y, Wang CT, Chen HYH, Ruan HH (2020) Responses of C:N stoichiometry in plants, soil, and microorganisms to nitrogen addition. Plant Soil 456:277–287. https://doi.org/10.1007/s11104-020-04717-8
- Tang B, Rocci KS, Lehmann A, Rillig MC (2023) Nitrogen increases soil organic carbon accrual and alters its functionality. Glob Chang Biol 29:1971–1983. https://doi.org/10.1111/gcb.16588
- Tian J, Dungait JAJ, Lu XK, Yang YF, Hartley IP, Zhang W, Mo JM, Yu GR, Zhou JZ, Kuzyakov Y (2019) Long-term nitrogen addition modifies microbial composition and functions for slow carbon cycling and increased sequestration in tropical forest soil. Glob Chang Biol 25:3267–3281. https://doi.org/10.1111/qcb.14750

571

572

573574

- Wang R, Wu H, Sardans J, Li T, Liu H, Peñuelas J, Dijkstra FA, Jiang Y (2020) Carbon storage and plant-soil linkages among soil aggregates as affected by nitrogen enrichment and mowing management in a meadow grassland. Plant Soil 457:407–420. https://doi.org/10.1007/s11104-020-04749-0
- Wang YF, Lv W.W, Xue K, Wang SP, Zhang LR, Hu RH, Zeng H, Xu XL, Li YM, Jiang LL, Hao YB, Du JQ, Sun JP, Dorji T, Piao SL, Wang CH, Luo CY, Zhang ZH, Chang XF, Zhang MM, Hu YG, Wu TH, Wang JZ, Li BW, Liu PP, Zhou Y, Wang A, Dong SK, Zhang XZ, Gao QZ, Zhou HK, Shen MG, Wilkes A, Miehe G, Zhao XQ, Niu HS (2022) Grassland changes and adaptive management on the Qinghai–Tibetan Plateau. Nat Rev Earth Env 3:668–683. https://doi.org/10.1038/s43017-022-00330-8
- Wang Z, Tao T, Wang H, Chen J, Small GE, Johnson D, Chen J, Zhang Y, Zhu Q, Zhang S, Song Y, Kattge J, Guo P, Sun X (2023) Forms of nitrogen inputs regulate the intensity of soil acidification. Glob Chang Biol 29:4044–4055. https://doi.org/10.1111/gcb.16746
- Widdig M, Heintz-Buschart A, Schleuss PM, Guhr A, Borer ET, Seabloom EW, Spohn M (2020)
 Effects of nitrogen and phosphorus addition on microbial community composition and
 element cycling in a grassland soil. Soil Biol and Biochem 151:108041.
 https://doi.org/10.1016/j.soilbio.2020.108041
- Xu GH, Fan XR, Miller AJ (2012) Plant nitrogen assimilation and use efficiency. Annu Rev Plant Biol 63:153–182. https://doi.org/10.1146/annurev-arplant-042811-105532
- 590 Xu H, Qu Q, Li G, Liu G, Geissen V, Ritsema CJ, Xue S (2022) Impact of nitrogen addition on 591 plant-soil-enzyme C–N–P stoichiometry and microbial nutrient limitation. Soil Biol and 592 Biochem 170:108714. https://doi.org/10.1016/j.soilbio.2022.108714
- Xue X, Guo J, Han BS, Sun QW, Liu LC (2009) The effect of climate warming and permafrost thaw on desertification in the Qinghai–Tibetan Plateau. Geomorphology 108:182–190.

| 595 | https://doi.org/10.1016/j.geomorph.2009.01.004 |
|-----|---|
| 596 | Yang Y, Liu H, Yang X, Yao HJ, Deng XQ, Wang YQ, An SS, Kuzyakov Y, Chang SX (2022) |
| 597 | Plant and soil elemental C:N:P ratios are linked to soil microbial diversity during |
| 598 | grassland restoration on the Loess Plateau, China. Sci Total Environ 806:150557. |
| 599 | https://doi.org/10.1016/j.scitotenv.2021.150557 |
| 600 | Yang S, Liu WX, Guo LL, Wang CZ, Deng MF, Peng ZY, Liu LL (2022) The changes in plant |
| 601 | and soil C pools and their C:N stoichiometry control grassland N retention under |
| 602 | elevated N inputs. Ecol Appl 32:e2517. https://doi.org/10.1002/eap.2517 |
| 603 | Yang T, Long M, Smith MD, Gu Q, Yang YD, He NP, Xu C, Wu HH, Vilonen L, Zhao JL, Jentsch |
| 604 | A, Yu Q (2021) Changes in species abundances with short-term and long-term nitrogen |
| 605 | addition are mediated by stoichiometric homeostasis. Plant Soil 469:39–48. |
| 606 | https://doi.org/10.1007/s11104-021-05141-2 |
| 607 | Zhang L, Liu J, Xi JZ, Pang R, Gunina A, Zhou SR (2023) Competition for nitrogen between |
| 608 | plants and microorganisms in grasslands: effect of nitrogen application rate and plant |
| 609 | acquisition strategy. Biol Fertil Soils 60:227–236. https://doi.org/10.1007/s00374-023- |
| 610 | 01782-w |
| 611 | Zhang X, Wang Z, Lü XT, Han X, Yu FH (2026) Contrasting priming effects in soils subjected to |
| 612 | long-term urea versus inorganic nitrogen addition. Soil Till Res 256:106918. |
| 613 | https://doi.org/10.1016/j.still.2025.106918 |
| 614 | Zhang Y, Yang YS, Osborne B, Zhou HK, Wu JY, Zhang WW, Zou JL (2024) Edaphic factors |
| 615 | control microbial biomass and elemental stoichiometry in alpine meadow soils of the |
| 616 | Tibet Plateau. Plant Soil. https://doi.org/10.1007/s11104-024-06578-x |
| 617 | Zhu ZK, Zhou J, Shahbaz M, Tang HM, Liu SL, Zhang WJ, Yuan HZ, Zhou P, Alharbi H, Wu JS, |
| 618 | Kuzyakov Y, Ge T (2021) Microorganisms maintain C:N stoichiometric balance by |
| 619 | regulating the priming effect in long-term fertilized soils. Appl Soil Ecol 167:104033. |
| 620 | https://doi.org/10.1016/j.apsoil.2021.104033 |
| | |