

1           **ADMS simulation and influencing factors of bioaerosol**  
2           **diffusion from BRT under different aeration modes in six**  
3           **wastewater treatment plants**

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34 **Abstract**

35 Bioaerosols produced by municipal wastewater treatment plants (MWTP) can spread  
36 in air, thereby polluting the atmosphere and causing safety hazards to workers and  
37 surrounding residents. In this study, the biological reaction tanks (BRTs) of six MWTPs  
38 undergoing typical processes in North China, Yangtze River Delta, and the Greater Bay  
39 Area were selected to set up sampling points and investigate the production  
40 characteristics of bioaerosols. The Atmospheric Dispersion Modelling System method  
41 was used to simulate the diffusion of bioaerosols in the MWTPs. The concentrations of  
42 bacteria and, specifically, intestinal bacteria in the bioaerosols ranged from 389 CFU/m<sup>3</sup>  
43 to 1,536 CFU/m<sup>3</sup> and 30 CFU/m<sup>3</sup> to 152 CFU/m<sup>3</sup>, respectively, and the proportion of  
44 the intestinal bacteria was 8.85%. The concentration of soluble chemicals (SCs) in the  
45 bioaerosols was 18.36 µg/m<sup>3</sup>–82.19 µg/m<sup>3</sup>, and the main SCs found were Mg<sup>2+</sup>, Ca<sup>2+</sup>,  
46 and SO<sub>4</sub><sup>2-</sup>. The proportion of intestinal bacteria (75.79%) produced via surface aeration  
47 by a BRT attached to large-sized bioaerosol particles was higher than that of a BRT  
48 undergoing the bottom aeration process (37.28%). The main microorganisms found in  
49 the bioaerosols included Moraxellaceae, Escherichia–Shigella, Psychrobacter, and  
50 Cyanobacteria. The generation of bioaerosols exhibited regional characteristics. The  
51 wastewater treatment scale, wastewater quality, and aeration mode were the main  
52 factors influencing bioaerosol production. Model simulation showed that, after 1 h, the  
53 diffusion distance of bioaerosol was 292 m–515 m, and the affected area was 42,895  
54 m<sup>2</sup>–91,708 m<sup>2</sup>. The diffusion distance and range of the bioaerosols were significantly  
55 correlated with the concentration at the bioaerosol source and the aeration mode  
56 adopted by the BRTs. Wind speed and direction were two environmental factors that  
57 affected the diffusion of bioaerosols. With an increase in the diffusion distance, the  
58 concentration of microorganisms, intestinal bacteria, ions, and fine particles in the  
59 bioaerosols decreased significantly, resulting in a corresponding reduction in the  
60 exposure risk. This study provides new insights to help predict bioaerosol risks at  
61 MWTPs and identify safe areas around MWTPs. The study also provides a basis for  
62 selecting safe MWTP sites and reducing bioaerosol pollution risks.

63 **Key words:** Wastewater treatment, Bioaerosol, Generation characteristics, Trajectory  
64 prediction, Diffusion factors

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68 **1. Introduction**

69 Wastewater treatment is considered a major source of bioaerosols. Most urban  
70 wastewater treatments use activated sludge processes that use microorganisms to  
71 degrade organic matter in wastewater into harmless substances. However, oxygen is  
72 required to maintain the growth of the microorganisms and the reaction. Therefore,  
73 biochemical treatment processes usually adopt aeration, and microorganisms may enter  
74 the air during the process of bioaerosol generation. When bioaerosols contain intestinal  
75 bacteria, viruses, or fungi and their spores that cause allergic reactions, they can cause  
76 diseases in the human respiratory tract (Brenner et al. 1988) . The exposure risk and  
77 pathogenesis of bioaerosol requires further investigation.

78  
79 Bioaerosols containing bacteria and fungi have been detected in the air around  
80 biological reaction tanks (BRTs) during oxidation ditch, Sequencing Batch Reactor  
81 Activated Sludge Process(SBR), and Anaerobic-Anoxic-Oxic (A<sup>2</sup>/O) processes(Li et al.  
82 2011, Xu et al. 2018). In a previous study, the concentration of particles in bioaerosol  
83 produced as a result of wastewater treatment reached 1,233/m<sup>3</sup>–6,533/m<sup>3</sup>, and the  
84 microbial concentration reached more than 1,690 CFU/m<sup>3</sup> (Li et al. 2016). The particle  
85 size of bioaerosols in municipal wastewater treatment plants (MWTPs) is generally  
86 2.1µm–3.3 µm (Laitinen et al. 1994). Bioaerosols produced by MWTPs may contain  
87 various pathogenic microorganisms, such as viruses (rotavirus, norovirus, adenovirus,  
88 hepatitis virus, and coliphages) and intestinal bacteria (Micrococcus, Bacteroides,  
89 Chryseobacterium, Pseudomonas, and Acinetobacter). They can cause infections  
90 following inhalation, swallowing, and skin contact (Gotkowska-Plachta et al. 2013,  
91 Wang et al. 2018b, Wang et al. 2018c). Previous studies on wastewater treatment of  
92 bioaerosols have mainly focused on their emission levels, particle size characteristics,  
93 and population structure (Li et al. 2011, Wang et al. 2018c).

