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TITLE: Does freeze-thaw action affect the extractability and bioavailability of Pb and As in contaminated soils?

RUNNING TITLE: Pb and As bioavailability changes in control and freeze-thaw soils

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Highlights

- Freeze-thaw increased the bioavailability of soil Pb and As;
- Changes were detectable under free-thaw treatments using a sequential extraction;
- In vitro simulation methods showed differences in freeze-thaw treated soils;
- Particle size changes after freeze-thaw were linked to Pb/As bioavailability change.

21 **Abstract**

22 As global warming intensifies, there will be increased uncertainty as to the environmental behavior and risks from
23 heavy metals in industrial/legacy contaminated sites in permafrost regions. Bioavailability has been increasingly used
24 for human health risk assessment of heavy metals in contaminated soils. Soil heavy metal bioavailability depends on
25 soil physicochemical properties, and freeze-thaw affects soil physical, chemical and biological processes. However
26 it is not clear whether freeze-thaw has an effect on the bioavailability of soil heavy metals. In this study, soils
27 contaminated with Pb and As were collected from 10 industrial sites in northeast China, an area prone to freeze-thaw
28 action. Extractability and bioavailability of soil As and Pb were determined by the Tessier sequential extraction
29 method and four in vitro gastronomic-intestinal simulation methods under control and freeze-thaw treatments. The aims
30 were: to compare the results of extraction and bioavailability from laboratory experiments which artificially simulate
31 freeze-thaw conditions against control soils; to explore the correlation between bioavailability of Pb/As and soil
32 properties. Freeze-thaw cycling significantly decreased soil pH, and increased the soil weight surface area. Results
33 using the Tessier sequential extraction procedure showed that freeze-thaw decreased the percentage in the residual
34 fraction, and increased the percentage of Pb and As in the exchangeable fraction, carbonate-bound fraction, Fe-Mn
35 oxides-bound fraction and organic-bound fraction, relative to control soils. Results using four simulation methods
36 showed that freeze-thaw significantly increased Pb and As bioavailability compared to the controls. Pb and As
37 released in the gastric digestion phase of the four methods was significantly higher than that in the intestinal digestion
38 phase. Further analysis of correlations between Pb and As bioavailability and soil properties indicated that total
39 concentrations of Al, Fe and Mn, particle size, and weight surface area significantly correlated to Pb and As
40 bioavailability. Overall, this study demonstrated that freeze-thaw did influence the bioavailability of soil heavy metals.
41 It suggests the freeze-thaw action should be comprehensively considered in the human risk assessment of soil
42 pollutants in permafrost regions.

43

44 **Keywords:** Heavy metals; Bioavailability; Tessier sequential extraction method; in vitro simulation methods;
45 Seasonal permafrost regions; Industrial contaminated land

46

47 **1. Introduction**

48 Soils are subjected to freezing-thawing over large parts of the globe. Indeed, freezing-thawing is a key factor in
49 soil formation in many parts of the world (Henry, 2007; Kim et al., 2012). Permanent and seasonal permafrost areas

50 cover ~23% of the global land area (Zhou et al., 2018). Seasonal permafrost freezes for one month or more, at depths
51 of a few tens of millimeters to a few meters; seasonal permafrost is cyclic, freezing in winter and thawing in spring
52 and summer. It is estimated that the global area of stable permafrost will decrease from $1.07 \times 10^5 \text{ km}^2$ in 2011 to 5.6
53 $\times 10^4 \text{ km}^2$ by 2110 (Wei et al., 2011). It has been hypothesized that pollutants accumulated in permafrost areas as a
54 "sink" may then be re-mobilised and transformed during permafrost thawing, with unknown consequences (Wei et
55 al., 2011). In China, soil heavy metal contamination is a key type of soil pollution, especially for As and Pb (Ministry
56 of Environmental Protection of the People's Republic of China Ministry of Land and Resources of the People's
57 Republic of China, 2014). As part of the increased development and re-development of urban areas, a large number
58 of polluting enterprises in seasonal tundra areas have moved out from urban centers, leaving behind potentially
59 contaminated sites, for example, old industrial bases in northeast China. Therefore, as global warming intensifies,
60 there will be increased uncertainty as to the environmental behavior and risks from heavy metals in industrial/legacy
61 contaminated sites in permafrost regions.

62 Understanding the fraction of contaminants that potentially enter human body is a key task of human health risk
63 assessment. Often the focus for risk assessment of heavy metals in contaminated sites/soils is their potential to be
64 absorbed by humans, through oral, dermal, and respiratory routes (Kelley et al., 2002). The use of bioavailability for
65 human health risk assessment of soil heavy metals has been increasingly recognized by researchers and has resulted
66 in a more mature human health risk system (USEPA, 2001; USEPA, 2017a; USEPA, 2017b). Many countries have
67 also included bioavailability into the human health risk assessment of soil heavy metals (Ng et al., 2009; USEPA,
68 2007). The essence of freeze-thaw action is obviously the freezing and thawing of soil water, i.e., the repeated freezing
69 and thawing of the soil surface layer and soil layers at certain depths when the temperature drops below 0 degrees
70 and rises above 0 degrees. The physical effect produced by freezing and thawing will accelerate the fragmentation of
71 large solid media, enhance water release and the water permeability coefficient of soil, and be accompanied by the
72 leaching and migration of soluble components during the thawing process (Gao et al., 2016; Hou et al., 2021; Luo
73 and Li, 2021). Freeze-thaw affects soil physical (e.g., particle size, soil porosity, and specific surface area), chemical
74 (e.g., pH and organic matter), and biological processes (e.g., soil microbes and soil fauna) (Edwards, 2010; Lehrs, 1998).
75 Although it is not clear whether freeze-thaw has a particularly important effect on soil heavy metal
76 bioavailability, recent studies have found that freeze-thaw can affect the transformation of heavy metal forms (Sun
77 et al., 2016; Wang, 2017). Freeze-thaw cycles affected the mobilization and enrichment of heavy metals (Hou et al.,
78 2020, 2022). The increase in pH under freeze-thaw can reduce metal bioavailability (Chen, 2021), while organic

79 matter can complex and chelate with heavy metal ions, thus reducing the bioavailability and mobility of heavy metals
80 (Li et al., 2013). Based on this, it is reasonable to assume that freeze-thawing may affect the release, forms and
81 bioavailability of soil heavy metals, although very little research has systematically investigated these processes to
82 date.

