# Pantropical modelling of canopy functional traits using Sentinel-2 remote sensing data

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- 55 Abstract

56 Tropical forest ecosystems are undergoing rapid transformation as a result of changing environmental 57 conditions and direct human impacts. However, we cannot adequately understand, monitor or simulate 58 tropical ecosystem responses to environmental changes without capturing the high diversity of plant 59 functional characteristics in the species-rich tropics. Failure to do so can oversimplify our understanding 60 of ecosystems responses to environmental disturbances. Innovative methods and data products are 61 needed to track changes in functional trait composition in tropical forest ecosystems through time and 62 space. This study aimed to track key functional traits by coupling Sentinel-2 derived variables with a 63 unique data set of precisely located in-situ measurements of canopy functional traits collected from 2434 64 individual trees across the tropics using a standardised methodology. The functional traits and vegetation censuses were collected from 47 field plots in the countries of Australia, Brazil, Peru, Gabon, Ghana, and 65 Malaysia, which span the four tropical continents. The spatial positions of individual trees above 10 cm 66 67 diameter at breast height (DBH) were mapped and their canopy size and shape recorded. Using geo68 located tree canopy size and shape data, community-level trait values were estimated at the same spatial 69 resolution as Sentinel-2 imagery (i.e. 10 m pixels). We then used the Geographic Random Forest (GRF) to 70 model and predict functional traits across our plots. We demonstrate that key plant functional traits can 71 be measured at a pantropical scale using the high spatial and spectral resolution of Sentinel-2 imagery in 72 conjunction with climatic and soil information. Image textural parameters were found to be key 73 components of remote sensing information for predicting functional traits across tropical forests and 74 woody savannas. Leaf thickness ( $R^2$ =0.52) obtained the highest prediction accuracy among the 75 morphological and structural traits and leaf carbon content ( $R^2 = 0.70$ ) and maximum rates of 76 photosynthesis ( $R^2 = 0.67$ ) obtained the highest prediction accuracy for leaf chemistry and photosynthesis 77 related traits, respectively. Overall, the highest prediction accuracy was obtained for leaf chemistry and 78 photosynthetic traits in comparison to morphological and structural traits. Our approach offers new 79 opportunities for mapping, monitoring and understanding biodiversity and ecosystem change in the most 80 species-rich ecosystems on Earth.

81 Keywords: Plant traits, Sentinel-2, Tropical forests, Random Forest, Pixel-level predictions, Image texture

82 1. Introduction

83 Some of the most urgent questions in ecology and ecosystem science today focus on how communities 84 of organisms respond to global environmental changes (Naeem et al., 2009), how biodiversity and 85 ecosystem changes across the world can be consistently mapped and monitored (Navarro et al., 2017), 86 and how spatial, temporal and taxonomic variability in biodiversity influences ecosystem resilience to 87 climate change (Oliver et al., 2015). In terms of Earth system science, we need to understand and model 88 how the terrestrial biosphere will respond (and already is responding) to global environmental change, 89 and whether there are critical thresholds or "tipping points" beyond which major biomes may not be able 90 to recover. Nowhere is the challenge more urgent than in the species-rich tropical forest and woody 91 savanna biomes, which together are home to more than half of global biodiversity and over 60% of 92 terrestrial productivity (Beer et al., 2010). There is evidence that atmospheric change may have effects on

tropical forest productivity and tree functional composition (Esquivel-Muelbert et al., 2019, Hubau et al.,
2020). These effects may include a stimulation of productivity (perhaps due to rising CO<sub>2</sub>) and/or a
degradation or dieback, possibly caused by increased seasonality and incurred intensity of extreme
drought events (Malhi et al., 2008, Malhi et al., 2018). Such events are partly responsible for the increased
tree mortality and decreased carbon residence time in tropical forests worldwide (McDowell et al., 2018).
However, to adequately understand such responses we need to capture and map the high diversity of
plant ecosystem function in the species-rich tropics and savannas.