94  
95 Infective pathogens can be transported over long distances via airflow(van Doremalen  
96 et al. 2020). It was found that the concentration of microorganisms decreased  
97 significantly at 500 meters downwind of the bioaerosol source, and each MWTP  
98 presented its own bioaerosol diffusion characteristics. BRTs are sewage treatment  
99 facilities built outdoors. The bioaerosol generated by them will spread around the  
100 MWTP with the wind, bringing potential risks to the downwind area. Bioaerosol  
101 transport and dispersion models that describe the spatial and temporal distributions of

102 aerosol concentrations are commonly generated using computational fluid dynamics  
103 (CFD), which usually represent the turbulent transport of momentum and energy using  
104 eddy diffusivities in the corresponding governing equations (Hayashi et al. 2002, Yang  
105 et al. 2014). The viruses in public places, bioaerosols in hospital diagnosis and  
106 treatment sites and particulate matter emissions from aquaculture farms were calculated  
107 and simulated by this method to predict the infection caused by the spread of particulate  
108 pollutants and prevent the exposure risk brought about by them (Archer et al. 2019,  
109 Zhang et al. 2020). Analytical solutions obtained using the Gaussian-like Atmospheric  
110 Dispersion Modelling System (ADMS) based on the CFD principle can provide a quick  
111 prediction of the probability of taking in bioaerosol. In this study, the ADMS was used  
112 to study bioaerosol diffusion at the MWTP scale.

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114 Because it is helpful to reduce bioaerosols and their exposure risk, studies on bioaerosol  
115 diffusion characteristics have attracted increasing attention. In this study, six MWTPs  
116 undergoing oxidation ditch, SBR, and A<sup>2</sup>/O processes were investigated in three regions  
117 of China. The chemical composition of bioaerosol was analyzed by ion chromatography.  
118 High throughput sequencing was used to analyze microbial population in bioaerosols.  
119 The ADMS was used to simulate the diffusion of bioaerosols from the MWTPs to  
120 explore the diffusion law of bioaerosols. Analyzing the influencing factors of bioaerosol  
121 diffusion and evaluating the bioaerosol transport risk may contribute to risk predictions  
122 of bioaerosols from MWTPs, which, in turn, can provide a basis for safely selecting  
123 MWTP sites and reducing bioaerosol pollution risks.

124

## 125 **2 Materials and methods**

### 126 **2.1 Municipal MWTP descriptions**

127 Sampling sites for bioaerosol capture were set up 1.5 m above the aeration tanks of six  
128 MWTPs. The six MWTPs, NC-O and NC-A in North China, YZD-O and YZD-A in the  
129 Yangtze River Delta, GBA-O and GBA-A at the Greater Bay Area. Treatment processes  
130 are listed in Table S1. Samples were collected during the morning from 8 to 10 a.m.  
131 without disturbing the normal operation of the aerators. Meteorological parameters, e.g.,  
132 temperature (Tem), relative humidity (RH), wind speed (WS) and solar radiation (SR),  
133 at the given sampling site were also recorded using portable equipment (Table S1). The  
134 air temperature and RH were determined using a Dewpoint Thermohygrometer (WD-  
135 35612, OAKTON, Germany). The WD and WS were recorded using a portable

136 anemometer (HD2303, Delta OHM, Padova, Italy) while the SR was measured using a  
137 portable irradiance meter (HD2302.0, Delta OHM, Italy).

138

## 139 **2.2 Bioaerosol capture**

140 A six-stage viable Andersen Impactor (228–9530 K, SKC Gulf Coast, Inc., Houston,  
141 TX, USA) with 400 holes was used to capture bioaerosols at each sampling site as  
142 described in a previous study (Wang et al. 2018c). A culture medium was kept under  
143 each stage of the sampler. The sampling time was typically 3 min at a speed of 28.3  
144 L/min and the total impaction volume was 84.9 L (Table S1). Dishes were removed  
145 from the sampler when the required volume of air had been drawn. Bacteria and  
146 intestinal bacteria were cultured by LB and MacConkey Agar Medium (Hopebio,  
147 China), respectively. In this study, air samples were collected every hour (8 a.m., 9 a.m.,  
148 10 a.m.), and two parallel samples were collected each time. A total of 6 samples were  
149 taken at each sampling site and the data was the average of the 6 samples. After each  
150 sample was collected, the sampler was sterilised with a solution of 75% ethanol. Results  
151 were calculated as the geometric mean of the replicates, expressed as colony-forming  
152 units per cubic meter of air (CFU/m<sup>3</sup>). A TH-150 medium flow sampler (Medium Flow  
153 sampler, Wuhan, China) was applied to collect the total suspended particulates (TSPs)  
154 in the atmosphere. The glass fiber membrane with diameter of 90 mm was utilized as  
155 the deposition medium. The particle rejection coefficient of medium was 99.90%. The  
156 glass fiber membranes were dried in a desiccator for 48 hours in advance.

157

## 158 **2.3 Analysis method**

### 159 **2.3.1 Chemicals analysis**

160 Concentrations of SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, Cl<sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, and NO<sub>2</sub><sup>-</sup> anions in each sample were  
161 determined with an ion chromatogram analyser (ICS-3000, Dionex, Sunnyvale, CA,  
162 USA). Concentrations of NH<sub>4</sub><sup>+</sup>, Ca<sup>2+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, and Na<sup>+</sup> cations in each sample were  
163 determined with an ion chromatogram analyser (IC plus 883, ion chromatography  
164 system, Metrohm, Herisau, Switzerland). The chromatography apparatus included a  
165 column oven, conductivity detector, manual injector, and chromatography workstation  
166 (Metrohm); AS19 Column and Metrosep C 4150/4.0 separation column; eluent: 20 mM

167 NaOH (anions), 2.0 mM HNO<sub>3</sub> (cations); column temperature: 30°C; flow-rate: 1.0  
168 mL/min; inject volume: 10 L. The detection limit of the method was less than 0.05 mg/L  
169 for anions and cations.