83 Chemical extraction methods (Sun et al., 2006), in vivo animal-based methods (Brattin & Casteel, 2013; Juhasz
84 et al., 2010) and in vitro simulation methods (Li et al., 2016; Zhu et al., 2019) have been commonly used as the
85 assessment methods for soil heavy metal bioavailability. Chemical extraction methods have been widely used to
86 estimate the bioavailability of heavy metals; primary extraction methods (e.g. EDTA) and sequential extraction
87 methods (e.g. the Tessier sequential extraction methodology) have been used. Although the evaluation of the
88 mobilisation of heavy metals in soil by chemical extraction methods is relatively simple to perform, it obviously will
89 not accurately mimic conditions in the gastro-intestinal tract, where absorption occurs. It has disadvantages and
90 shortcomings, such as inaccurate characterization, poor comparability, and susceptibility to the influence of soil
91 physical and chemical properties (Sun et al., 2006). The limitations of chemical extraction methods have become
92 increasingly evident, with the increasing requirements of environmental health risk assessment. In vivo bioassays in
93 different animal models have been used to assess the bioavailability of soil heavy metals, but due to ethical and cost
94 issues, in vitro gastrointestinal simulation methods have been developed in recent years. Gastrointestinal simulation
95 methods are intended to determine the dissolution of soil contaminants under conditions designed to simulate the
96 human gastrointestinal tract, which is a key step determining bioavailability (i.e. absorption across the GI-tract lining).
97 Examples of gastro-intestinal simulation methods include: the Physiologically Based Extraction Test (PBET), the
98 Unified BARGE Method (UBM), the In Vitro Gastrointestinal Method (IVG) and the Soluble Bioavailability
99 Research Consortium (SBRC) method. Around 10 different gastrointestinal simulation methods are available
100 internationally now, but there is still a lack of carefully designed inter-comparison and calibration exercises, and there
101 is not yet a recommended or preferred procedure, supported by the wider research community. The results from
102 different gastrointestinal simulation methods for the same metals were found to vary widely in previous studies, for
103 example (Li et al., 2020; Li et al., 2016). The SBRC has now been recommended by the USEPA as the standard
104 method for predicting the biological effectiveness of soil Pb. Li et al. (2016) showed the ability of the IVG method
105 to effectively predict the relative biological effectiveness of As, based on the in vivo - in vitro correlation (IVIVC)
106 model. Zhu et al. (2019) confirmed that the UBM can be used as a valid method to predict the relative biological
107 effectiveness of Pb, by constructing a mouse-UBM correlation experiment.

108 The objectives and design of this study were: to sample 10 soils from contaminated industrial legacy sites in
109 northeast China, a major seasonal permafrost region; to compare the performance of established extraction (Tessier
110 sequential extraction method) and in vitro simulation methods (PBET, IVG, SBRC and UBM) for Pb and As present
111 in the soils from the legacy sites, in relation soil properties, ease of test performance etc.; to compare the extraction
112 and bioavailability results from laboratory experiments which artificially simulate freeze-thaw conditions against
113 soils maintained under control conditions. The results are discussed with respect to the test performances, influence
114 of soil properties, the assays used and the potential importance of freeze-thaw actions on heavy metal behavior and
115 bioavailability in soils.

116

117 **2. Materials and methods**

118 *2.1. Soil sample collection and freeze-thaw experiment*

119 Pb and As contaminated sites left by industries in cities in northeast China were selected for this study. A grid
120 of 40 × 40 square meters was delineated at the geographic center of the sites. The diagonal method was used to collect
121 samples from 5 points; the manual drilling method was used for sampling, to 0-20 cm depth. A total of 10
122 contaminated soil samples were collected from three types of industrial site: 4 steel plant sites, 5 non-ferrous metal
123 smelting and processing plant sites, and 1 pharmaceutical factory site. The soil for testing and experiments was air-
124 dried at room temperature, then debris such as gravel and plant residues were removed. The soils were then loosely
125 disaggregated, sieved through 2 mm and 250 μm, then stored in sealed bags for chemical analysis and freeze-thaw
126 treatment, respectively. **The 250 μm soil samples were transferred into a sealed bag, and the relative field water**
127 **holding capacity of the soil was adjusted to 60%. Soils were incubated at room temperature (10 ± 5°C) for 15 days**
128 **before freeze-thaw experiments (Zhang et al., 2013).**

129 Using local meteorological data for northeast China, the freeze-thaw experiments used the following conditions:
130 soil moisture content, 60%; soil freezing/thawing temperatures, -15°C and 6°C. With reference to "Standard Test
131 Methods for Freezing and Thawing Compacted Soil-Cement Mixtures" (ASTMD560, 2016) issued by the American
132 Society for Testing and Materials, the number of freeze-thaw cycles was set to 12. **Incubated soils were placed in a**
133 **low-temperature freezer at -15°C for 24 hours, then taken out and thawed in a refrigerator at 6°C for 24 hours. One**
134 **freeze-thaw cycle was therefore 48 hours, and the process was repeated 12 times (Gu, 2020). At the same time, control**
135 **soils were maintained at room temperature (10 ± 5°C) with 0 cycles of freeze-thaw treatment.**

136 *2.2. Determination of soil properties and metal concentrations*

137 The <2 mm sieved samples were used for determining the soil properties and metal concentration. These samples
 138 were used to determine soil pH (potentiometric method), cation exchange capacity (CEC) (barium chloride-sulfuric
 139 acid forced exchange method) and soil organic matter content (SOM) (potassium dichromate titration method). Soil
 140 particle surface area was determined by the BET specific surface area test and particle size distribution was
 141 determined using a TopSizer laser. Total metal concentrations were determined using the U.S. Environmental
 142 Protection Agency's digestion method (USEPA 3050B), with quantification by inductively coupled plasma mass
 143 spectrometry (ICP-MS).

144 2.3. Properties of the soils used in the study

145 Soil physicochemical properties and total metal concentrations are given in Table 1. In summary, values range
 146 as follows: soil pH, 7.20-7.59; SOM, 1.78-91.4 g/kg; CEC, 1.31-12.6 cmol/kg; mean particle size, 11.18-115 μm ;
 147 weight specific surface area, 48.7-296 m^2/g ; conductivity, 45.3-251 $\mu\text{s}/\text{cm}$. The key variables soil pH and SOM are
 148 broadly typical for soils from northeast China (Sun et al., 2020; Zhang, 2011). The total concentrations of the major
 149 elements Al, Fe and Mn, which can influence the chemistry of Pb, were $\sim 18,000$ - $55,000$ mg/kg, $\sim 37,000$ - $130,000$
 150 mg/kg and ~ 690 - $4,000$ mg/kg, respectively. Pb concentrations ranged between ~ 130 - $27,000$ mg/kg, As between 77-
 151 930 mg/kg. Background (uncontaminated) soils are typically ~ 10 - 60 mg Pb/kg and ~ 2 - 35 mg As/kg (China National
 152 Environmental Monitoring Centre, 1990; Sun et al., 2019b), so these soils are clearly contaminated with Pb. The total
 153 concentrations of Pb, Al, Fe and Mn significantly correlated with each other (Figure S1).

154
 155 Table 1. Characteristics of the soils used in the study.

Samples	Soil 01	Soil 02	Soil 03	Soil 04	Soil 05	Soil 06	Soil 07	Soil 08	Soil 09	Soil 10
pH	7.40	7.59	7.25	7.48	7.53	7.55	7.32	7.50	7.49	7.20
σ ($\mu\text{s}/\text{cm}$)	59.4	58.9	76.6	47.8	77.9	52.7	214.0	80.2	45.3	251.0
CEC (cmol/kg)	7.45	10.22	6.13	6.86	8.54	1.31	3.06	9.30	12.6	8.41
SOM (g/kg)	34.5	28.2	30.5	20.1	42.3	25.9	25.1	61.4	86.5	91.4
BET (m^2/kg)	93.2	76.2	194	82.0	261.1	51.8	48.7	89.6	295.7	223.1
PS (μm)	95.4	112.	106	109	69.0	11.8	11.2	115.0	62.7	47.5
Total Al (mg/kg)	24200	36800	24400	23500	29300	21400	1800	23000	28700	54600
Total As (mg/kg)	76.8	498	146	931	99	701	536	445	130	168
Total Fe (mg/kg)	38100	45400	36800	49200	42200	129000	94400	54400	38100	38800
Total Mn (mg/kg)	796	692	888	3900	713	3710	1800	1900	909	1340
Total Pb (mg/kg)	527	13600	4060	22000	243	27000	22700	12100	3420	137

156 **Key:** pH (soil pH), conductivity (σ), CEC, SOM, BET (Weight specific surface area), PS (Mean particle size), and
 157 total concentration of Fe (Total Fe), Al (Total Al), Mn (Total Mn), and Pb (Total Pb).

158

159 2.4. Determination of Pb and As extractability

160 The Tessier 5-step sequential extraction (Tessier method) (Tessier et al., 1979) was used to determine Pb and As
161 extractability of the 10 soil samples (250 µm size fraction). The Tessier method determines five operationally defined
162 fractions: exchangeable fraction (F1), carbonate bound fraction (F2), iron and manganese bound fraction (F3),
163 organic bound fraction (F4) and residual fractions fraction (F5).

164 2.5. Heavy metal availability determined with simulated GI tract procedures

165 The soil fraction that passed through the 250 µm sieve was used for the experiments. Each soil sample was
166 subjected to two freeze-thaw treatments (0 and 12 cycles). Four widely used gastrointestinal simulation methods
167 (IVG, PBET, SBRC, UBM) were used to determine the bioavailability of Pb and As in the soils. All four methods
168 generate gastric phase and intestinal phase extracts, but they differ in solid-fluid ratio, pH and extraction time. For
169 example, pH of the gastric phase and intestinal phase of the UBM procedure are 1.2 and 6.3 respectively, and for
170 PBET are 2.5 and 7.0, respectively. Details of the gastrointestinal fluid composition and extraction parameters for the
171 four methods are provided in Table S1. The specific experimental components and experimental parameters refer to
172 the research of Juhasz et al. (2009) and Denys et al. (2012). Three replicate analyses were performed for each method.
173 Estimation of the bioavailable concentrations of Pb was as follows:

$$174 \quad C_g = \frac{\rho(G_{Pb/As}) \times V_g}{m} \quad (1)$$

$$175 \quad Bio_G = \frac{C_g}{C_s} \times 100\% \quad (2)$$

$$176 \quad C_{gi} = \frac{\rho(GI_{Pb/As}) \times V_{gi}}{m} \quad (3)$$

$$177 \quad Bio_{GI} = \frac{C_{gi}}{C_s} \times 100\% \quad (4)$$

178 where C_g is the available concentration of Pb/As in the stomach, mg/kg; C_{gi} is the available concentration of Pb/As
179 in the intestine, mg/kg; $\rho(G_{Pb/As})$ is the concentration of Pb/As in simulated gastric juice, µg/L; $\rho(GI_{Pb/As})$ is the
180 concentration of Pb/As in simulated intestinal fluid, µg/L; V_g is the volume of simulated gastric fluid, L; V_{gi} is the
181 volume of simulated intestinal fluid, L; m is the mass of the soil sample during the test, g; C_s is the total concentration
182 of Pb/As in soil samples, mg/kg; Bio_G is the human availability of Pb in the gastric stage; Bio_{GI} is the human
183 availability of Pb/As in the intestinal stage.