170

### 171 **2.3.2 Microbial population**

172 Illumina MiSeq high-throughput sequencing was applied to analyse the bacteria  
173 population in the bioaerosols. DNA was extracted, purified, and sequenced as described  
174 in previous studies(Wang et al. 2018c). After raw FASTQ files were demultiplexed and  
175 quality filtered, operational taxonomic units (OTUs) were clustered with a 97%  
176 similarity cut-off and chimeric sequences were identified and removed. The taxonomy  
177 of each 16S rRNA gene sequence was analysed using a confidence threshold of 70%  
178 (Amato et al. 2013, Jiang et al. 2015). Each sample was normalized at the same  
179 sequence depth (34,536 reads). The alpha and beta diversity and similarity of the  
180 bacterial community's structure were analysed(Table S2).

181

### 182 **2.3.3 Simulation on bioaerosol**

183 Atmospheric Dispersion Modelling System (ADMS) was an advanced dispersion  
184 model used to obtain the air quality impact of existing and proposed industrial  
185 installations which was originally developed for regulatory authorities in the UK. It has  
186 been widely applied to simulate the diffusion of gases or particles in various situations  
187 (Hood et al. 2020, Jaffe et al. 2020). ADMS was utilized in the simulation of bioaerosol  
188 diffusion in this study. Considering the change of microbial activity in bioaerosols, this  
189 study adjusted and improved the model parameters, such as the type selection of  
190 particles representing aerosols, dry and wet sedimentation, boundary layer processes,  
191 chemical reactions and topography. The adjusted model is compared with the observed  
192 results and fitted with the actual measured values. In order to simulate the diffusion  
193 characteristics of bioaerosols accurately, we measured and obtained ground structures  
194 and terrain conditions in more detail. The aerosols were simulated from the six MWTPs  
195 studied in this study. The simulated area size was within 1000000 m<sup>2</sup>, Meteorological  
196 data are obtained from the meteorological Bureau (<http://www.weather.com.cn/>) and  
197 underlying surface data are obtained from field monitoring obtain meteorological  
198 parameters, and input the obtained data including temperature, humidity, illumination,  
199 wind direction and wind speed; Underlying surface data such as terrain building height  
200 into building modules.

201

## 202 **2.4 Exposure Risk assessment**

203 The average daily doses of a respiratory inhalation model were calculated using Eqs.  
204 (1) (Liu et al. 2018, Vilavert et al. 2012).

$$205 \quad \text{ADD}_{\text{inhalation}} = \frac{c \times \text{InR} \times \text{EF} \times \text{ET}}{\text{BoW} \times \text{ALE}} \quad (1)$$

206 Where  $\text{ADD}_{\text{inhalation}}$  is the average daily dose to the respiratory system (CFU/d/kg),  $c$  is  
207 the average bioaerosol concentration (CFU/m<sup>3</sup>),  $\text{InR}$  is the inhalation rate (m<sup>3</sup>/d),  $\text{EF}$  is  
208 the exposure frequency (d/yr),  $\text{ET}$  is the respiratory inhalation exposure time (yr),  $\text{ALE}$   
209 is their average life expectancy (d), and  $\text{BoW}$  is the body weight of the exposed person  
210 (kg). The dose rate was estimated from the average bioaerosol concentrations. As the  
211 people exposed in this study were mainly adults, the objects of the greatest risk  
212 evaluation were adults. Most of the exposure parameters of Chinese people proposed  
213 by Wang et al. (2009) were employed in this study (Wang et al. 2009). Some parameters  
214 such as average body weight ( $\text{BoW}$ ) and average time ( $\text{ALE}$ ), were retrieved from the  
215 Exposure Factors Handbook of the Chinese Population (Adult) based on the  
216 behavioural characteristics of environmental exposure in China in 2017.

217

218 Functional Annotation of Prokaryotic Taxa (FAPROTAX) was a functional annotation  
219 database of culturable microorganisms (Liang et al. 2019). Substitute the 16S based  
220 OTU classification in the high-throughput sequencing of microorganisms into the  
221 python script linked to the OTU classification and the FAPROTAX database to obtain  
222 the prediction results of microbial community functions (Parfrey et al. 2016).  
223 FAPROTAX was utilized to forecast bioaerosols hazard in this study.

224

## 225 **3. Results**

### 226 **3.1 Aerosol emission**

#### 227 **3.1.1 Aerosol emission level**

228 Figure 1 demonstrates the bacteria and intestinal bacteria in bioaerosol escaped in the  
229 BRT section of the six MWTPs. The concentrations of bioaerosols presented upwind  
230 were also measured with 275 CFU/m<sup>3</sup> for NC-O, 177 CFU/m<sup>3</sup> for NC-A, 75 CFU/m<sup>3</sup>  
231 for YZD-O, 42 CFU/m<sup>3</sup> for YZD-A, 99 CFU/m<sup>3</sup> for GBA-O, and 51 CFU/m<sup>3</sup> for GBA-  
232 A, respectively. The emission level of bioaerosols found at all BRTs were higher than  
233 those presented in the air upwind (Table S1). The concentration of airborne bacteria at