184 2.6. Data analysis

185 Pb and As concentrations are presented as means, standard deviations determined on triplicate analyses. ANOVA
186 was used to test the effect of freezing and thawing on Pb extractability/bioavailability versus the controls. Spearman

187 correlation was used to test the correlation between soil Pb and soil factors (soil physicochemical properties and
 188 different occurrence forms). The R statistical package was used for all statistics and graphics.

189

190 3. Results

191

192 3.1. The influence of the freeze-thaw treatments on basic soil properties

193 The freeze-thaw treatment resulted in some changes to the soil properties, compared to the untreated control soils
 194 (see Table 2 & Table S2). Freeze-thaw treatment generally resulted in the following changes: slight reduction in soil
 195 pH (typically by 0.13-0.56 pH units); a decrease in SOM (by between 4-58 %), as determined by the potassium
 196 dichromate titration method; some differences in CEC, although this was not consistently up or down following
 197 treatment. However, the biggest change was in the surface area, as determined by the BET method. This likely reflects
 198 the effect of freeze-thaw action in disaggregating/fracturing soil particles. These results show freezing-thawing has
 199 important effects on the physical and chemical properties of soils.

200

201 Table 2. Differences in the soil properties between the freeze-thaw treatments and the controls. Values in bold show
 202 a significant difference between treatment and control soils. Values with minus and plus symbols show a reduction
 203 and increase in the treated soils versus the control soils, respectively.

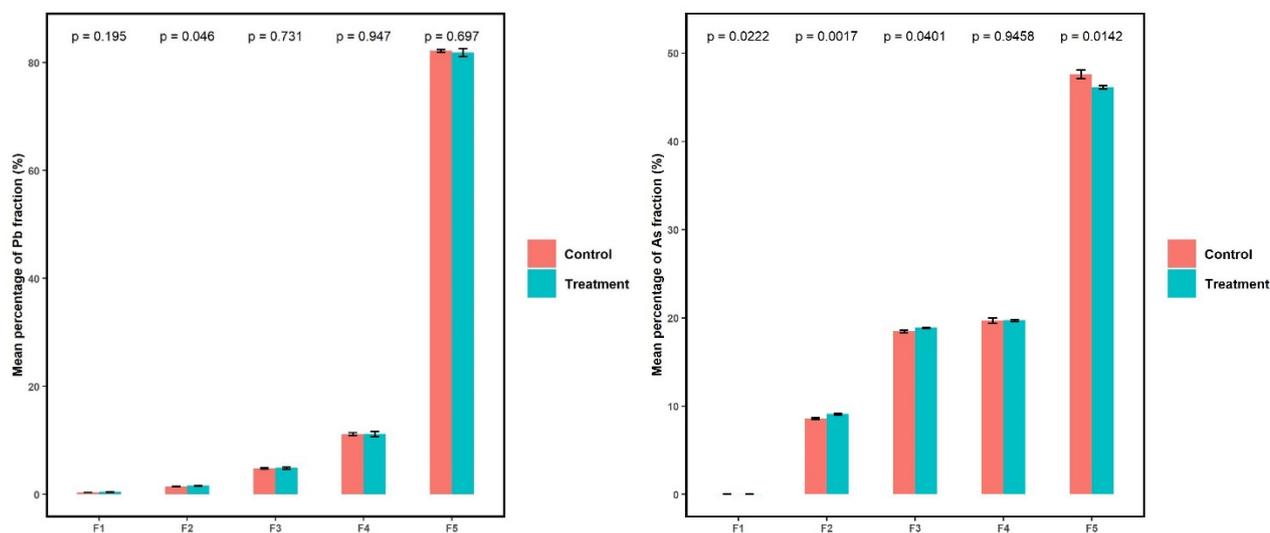
Treatment vs. Control	Soil 01	Soil 02	Soil 03	Soil 04	Soil 05	Soil 06	Soil 07	Soil 08	Soil 09	Soil 10	<i>P</i>
pH	-0.27	-0.55	-0.52	0.00	0.00	-0.13	-0.56	-0.52	-0.30	-0.41	0.0080
σ ($\mu\text{s}/\text{cm}$)	79.80	16.80	-15.50	18.30	5.70	20.00	-21.80	28.80	9.90	-66.10	0.8055
CEC (cmol/kg)	-1.87	-6.09	-1.37	0.33	-3.56	-0.73	-0.34	-0.09	-4.08	1.41	0.1430
SOM (g/kg)	-2.85	-12.31	-7.18	-6.25	-3.00	-7.75	-3.76	-0.79	-7.71	-16.62	0.2568
BET (m^2/kg)	569.54	628.35	319.39	436.19	523.60	3.49	51.41	169.00	552.75	155.17	0.0037
PS (μm)	-72.93	-90.89	-77.87	-71.96	-53.66	195.74	127.24	-35.74	-50.63	-19.07	0.0601

204 **Key:** pH (Soil pH), σ (Conductivity), CEC, SOM, BET (Weight specific surface area), PS (Mean particle size).

205

206 To analyze the effect of freeze-thaw on Pb and As speciation fractions, the Tessier sequential extraction method
 207 was used on the control soils and the freeze-thaw treated soils. When comparing the percentage of five fractions
 208 between freeze-thaw treated soils and control soils, higher F1, F2 ($P<0.05$), F3 and F4 of Pb were found under the
 209 freeze-thaw treatment than under the control, while a greater percentage of Pb occurred in F5 Pb under the control
 210 than that under the freeze-thaw treatment; Higher F1 ($P<0.05$), F2 ($P<0.05$), F3 ($P<0.05$) and F4 As was found under
 211 the freeze-thaw treatment than under the control, while higher F5 ($P<0.05$) As was found under the control than that
 212 under the freeze-thaw treatment (Figure 1 & Table S3). Although the differences were small, they were statistically

213 significant. These results for Pb and As indicated that freeze-thaw action enhanced movement from the residual
 214 fraction, organic bound fraction and Fe-Mn oxides bound fraction to the exchangeable and carbonate-bound states,
 215 (i.e. a move to the fractions typically associated with higher mobility and bioavailability).
 216

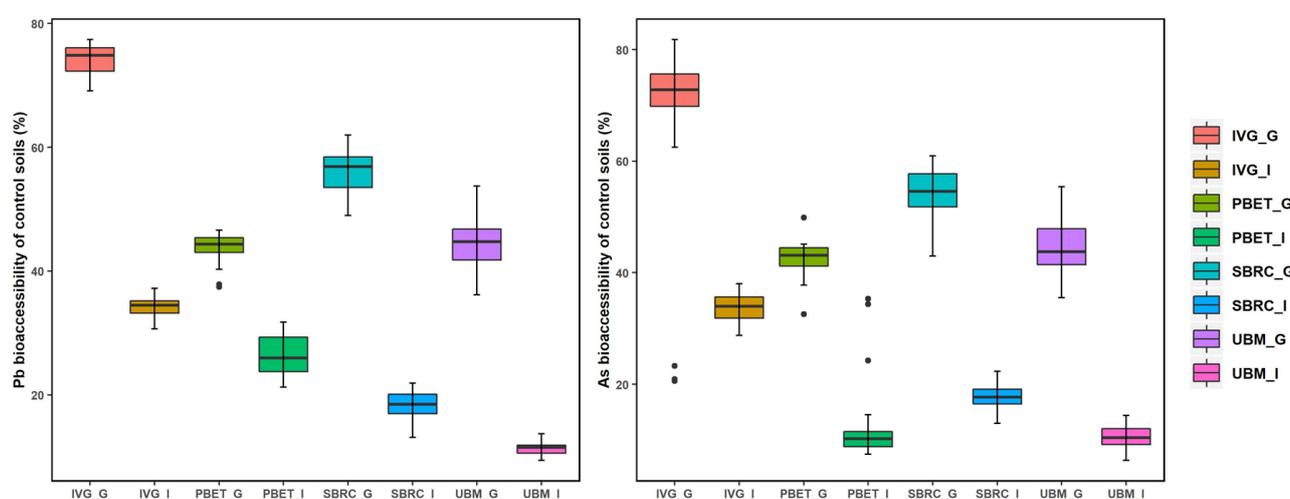


217
 218 Figure 1. Mean percentage of the five Tessier extracts of Pb (left) and As (right) under freeze-thaw treatment and
 219 control averaged for the 10 soils. F1: Exchangeable fraction; F2: Carbonate-Bound fraction; F3: Fe-Mn Oxides-
 220 Bound fraction; F4: Organic-Bound fraction; F5: Residual fraction. P values showed the significant difference of
 221 each fraction between the freeze-thaw and control treatments.

222
 223 *3.3. Pb and As bioavailability in the control (untreated) soils determined using the 4 in vitro GI methods*

224 Soil Pb and As bioavailability determined in the control soils using the 4 methods is shown in Figure 2 and Table
 225 S4. Mean values of Pb across the 10 soils range for each method as follows: IVG, 71.60±3.27-76.70±0.89 (gastric
 226 phase), 32.85±3.79-35.41±1.35 (intestinal phase); PBET, 38.68±1.79-46.00±0.70 (gastric phase), 22.96±1.48-
 227 31.09±0.60 (intestinal phase); SBRC, 49.79±1.09-58.98±0.56 (gastric phase), 16.04±1.52-20.67±0.59 (intestinal
 228 phase); UBM, 36.38±0.28-47.29±4.39 (gastric phase), 9.97±0.57-13.07±0.21 (intestinal phase). Mean values of As
 229 across the 10 soils range for each method as follows: IVG, 21.57±1.47-75.75±2.41 (gastric phase), 31.25±1.95-
 230 35.06±2.21 (intestinal phase); PBET, 36.46±3.44-44.94±4.30 (gastric phase), 8.03±0.72-31.31±6.14 (intestinal
 231 phase); SBRC, 46.88±3.44-57.53±2.21 (gastric phase), 15.62±2.26-19.03±1.89 (intestinal phase); UBM, 40.63±6.72-
 232 46.45±3.48 (gastric phase), 8.68±0.67-13.37±1.36 (intestinal phase). **Bioavailability in the control soils in the gastric
 233 and phase of the four in vitro GI methods were in the sequence: IVG > SBRC > UBM > PBET for both Pb and As;**

234 while in the intestinal phase: IVG>PBET>SBRC>UBM for Pb, IVG>SBRC>PBET>UBM for As. Both elements
 235 were lower in the intestinal phase than in the gastric phase for the control soils. From the 10 soils, Pb bioavailability
 236 followed the same order (IVG>SBRC>UBM>PBET) as the average trend in the gastric phase. The exceptions were
 237 Soils 02, 03 and 08 (IVG>SBRC>PBET>UBM). All 10 soils followed the same order (IVG>PBET>SBRC>UBM)
 238 in the intestinal phase. For As, bioavailability in 9 soils followed the same order (IVG>SBRC>UBM>PBET) in the
 239 gastric phase; the exception was Soil 08. In the intestinal phase, 9 soils also followed the same order, with the
 240 exception being Soil 05. For both Pb and As all 10 soils showed the closest agreement between the PBET and UBM
 241 GI methods (Figure S1).



242
 243 **Figure 2.** Mean Pb and As bioavailability in the control soils across 4 methods (n=10). G: Gastric phase, I: Intestinal
 244 phase.