234 the MWTPs ranged from 389 CFU/m<sup>3</sup> to 1,536 CFU/m<sup>3</sup>, with the concentrations in the  
235 NC-O, NC-A, YZD-O, YZD-A, GBA-O, and GBA-A being 1,536 CFU/m<sup>3</sup>, 1,009  
236 CFU/m<sup>3</sup>, 1,184 CFU/m<sup>3</sup>, 554 CFU/m<sup>3</sup>, 1,056 CFU/m<sup>3</sup>, and 389 CFU/m<sup>3</sup>, respectively.  
237 MWTPs using the surface aeration method had a higher concentration of bacteria (1,258  
238 CFU/m<sup>3</sup>). Intestinal bacteria were found at each MWTP, with concentrations ranging  
239 from 30 CFU/m<sup>3</sup> to 1,52 CFU/m<sup>3</sup>, accounting for 8.04% of the total bacterial population.  
240 MWTPs in the NC area generated more bacteria and, specifically, intestinal bacteria  
241 (with averages of 1,273 CFU/m<sup>3</sup> and 140 CFU/m<sup>3</sup>, respectively) than did those in the  
242 other areas (which had averages of 796 CFU/m<sup>3</sup> and 47 CFU/m<sup>3</sup>, respectively). Figure  
243 2 shows the soluble chemicals (SCs) detected in bioaerosols at the six MWTPs. SCs  
244 were detected at each MWTP, and the amount of ten kinds of SCs ranged from  
245 18.36µg/m<sup>3</sup>–82.19 µg/m<sup>3</sup>. Bioaerosols at the MWTPs of the NC and GBA areas had  
246 higher SC contents (with an average of 73.74 µg/m<sup>3</sup>) than did those in the YZD area  
247 (with an average of 37.89 µg/m<sup>3</sup>). Mg<sup>2+</sup>, Ca<sup>2+</sup>, and SO<sub>4</sub><sup>2-</sup> were the major SCs in the  
248 bioaerosols at each MWTP, accounting for 29.84% of the total SC content.

249

### 250 **3.1.2 Aerosol particle size distribution**

251 The size distributions of the bioaerosol particles to which the airborne and intestinal  
252 bacteria attached at each MWTPs are shown in Figure 3. The aerodynamic cut-size  
253 diameters of the Six Stage Viable Andersen Cascade Impactor are: over 7.0 µm in stage  
254 1, 4.7-7.0 µm in stage 2, 3.3-4.7 µm in stage 3, 2.1-3.3 µm in stage 4, 1.1-2.1 µm in  
255 stage 5, 0.65-1.1 µm in stage 6. The microbial aerosol particles collected in this study  
256 were classified as fine particles which picked from the stage 5 and the stage 6, and  
257 coarse particles that sampled from the stage 1 to the stage 4. The inhalation of particles  
258 smaller than 10 µm has detrimental health effects, such as causing asthma and  
259 cardiovascular disease (Urbán et al. 2015, Vestlund et al. 2014). At the sampling sites  
260 NC-O, YZD-O, and GBA-O, 69.73% to 80.56% of the intestinal bacteria were attached  
261 to coarse particles with an average concentration of 64 CFU/m<sup>3</sup>, while the average  
262 concentration of the fine particles was 23 CFU/m<sup>3</sup>. At the sampling sites NC-A, YZD-  
263 A, and GBA-A, 62.72% of the intestinal bacteria on average were attached to fine  
264 particles. The concentrations of fine and coarse particles were 4 CFU/m<sup>3</sup>-24 CFU/m<sup>3</sup>  
265 and 10 CFU/m<sup>3</sup>-68 CFU/m<sup>3</sup>, respectively.

266

### 267 **3.1.3 Microbial population in bioaerosols**



268 This study analyzed the microbial populations at each MWTP. Table S3 shows the  
269 composition and proportion of the microorganisms at each MWTP. The main  
270 microorganism species in the bioaerosols varied among MWTPs. Moraxellaceae  
271 (11.17%), Pseudomonas (7.11%), and Chroococciopsis (6.64%) dominated the  
272 bacterial population in NC-A. Pseudomonas, Escherichia–Shigella, and Psychrobacter  
273 accounted for 84.63% of the bacterial population in NC-O. Cyanobacteria was the  
274 dominant microorganism in YZD-A, accounting for 80.46% of the overall bacterial  
275 population, while in YZD-O, Cyanobacteria (28.65%), Candidatus (8.76%), and  
276 Saccharibacteria (4.55%) were the most abundant. The dominant species in GBA-A  
277 were Sphingomona (9.74%), Sphingomonadales (6.97%), and Mitochondria (6.29%),  
278 while Peptostreptococcaceae (13.79%), Mycobacterium (4.75%), and  
279 Sphingobacteriaceae (4.47%) were the dominant bacteria in GBA-O. Intestinal bacteria,  
280 including Moraxellaceae, Pseudomonas, Acinetobacter, Arcobacter, Neisseriaceae,  
281 Escherichia–Shigella, Mycobacterium, Saccharibacteria, Romboutsia, Serratia, and  
282 Flexibacter, were detected in bioaerosols at the MWTPs. Pseudomonas is a common  
283 pathogenic bacterium that can cause endocarditis, osteomyelitis, pneumonia, urinary  
284 tract infections, gastrointestinal infections, meningitis, and sepsis(Rice et al. 2012).  
285 Diseases caused by Pseudomonas are becoming more common(Gomez-Gamboa et al.  
286 2021). Acinetobacter mainly causes respiratory and urinary tract infections, sepsis, and  
287 secondary meningitis (Gao et al. 2014, Gou et al. 2016). Serratia can cause urinary tract,  
288 respiratory tract, and wound infections(Kowalski et al. 2017). Escherichia–shigella is a  
289 common pathogenic bacterium that can cause extra-intestinal infections, acute diarrhea,  
290 bacillary dysentery, and other diseases (Kotloff et al. 2018, Rodriguez-Angeles 2002).  
291 Therefore, in addition to strengthening the personal protection of workers, such as by  
292 wearing masks and goggles, BRTs, especially the oxidation ditch, need to be equipped  
293 with protective covers and other facilities to prevent the escape of bioaerosols.