246 3.4. Comparison of Pb bioavailability determinations between the control and freeze-thaw treated soils

247 Table 3 shows a comparison of Pb and As bioavailability derived using IVG, PBET, SBRC, and UBM between
 248 the control and the freeze-thaw treated soils. Significantly higher bioavailability of Pb and As determined by the
 249 gastric phase, the intestinal phase and the combined phases (total of G and I) of all four methods were found in freeze-
 250 thaw treated soils than in the control soils. The ratios of G:I in three of the methods (IVG, SBRC and UBM) were
 251 significantly lower in the freeze-thaw treated soils than in the control soils. Higher bioavailability of Pb and As
 252 occurred in the gastric phase, the intestinal phase and the combined phases using the IVG methodology, while the
 253 UBM and PBET methods gave the lowest values. Bioavailability of Pb and As determined by the gastric phase in the
 254 both of control soils and freeze-thaw treated soils was significant higher than that determined by the intestinal phases
 255 (Table 3). The distribution trend of As and Pb bioavailability in the ten control soils was the same as that in the ten

256 freeze-thaw treated soils. For example, the lowest bioavailability was found in Soil 05 across all four GI methods and
 257 the highest was found in Soil 08 with all four GI methods (Figure S1).

258
 259 Table 3. Comparison of Pb and As bioavailability of each method between treatments and phases averaged for the 10
 260 soils. Mean value and standard deviation of Pb and As bioavailability were provided in the table. Uppercase letters
 261 represent for the significant difference in the Pb and As bioavailability between gastric phase and intestinal phase for
 262 each method; Lowercase letters represent for the significant difference in the Pb and As bioavailability between
 263 control and treatment for each method. **G: Gastric phase; I: Intestinal phase.**

Bioacce- ssibility (%)	Control				Freeze-thaw treatment			
	Gastric phase	Intestinal phase	Ratio of G:I	Total of G and I	Gastric phase	Intestinal phase	Ratio of G:I	Total of G and I
IVG_Pb	74.15±2.40A/a	34.21±1.59B/a	2.17±0.012a	108.35±2.75a	91.96±3.09A/b	45.89±1.86B/b	2.00±0.002b	137.85±3.38b
PBET_Pb	43.71±2.41A/a	26.44±3.06B/a	1.67±0.19a	70.15±4.29a	58.51±3.22A/b	28±3.28B/a	2.11±0.24b	86.51±5.11b
SBRC_Pb	56.02±3.46A/a	18.47±2.13B/a	3.05±0.29a	74.49±3.85a	59.4±3.40A/b	21.45±2.05B/b	2.79±0.26b	80.86±4.17b
UBM_Pb	44.40±4.87A/a	11.35±1.07B/a	3.93±0.38a	55.75±3.36a	54.21±3.84A/b	14.72±1.27B/b	3.70±0.36a	68.93±4.13b
IVG_As	68.01±16.25A/a	33.88±2.48B/a	2.01±0.49a	101.89±16.21a	90.90±4.79A/b	45.45±2.92B/b	2.00±0.02a	136.35±4.85b
PBET_As	42.43±3.13A/a	12.19±6.97B/a	4.06±1.25a	54.62±7.51a	57.01±4.05A/b	12.89±7.27B/a	5.17±1.60a	69.90±8.17b
SBRC_As	54.50±4.09A/a	17.51±2.21B/a	3.12±0.21a	72.01±3.99a	57.73±4.08A/b	20.42±2.45B/b	2.83±0.19b	78.12±4.33b
UBM_As	44.41±5.13A/a	10.70±2.09B/a	4.23±0.64a	55.12±2.18a	54.20±2.64A/b	13.86±1.93B/b	3.97±0.60a	68.09±2.68b

264
 265 *3.5. Correlation of soil chemical properties and Pb and As bioavailability under the freeze-thaw treatments*

266 Spearman correlation analysis was performed on the results from the 10 soils to further analyze whether any of
 267 the measured soil properties influenced Pb and As bioavailability. A significant correlation was found between the
 268 total concentration of Al, Fe and Mn with Pb and As bioavailability under both the control and freeze-thaw treatment
 269 (see Table 4). Particle size and BET had significant correlation with Pb and As bioavailability under the control
 270 treatment. Although most pH, all SOM, all CEC and all connectivity gave no significant correlation with Pb and As
 271 bioavailability (Table 4), they showed some trends with the change of Pb and As bioavailability from the control to
 272 the freeze-thaw treatment (Table 2).

273
 274 Table 5. Correlation between Pb and As bioavailability and soil properties across different treatments. Values in the
 275 columns represent correlation coefficient between Pb and As bioavailability and soil properties. Values in bold red
 276 indicate $P < 0.05$.

Treatment	Method	Total Al		Total Fe		Total Mn		BET		CEC		pH		PS		SOM		σ	
		Pb	As	Pb	As	Pb	As	Pb	As	Pb	As	Pb	As	Pb	As	Pb	As	Pb	As
Control	IVG_G	-0.5	-0.31	0.28	0.5	0.66	0.43	-0.16	-0.48	0.1	-0.41	0.04	0.18	0.21	0.36	0.03	-0.56	-0.33	-0.37
Control	IVG_I	-0.59	-0.56	0.28	0.72	0.53	0.73	-0.22	-0.41	0.12	-0.07	0.25	0.26	0.36	0.54	-0.15	-0.09	-0.49	-0.01