294

#### 295 **3.1.4 Bioaerosol generation**

296 There were differences in the number, particle size distribution, and composition of  
297 total bacteria and Enterobacteriaceae in aerosols released from each sewage treatment  
298 plant, and the concentrations of SCs in the aerosols differed. Similar results were  
299 reported in previous studies(Wang et al. 2018a, Wang et al. 2018d). Reasons for this  
300 difference might be differences in regional characteristics and aeration modes. The  
301 water quality and environmental conditions of sewage treatment plants differ regionally.

302 Aerosol generation is related to water quality and environmental conditions. Therefore,  
303 the concentration of microorganisms generated in NC, which has a high concentration  
304 of microorganisms in its sewage, was high. The diffusion of bioaerosols produced by  
305 wastewater treatment plants undergoing surface oxygenation and bottom oxygenation  
306 differed within the same area. This may be related to the different mechanisms of  
307 bioaerosol generation as a result of bottom aeration and surface aeration. The bottom  
308 aeration process filled the bottom of the water body with gas, and the gas formed  
309 bubbles at the bottom of the liquid and gradually rose to the liquid surface. During this  
310 process, sludge flocs from the bottom and middle of the liquid were carried to the  
311 surface. During the process of surface rupture, the bubbles reaching the surface  
312 transported substances in the sewage to the air, resulting in a loss of water and  
313 aerosolizing in the air, thereby forming aerosols. Surface aeration oxygenation facilities  
314 are located at liquid surfaces and often disturb water surfaces owing to the motion of  
315 rotating brushes. After flapping and raising, water surface liquid is wrapped with  
316 substances in sewage and is transported into the air, and the splashing of large droplets,  
317 water loss, and atomization of small droplets leads to bioaerosols generation near the  
318 specific brush. Data on aerosol production characteristics in this study also confirmed  
319 that both surface and bottom aeration produces bioaerosols. The higher the amount of  
320 bioaerosols produced, the higher the concentration in the air at 200 m-DW. Compared  
321 with surface aeration, bottom aeration produces less bioaerosol. The concentration in  
322 the air emitted by bottom aeration was also lower at 200 m-DW.

323

### 324 **3.2 Bioaerosol diffusion**

325 The ADMS was used to calculate the diffusion of bioaerosols, microorganisms,  
326 particles, and water-soluble ions. The FLOWSTAR model was used to calculate flow  
327 and turbulence fields over the terrain by introducing meteorological parameters. Wind  
328 direction and wind speed are the dominant factor influencing bioaerosol diffusion  
329 direction and range (Wei et al. 2020). The rose chart of the wind direction in North  
330 China, the Greater Bay Area, and the Yangtze River Delta are shown in Figure 5. The  
331 dominant wind directions in North China were south (44.66%) and north (22.74%).  
332 East (20.55%), northeast (16.71%), and north (15.07%) winds were dominant in  
333 Yangtze River Delta. North (25.21%), southeast (21.10%), and northeast (20.01%)  
334 winds were dominant in the Greater Bay Area.

335

336 **3.2.1 Aerosol diffusion direction**

337 The concentrations of bioaerosols produced at the MWTPs were used to simulate the  
338 diffusion of bacteria in air using the ADMS model (Figure 6 and Table S3). The  
339 simulation results showed that bioaerosols from the MWTPs diffused evenly. After 1 h  
340 of diffusion, high aerosol concentrations were found close to the generation source, and  
341 their diffusion was distributed in clumps. MWTPs in the same region had similar  
342 diffusion directions that closely correlated to the wind direction. The diffusion trends  
343 in NC-A and NC-O had a southwest-northwest (139°) elliptic distribution. The  
344 diffusion trends in GBA-A and GBA-O exhibited southeast to northwest (164°) elliptic  
345 distributions. Diffusion in YZD-A exhibited an elliptic distribution with a southwest-  
346 northeast (243°) trend, while that in YZD-O exhibited a west-east trend (285°) elliptic  
347 distribution.

348

349 **3.2.2 Aerosol diffusion scope**

350 The concentrations of bacteria in the aerosol diffusion direction at 100 m (100 m-DW;  
351 the boundary of the MWTPs) and 200 m (200 m-DW; residential areas) varied among  
352 the MWTPs. The bioaerosol concentrations at 100 m-DW and 200 m-DW in NC-A and  
353 NC-O ranged from 63 CFU/m<sup>3</sup>–75 CFU/m<sup>3</sup> and 25 CFU/m<sup>3</sup>–35 CFU/m<sup>3</sup>, respectively.  
354 The spread of bioaerosol from YZD-A and YZD-O at 100 m-DW and 200 m-DW  
355 ranged from 41 CFU/m<sup>3</sup>–66 CFU/m<sup>3</sup> and 18 CFU/m<sup>3</sup>–50 CFU/m<sup>3</sup>, respectively. The  
356 spread of bioaerosol in GBA-A and GBA-O also ranged from 20 CFU/m<sup>3</sup>–72 CFU/m<sup>3</sup>  
357 and 50 CFU/m<sup>3</sup>–55 CFU/m<sup>3</sup> at 100 m-DW and 200 m-DW, respectively. The  
358 concentration of intestinal bacteria at 100 m-DW and 200 m-DW at each MWTP ranged  
359 from 1 CFU/m<sup>3</sup>–8 CFU/m<sup>3</sup> and 1 CFU/m<sup>3</sup>–3 CFU/m<sup>3</sup>, respectively. Bacteria attached to  
360 fine particles also decreased with increasing distance, with concentrations ranging from  
361 4 CFU/m<sup>3</sup>–22 CFU/m<sup>3</sup> and 3 CFU/m<sup>3</sup>–11 CFU/m<sup>3</sup> at 100 m-DW and 200 m-DW,  
362 respectively. The total concentration of SCs decreased to 3.62 µg/m<sup>3</sup> and 2.02 µg/m<sup>3</sup> on  
363 average at 100 m-DW and 200 m-DW, respectively.

364

365 The bioaerosol diffusion distance (292–515 m) and affected area (42,895–91,708 m<sup>2</sup>)  
366 differed among the MWTPs. Aerosols in NC-O spread 423 m in the northwest direction  
367 and affected an area of 61,385 m<sup>2</sup>. The microorganisms in NC-A spread 301 m  
368 northwest, with an affected diffusion area of approximately 42,895 m<sup>2</sup>. Microorganisms  
369 in YZD-O spread 307 m northwest, affecting an area of 47,333 m<sup>2</sup>. The diffusion

370 distance of microorganisms in YZD-A was 554 m, and the diffusion area was 80,614  
371 m<sup>2</sup>. Microorganisms in GBA-O spread 292 m northwest, with an affected diffusion area  
372 of approximately 48,073 m<sup>2</sup>. Microorganisms in GBA-A spread 515 m northwest, with  
373 an affected diffusion area of approximately 91,708 m<sup>2</sup>. The concentration at the  
374 diffusion edge was as low as in the range of 2 CFU/m<sup>3</sup>–20 CFU/m<sup>3</sup>.

375

### 376 **3.2.3 Factors influencing diffusion**

#### 377 *Influence of source strength on the diffusion range*

378 Diffusion analysis of aerosols generated at the MWTPs indicated that there were  
379 differences in the aerosol diffusion results during the BRT stage. Mechanical  
380 disturbances were major causes of aerosol generation. The high microbial content and  
381 intense sewage aeration led to a large amount of aerosol being generated in the BRT  
382 section, as supported by previous studies(Wang et al. 2019, Yang et al. 2019). Therefore,  
383 the bioaerosol concentrations in NC-O were higher at 200 m-DW than at the MWTPs  
384 in the other areas. The level of bioaerosol generated at the source influenced bioaerosol  
385 diffusion; the higher the level at the source, the more microorganisms dispersed in the  
386 downward direction.

387

388 The species and concentrations of bacteria varied with place, depending on the process  
389 selected and meteorological parameters (e.g., temperature, RH, and WS). Canonical  
390 correlation analysis (CCA) was conducted to describe the influence of these factors on  
391 bacterial populations in bioaerosols (Figure S1) (Li et al. 2017). The results  
392 demonstrated that environmental factors influence bacterial emissions. Among them,  
393 RH, temperature, SR, and WS were the major factors influencing the level of airborne  
394 bacteria in the air, which is consistent with results obtained in previous reports(Yan et  
395 al. 2019a). The RH, temperature, and SR were positively correlated with airborne  
396 intestinal bacteria release and dispersal, whereas WS was negatively correlated with  
397 airborne intestinal bacteria release and dispersal (Table 3). High temperatures and RH  
398 favor microbiological growth, whereas strong winds help disperse bioaerosol particles.  
399 Owing to the suitable temperature, appropriate RH, and low WS, higher bioaerosol  
400 concentrations were maintained in the air. Similar results were obtained from CCA of  
401 strain-influencing factors. Some microbes were sensitive to meteorological conditions.  
402 CCA results also showed that environmental factors have different effects on different  
403 types of microorganisms. The RH had a strong positive effect on *Pseudomonas*,

404 Psychrobacter, and Serratia that were positively affected by temperature, and most of  
405 the bacteria were negatively correlated with WS and SR.

406

407 Changes in microbial concentrations in bioaerosols are related to WS. The results  
408 showed that a high WS is conducive to aerosol diffusion, whereas light intensity has  
409 the opposite effect. A high RH is favorable for bacterial activity and diversity during  
410 diffusion. WS and RH are major factors influencing aerosol diffusion and are important  
411 parameters in estimating aerosol diffusion. The range of influence of bioaerosol  
412 diffusion is related to WS; the higher the WS, the greater the range of influence of  
413 aerosols. Pseudomonas, Escherichia–Shigella, Psychrobacter, and Serratia were  
414 strongly affected by environmental factors. Intestinal bacteria accounted for  
415 approximately 8.85% of the total bacterial population and may pose risks to human  
416 health following breathing or skin contact.