Control	PBET_G	-0.42	-0.53	0.66	0.76	0.42	0.41	-0.75	-0.68	-0.07	-0.16	0.35	0.55	0.7	0.84	-0.39	-0.45	-0.04	-0.03
Control	PBET_I	-0.18	0.14	0.39	-0.16	0.15	0.2	-0.58	0.49	-0.13	-0.09	0.28	-0.02	0.52	-0.48	-0.52	-0.05	-0.31	-0.35
Control	SBRC_G	-0.47	-0.54	0.62	0.72	0.39	0.44	-0.76	-0.67	-0.1	-0.13	0.36	0.53	0.75	0.83	-0.45	-0.43	-0.09	-0.1
Control	SBRC_I	-0.7	-0.3	0.5	0.59	0.62	0.58	-0.33	-0.45	-0.13	-0.01	-0.04	-0.08	0.39	0.41	-0.21	-0.26	-0.08	0.22
Control	UBM_G	-0.31	-0.41	0.5	0.53	0.09	0.22	-0.42	-0.32	-0.14	0.26	0.42	0.47	0.38	0.56	-0.42	-0.08	-0.31	-0.02
Control	UBM_I	0.09	0.05	0.18	0.03	0.04	0.43	-0.28	0.31	0.14	-0.12	0.24	0.16	0.19	-0.3	-0.33	-0.02	-0.41	-0.47
Freeze-thaw	IVG_G	-0.54	-0.61	0.37	0.81	0.73	0.73	-0.25	-0.55	0.36	-0.08	0.19	0.07	0.39	0.65	0.13	-0.16	-0.17	-0.01
Freeze-thaw	IVG_I	-0.58	-0.64	0.31	0.82	0.77	0.68	-0.28	-0.49	0.37	-0.14	0.14	0.15	0.42	0.6	0.16	-0.19	-0.2	-0.05
Freeze-thaw	PBET_G	-0.47	-0.54	0.62	0.72	0.39	0.44	-0.45	-0.52	-0.16	-0.37	-0.13	-0.09	0.56	0.58	-0.39	-0.43	0.07	-0.13
Freeze-thaw	PBET_I	-0.21	0.14	0.37	-0.16	0.16	0.2	-0.21	0.31	-0.54	0.15	-0.2	0.68	0.25	-0.22	-0.58	0.13	-0.21	-0.44
Freeze-thaw	SBRC_G	-0.42	-0.55	0.66	0.75	0.42	0.45	-0.47	-0.54	-0.09	-0.47	-0.1	-0.04	0.58	0.6	-0.36	-0.49	0.15	-0.17
Freeze-thaw	SBRC_I	-0.7	-0.21	0.5	0.54	0.62	0.55	-0.22	-0.36	0.12	0.18	0.19	-0.18	0.39	0.45	0.02	-0.18	0.18	0.28
Freeze-thaw	UBM_G	-0.31	-0.38	0.5	0.45	0.09	0.15	-0.03	-0.18	-0.58	-0.19	0.33	-0.15	0.1	0.18	-0.36	0.09	0.01	-0.02
Freeze-thaw	UBM_I	0.04	0.05	0.14	0.03	0.02	0.43	0.15	0.1	-0.37	0.18	-0.02	0.7	-0.09	0.01	-0.49	-0.03	-0.41	-0.58

277 **Key:** pH (Soil pH), σ (conductivity), CEC, SOM, BET (Weight specific surface area), PS (Mean particle size), total
278 concentration of Fe (Total Fe), Al (Total Al), and Mn (Total Mn).

279

280 4. Discussion

281 4.1. Effect of freeze-thaw on Pb and As bioavailability

282 Our results indicated that the effect of freeze-thaw increased Pb and As bioavailability with all four in vitro
283 methods. Although previous studies (Gao et al., 2016; Sun et al., 2016; Wang, 2017) did not show the direct effect of
284 freeze-thaw on the bioavailability of heavy metals, authors have speculated that freeze-thaw will change soil
285 physicochemical properties, thereby directly affecting metal speciation and mobility, and then indirectly affecting the
286 bioavailability of soil heavy metals (Guo et al., 2012; Luo and Li, 2021; Wang et al., 2013). Metal-soil adsorption
287 and desorption will presumably be affected by freeze-thaw water content and freeze-thaw temperature (Meeravali et
288 al., 2020; Sun et al., 2019a; Yang et al., 2017). Guo et al. (2012) analyzed black soil and brown loam soil from
289 agricultural land under freeze-thaw treatments and found that the amount of Pb immobilized and adsorbed in the
290 freeze-thaw soil was lower than that in non-freeze-thaw soils. The soil samples in this study were all black soil and
291 brown loam, and the results were consistent with this study. The adsorption capacity of soil for heavy metals
292 decreased with decreasing freeze-thaw temperature, decreased with increasing freeze-thaw cycles, and first increased,
293 then decreased and then stabilized with the increase of freeze-thaw water content (Wang et al., 2011), indicating that
294 freeze-thaw may not increase Pb bioavailability in all studies/scenarios. Although only 12 freeze-thaw cycles with
295 fixed temperature and water content parameters were used for the freeze-thaw treatments in this study, the 12 freeze-
296 thaw cycles have been shown previously to be a threshold value of freeze-thaw cycles (ASTMD560, 2016).

297 The proportions of bioavailable Pb and As varied with the four methods and the two phases (gastric phase >

298 intestinal phase; $P < 0.05$, Table 3) in this study. This is consistent with other studies, which have found that more Pb
299 is solubilized in the first gastric phase (Chen, 2021) and that there are also methodological differences (Aurélié et
300 al., 2010; Li et al., 2020). Previous studies have shown that increasing soil pH can increase the adsorption of heavy
301 metal ions in soils (Harter, 1983). The availability of heavy metals in soil is significantly affected by pH, but the
302 relationship between bioavailability and pH remains complex. Yang et al. (2003) found that with the increase of soil
303 pH, the bioavailability of Pb decreased significantly. However, in this study, we found that there was no significant
304 relationship between Pb bioavailability and pH in the control soils (Table 4). This difference between our study and
305 previous studies may be due to the relatively small pH variations for the soils studied here (7.20-7.59), so that the
306 effect of pH on the bioavailability of heavy metals in the soil could not be detected.

307 In the intestinal phase under the control treatment, the highest values of Pb bioavailability were found in IVG,
308 followed by PBET, SBRC and UBM. IVG has the highest Pb bioavailability in gastric phase, PBET has the lowest
309 pH in gastric phase (1.2 versus 1.5, 1.8, 2.5) of these four methods (Table S1). This indicated that pH is the factor
310 controlling bioavailability in the gastric stage. Previous studies have indicated that the bioavailability of soil heavy
311 metals in the gastric stage of in vitro simulation methods is affected by many factors, with the pH of gastric juice an
312 important factor. For example, Oomen et al. (2002) compared the results of five in vitro digestion methods and
313 concluded that gastric pH was the most significant factor affecting the bioavailability of heavy metals in soils. In
314 addition, some studies have reported that the solid-liquid ratio is also an important influencing factor. Among the four
315 methods used in this paper, the IVG method has the highest soil-liquid ratio (1:150) in the gastric stage, and related
316 studies show that when the soil-liquid ratio increases from 1:25 to 1:1000, the bioavailability of heavy metals
317 increased.