417

#### 418 ***Influence of meteorological parameters on diffusion***

419 Temperature, RH, SR, WS, and other environmental conditions influenced aerosol  
420 diffusion. Sampling of the six MWTPs occurred during summer at an air temperature  
421 of 31.9°C–37.2 °C. The optimal growth temperature for most bacteria in bioaerosols is  
422 10°C–30 °C (Rai et al. 2021). The RH ranged from 31.5%–57.5% at the MWTPs. The  
423 concentration of bioaerosols in the air at the MWTP was high when the RH was high.  
424 The CCA results also showed that a high RH is favorable for bacterial survival in air.  
425 Most microorganisms require water for survival; therefore, the RH is positively  
426 correlated with the abundance of microorganisms. A higher RH in air is conducive to  
427 the survival of microorganisms. (Yan et al. 2019b). The SR was in the range of 216.16  
428 W/m<sup>2</sup>–585.6 W/m<sup>2</sup>, and the CCA result revealed a negative correlation with  
429 microorganism abundance. Most bacteria are sensitive to light, and UV light may  
430 inactivate and kill microorganisms, thereby negatively affecting the diffusion of  
431 bioaerosols (Fahimipour et al. 2018). WS had an important relationship with the  
432 diffusion speed and distance of the bioaerosols. The wind direction affected the  
433 diffusion direction. Therefore, temperature, RH, SR, and WS were affecting bioaerosol  
434 diffusion. When simulating diffusion, it was suggested to consider the influence of  
435 pathogenic bacteria, particle size, and ions to make the evaluation results more accurate  
436 and conducive to risk prevention.

437

### 438 3.3.1 Risk assessment

439 As bioaerosols contain intestinal bacteria, they can be used to estimate the exposure  
440 risk of people downwind of aerosols produced by sewage plants. The results of an  
441 exposure dose calculation and a risk assessment are shown in Figure 7. The ranges of  
442  $HQ_{\text{inhalation}}$  inside the MWTPs were  $3.59 \times 10^{-2}$ – $1.42 \times 10^{-1}$ . The concentration of bacteria  
443 was higher in the air undergoing surface aeration ( $1.16 \times 10^1$ ) than in that undergoing  
444 submerged aeration ( $6.01 \times 10^{-2}$ ). The hazard quotient (HQ) and HI decreased with  
445 increasing distance downwind. The  $HQ_{\text{inhalation}}$  of the bacteria at 100 m-DW and 200 m-  
446 DW at each MWTP ranged from  $1.85 \times 10^{-3}$ – $6.93 \times 10^{-3}$  and  $9.24 \times 10^{-4}$ – $4.62 \times 10^{-3}$   
447 respectively, which were 5.88% and 3.28% on average in the BRT section, respectively.  
448 The  $HQ_{\text{inhalation}}$  of the bacteria in air undergoing surface aeration at 100 m-DW and 200  
449 m-DW were  $6.56 \times 10^{-3}$  and  $4.16 \times 10^{-3}$ , respectively, which were higher than those in air  
450 undergoing submerged aeration ( $3.82 \times 10^{-3}$  and  $1.63 \times 10^{-3}$ , respectively). Bioaerosol  
451 diffusion probably exerted higher risks at 100 m-DW and 200 m-DW in GBA-A than  
452 at the other MWTPs due to its higher  $HQ_{\text{inhalation}}$  ( $1.66 \times 10^{-3}$  and  $3.79 \times 10^{-3}$ , respectively).  
453 The  $HQ_{\text{inhalation}}$  of intestinal bacteria at 100 m-DW and 200 m-DW at each MWTP  
454 ranged from  $9.24 \times 10^{-5}$ – $7.39 \times 10^{-4}$  and  $9.24 \times 10^{-4}$ – $4.62 \times 10^{-3}$ , respectively. The individual  
455 risks were low at the sites investigated, and they remained below the acceptable levels  
456 stipulated by the USEPA (1 for noncarcinogenic pollutants). As the risk to the skin was  
457 several orders of magnitude lower than that following inhalation and was negligible,  
458 respiratory inhalation was regarded as the main pathway for microbial bioaerosol entry  
459 into the human body. The HI reached its highest level in NC-O. The order of exposure  
460 HQ following inhalation was: children>adult males>adult females. It is worth noting  
461 that the exposure hazard for children is generally higher than that for adults owing to  
462 their lower weight.

463

464 Figure 2S demonstrated the prediction results of FAPROTAX microbial community  
465 function at BRT of all MWTPs. A total of 7 dominant predicted functional groups were  
466 detected, including chemoheterotrophy, aerobic chemoheterotrophy, nitrate reduction,  
467 fermentation, human associated functional group and pathogenic functional groups  
468 (animal parasites or symbionts, human pathogens). Among them, human associated  
469 functional group, human pathogens and animal parasites or symbionts, were higher at  
470 NC-A, indicating that the microorganism at NC-A has potential risk to human health.

471

### 472 **3.3.2 Factors influencing exposure risk**

473 The results of the evaluation obtained using the aerosol generation source and total  
474 bacterial value diffused to the outside indicated that the exposure risk of bioaerosols  
475 was negligible. The MWTPs in the NC area had higher internal and downwind risks.  
476 Meanwhile, because of the high WS, the NC area had a wider bioaerosol exposure risk  
477 range. It is worth noting that, compared with bottom aeration, the risk of bioaerosols  
478 produced by surface aeration was greater. With an increase in the distance from the  
479 aerosol generation source, the concentration of bioaerosols in the air gradually  
480 decreased, and the exposure risk decreased accordingly.

481

#### 482 ***Impact of microorganism abundance on bioaerosol risk***

483 The microorganism concentrations were used to calculate the bioaerosol risk in this  
484 study. However, this risk of bioaerosol in reality might be higher than evaluated.  
485 Microorganisms present in the air may multiply during transportation. (Fahimipour et  
486 al. 2018). In addition, air contains  $Mg^{2+}$ ,  $Ca^{2+}$ ,  $SO_4^{2-}$ , and other chemicals, which may  
487 create a microenvironment suitable for the survival of airborne microorganisms. With  
488 the progression of diffusion, microorganisms in air generally lose their activity or die.  
489 However, aerosols produced in water may behave differently from those in other  
490 environments, such as soil and vegetation. Aeration generates a droplet membrane that  
491 breaks the water surface to generate aerosols. Therefore, most aerosols produced in  
492 water contain microorganisms in the droplet membrane. The droplets help the  
493 microorganisms to attach to the aerosol particles, reduce the direct irradiation of some  
494 ultraviolet rays, protect the viability of some microorganisms, and protect  
495 microorganisms from external damage (e.g., wind-induced damage) or death. Droplet  
496 membranes also increase the humidity in the environment of microorganisms and allow  
497 bacteria to survive. Meanwhile, water and soluble chemicals in the droplet membrane  
498 can provide nutrients for microorganisms.

499

500 The current risk assessment calculation method does not consider intestinal bacteria,  
501 small particles, SCs, or other factors. Table S3 presents the species and genera of  
502 intestinal bacteria. After diffusion, intestinal bacteria, SCs, and fine particles were still  
503 present. The detected concentrations of intestinal bacteria were 2 CFU/m<sup>3</sup>–8 CFU/m<sup>3</sup>  
504 and 1 CFU/m<sup>3</sup>–3 CFU/m<sup>3</sup> when intestinal bacteria spread to 100 DW and 200 DW,  
505 respectively; therefore, intestinal bacteria pose a potential risk. Intestinal bacteria,

506 including *Pseudomonas*, *Acinetobacter*, and *Serratia*, were detected in the aerosols.  
507 They may proliferate and maintain their activity during diffusion, thereby increasing  
508 the risk associated with aerosols. Second, although the number of intestinal bacteria  
509 was low, the current calculation method does not consider pathogenicity. If bacteria are  
510 pathogenic, although the risk of exposure is low, the risk of long-term exposure for on-  
511 site workers cannot be ignored.

512

513 Fine particles of 4 CFU/m<sup>3</sup>–24 CFU/m<sup>3</sup> and 3 CFU/m<sup>3</sup>–11 CFU/m<sup>3</sup> were also detected  
514 at 100-DW and 200-DW, respectively. The exposure risk was mainly related to the  
515 concentration of bacteria. The number of bacteria decreased gradually from the  
516 proximal end to the distal end of diffusion, and the amount of coarse and fine particles  
517 also decreased. At the proximal end of diffusion, the aeration mode had a greater impact.  
518 Surface aeration produces coarser particles that mainly affect the upper respiratory tract.  
519 Because of the sedimentation of coarse particles, the distal end (100 m-DW and 200 m-  
520 DW) of diffusion is dominated by fine particles, which may enter deeper parts of the  
521 human body.

522

523 Soluble ions at concentration of 1.36μg/m<sup>3</sup>–5.60 μg/m<sup>3</sup> and 0.60μg/m<sup>3</sup>–3.89 μg/m<sup>3</sup> were  
524 detected at 100 m-DW and 200 m-DW, respectively. Soluble chemicals diffused far into  
525 the air to provide nutrients for microorganisms and help them maintain vitality.  
526 Therefore, the bioaerosols produced by sewage treatment plants may contain more  
527 active intestinal bacteria.

528

529 In the current risk assessment calculation method, intestinal bacteria, fine particles, SCs,  
530 and other factors are not considered. Therefore, on-site protection should be  
531 strengthened. Suggestions for protecting workers include wearing masks and protective  
532 clothing.

533

#### 534 **4. Conclusions**

535 Concentrations, particle size distribution, population, and soluble ion number of the  
536 total bacteria and intestinal bacteria of bioaerosol in the six MWTPs present variation  
537 because of regional differences and aeration modes. MWTPs in North China generated  
538 more bioaerosol compared with other areas, while MWTPs with surface aeration  
539 process produced higher amounts of bioaerosol than submerge aeration process. Wind



540 direction affected the diffusion direction. The diffusion distance and range were related  
541 to the WS and source strength of the bioaerosol. The dispersion of bioaerosols with  
542 different particle sizes varied. Surface aeration resulted in more aerosol diffusion. With  
543 an increase in the distance from the MWTPs, the exposure risk gradually decreased.  
544 The number of microorganisms, pathogenic bacteria, ions and fine particles affect the  
545 risk of diffusion. Since there may be more than one source of bioaerosol production,  
546 this part of the study can be used to distinguish whether bioaerosols in residential  
547 exposure areas come from MWTPs, thus speculate the impact of bioaerosol generated  
548 by MWTPs on surrounding area. This study simulated the bioaerosol diffusion  
549 generated from MWTP, helping MWTPs' sites selection and reducing bioaerosols'  
550 possible risks.

551

552

### 553 **[Acknowledgements]**

554 We would like to thank Editage ([www.editage.cn](http://www.editage.cn)) for English language editing.

555

### 556 **[Funding sources]**

557 This project is supported by the special fund of State Key Joint Laboratory of  
558 Environment Simulation and Pollution Control (21K01ESPCR).

559

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