318

319 *4.2. General consideration of soil changes and factors affecting Pb and As bioavailability under freeze-thaw*
320 *conditions*

321 The results showed soil particle size, BET weight surface area and metal concentrations (As, Pb, Al, Fe and Mn)
322 were correlated under both the freeze-thaw and control treatments, suggesting they had similar sources/behaviours.
323 The traditional soil extraction methods were not sensitive/responsive enough to show major differences in Pb
324 behaviour in the freeze-thaw experiments. There was no direct significant relationship between Pb bioavailability
325 and most soil properties (pH, SOM, and CEC), even though these variables are known to be important in affecting
326 heavy metal fate and behaviour. This may be because: (1) the limited amount of samples collected in this study; (2)

327 the pH variation between the different soils studied and with the treatments was relatively small, so that an effect of
328 pH on Pb and As bioavailability could not be detected. However, the results still indicated changes of these properties
329 under the freeze-thaw treatment (Table 2). Changes in soil physicochemical properties are important factors affecting
330 the environmental behavior of soil heavy metals, among which soil porosity, aggregate stability, pH and organic
331 matter are all important factors affecting soil bioavailability of heavy metals (Liu et al., 2017a; Zhao et al., 2010).

332 The Our results showed that soil particle size and BET weight surface area significantly correlated to the
333 bioavailability of Pb and As. Although there are no direct previous studies on freeze-thaw soils, other studies have
334 shown soil particle size is a key factor affecting the bioavailability of Pb in soil (Anthony et al., 2007). For example,
335 Ljung et al. (2007) showed that when the soil particle size was 0-50 μm , the bioavailability of heavy metals in
336 simulated gastric phase had a significant correlation with the pH of gastric phase, but when the particle size increased,
337 the correlation was not significant. In addition, our results found that lower values of soil particle size and higher
338 values of BET weight surface area were in soils under the freeze-thaw treatments than in the non-freeze-thaw
339 treatments. The physical action produced by freeze-thaw (Liu et al., 2017b; Grogan et al., 2004; Oztas and Fayetorbay,
340 2003; Zhao et al., 2004) will accelerate the fragmentation of large-particle solid media, so soil pore particle size will
341 decrease and BET weight surface area in soils will increase.

342 A total of 10 soils contaminated with Pb and As from Northeast of China were used in this study. Soils in the
343 north area of China are generally alkaline (Sun et al., 2020), which is consistent with the results of soil pH (7.20-
344 7.59) under the non-freeze-thaw treatments across all study sites. However, soil pH under the freeze-thaw treatments
345 generally decreased slightly, except for two samples). A decreasing trend was also found in previous studies (Gao et
346 al., 2016; Sun et al., 2016); this may be because freeze-thaw could promote soil nitrification and release of dissolved
347 organic acids, thereby increasing the concentration of H ions in soil and reducing soil pH (Freppaz et al., 2007; Wang
348 et al., 2007; Zhou et al., 2011). The decrease of soil solution pH reduces the negative charges carried on the soil
349 colloid surface, resulting in the decrease of soil CEC, which is basically consistent with our results of soil CEC under
350 the freeze-thaw treatments (see Table 2). In addition, some studies have reported that freeze-thaw can release easily
351 decomposed organic matter by destroying soil aggregates, but our results showed SOM decreased under the freeze-
352 thaw treatment compared to the controls. This might be because freeze-thaw can promote the mineralization of
353 organic matter by microorganisms, resulting in decreasing SOM with the freeze-thaw cycles (Wang et al., 2007).

354 To sum up, freeze-thaw action can have a comprehensive effect of soil properties which effect the adsorption and
355 desorption of heavy metals, not just one aspect of it. This study to analyze the effect of freeze-thaw action on Pb and

356 As has some limitations. First, because of the close and small size (n=10) of sampling sites, the pH, SOM and CEC
357 results may not have ranged widely enough to show a significant correlation to Pb and As bioavailability. Second,
358 different times, temperatures and water contents could have been investigated in freeze-thaw treatments in this study.
359 Larger sampling size and design of freeze-thaw experiments, perhaps also combined with in vivo experiments could
360 be conducted in future.

361

362 **5. Conclusions**

363 We investigated the effect of freeze-thaw action on the extractability and bioavailability of Pb and As in
364 contaminated soils, based on the Tessier sequential extraction method and four G-I tract simulation methods (IVG,
365 PBET, SBRC and UBM). Our results indicated that (1) the freeze-thaw action significantly increased the
366 bioavailability of Pb and As in soil; (2) the freeze-thaw action decreased the percentage of residual fraction (F5), and
367 increased the percentage of exchangeable fraction (F1), carbonate-bound fraction (F2), Fe-Mn oxides-bound fraction
368 (F3) and organic-bound fraction (F4) of Pb and As in soil; (3) Total concentrations of Al, Fe and Mn, particle size,
369 and weight surface area were the main factors correlated with soil Pb and As bioavailability. Our findings suggest
370 that the assessment of soil pollutants in frozen regions needs to comprehensively consider the effect of freeze-thaw
371 on soil properties and pollutant behaviour. In addition, due to the limitation of sampling size and design of freeze-
372 thaw treatments, further studies related to the larger sampling size and more deep design of freeze-thaw experiments
373 (e.g., temperature and water content) for in vivo experiments are required to confirm our findings and explore how
374 the freeze-thaw action affecting on the bioavailability.

375

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381

382 **Conflicts of Interest Statement**

383 The authors declare that they have no known competing financial interests or personal relationships that could
384 have appeared to influence the work reported in this paper.

385

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