100 Species functional traits are defined as the morphological, physiological or phenological attributes 101 which determine the fitness of organisms, their response to changes in the environment and their 102 influence on ecosystem functions (Kissling et al., 2018, Díaz & Cabido, 2001). Functional traits provide 103 tangible and mechanistic means of assessing the ability of communities to adapt to climate change 104 (Pacifici et al., 2015) and play a major role in determining ecosystem productivity, functioning and notably 105 nature's contribution to people (e.g. water and wood availability) (Díaz et al., 2019, Carmona et al., 2016). 106 Any tools or methods that facilitate quantification of functional traits across large spatial scales and at 107 high spatial resolution would be invaluable for quantifying ecosystem functioning and ecological 108 responses to disturbance at scales relevant for policy and management (Kissling et al., 2018). However, it is still challenging to map functional trait diversity in tropical regions given the lack of plant trait data 109 110 available for most of those locations (Jetz et al., 2016). Additional challenges come from different and 111 often incompatible trait collection protocols and the lack of systematic high spatial, spectral and temporal 112 resolution remote sensing imagery that coincides with data for functional traits at the canopy level and 113 the lack of geo-located tree stems at the plot level. Thus, there is a need for spatially-explicit methods to 114 map and quantify plant functional traits at high spatial resolution in tropical forest and woody savanna 115 ecosystems.

116 Tracking functional traits can shed light on differences in ecosystem functioning across broad 117 spatial extents and therefore aid policy and decision making, e.g. for creating adequate biodiversity 118 conservation policies or for providing early warning of directional shifts in ecosystems. The key challenges

119 of any functional trait approach are scalability and monitoring: how can functional shifts in highly diverse 120 tropical forests and woody savannas be monitored and tracked over large spatial extents? Intensive field 121 sampling of plant functional traits at a pantropical scale is time-consuming and economically unviable. 122 There are large gaps in the availability of plant trait data globally, and the largest gaps are in the tropics 123 (Jetz et al., 2016). Large plant trait datasets aim to overcome this issue and have advanced our ability to 124 carry out plant functional trait analysis in an unprecedented way (Kattge et al., 2020, Gallagher et al., 125 2020). However, as with any database, the plant trait values from such databases will represent the local 126 trait-environment relationships for the site where they were collected, which may not be the area of 127 interest. A key assumption in trait-based ecology is that the environment is filtering for an optimal set of 128 trait characteristics so that the resulting communities are adapted to the environment where they are 129 distributed (Fell & Ogle, 2018, Lebrija-Trejos et al., 2010, Lortie et al., 2004). Hence, we might expect an 130 optimal set of trait characteristics for a given location, which when analysed over time could quantify the 131 dynamics of community trait distributions or shifts in functional composition relating to environmental 132 changes (Enquist et al. 2015).

133 Recently, there has been an increasing investment into mapping plant functional trait 134 distributions given economic and data availability constraints such efforts have mostly focused on 135 hyperspectral imagery at local (Schneider et al., 2017) to regional scales (Asner et al., 2015, Asner et al., 136 2016). However, high resolution hyperspectral imagery is not widely available (Clark, 2017, Szabó et al., 137 2019). Landsat-8 imagery at coarser spatial (30 m pixel), spectral and temporal resolution than Sentinel-2 138 imagery has been used to map four traits over small  $(20 \times 20 \text{ m})$  vegetation plots covering small spatial 139 extents (Wallis et al. 2019). The spatial mismatch between site-level trait sampling and the spatial 140 resolution of pixels may partly affect overall model predictions (Wallis et al. 2019). Other studies restricted to European forests (Ma et al., 2019) show how Sentinel-2 imagery could be used to map functional trait 141 142 diversity in the comparatively low tree diversity forests of Europe (Ma et al., 2019) and to retrieve specific 143 leaf area from Landsat-8 imagery (Ali et al., 2017). However, the tropics present a different set of 144 challenges, such as the high species richness, low accessibility and comparatively low availability of trait

data, plus the low coverage of remote sensing data because of persistent high cloud cover. These
challenges have hampered developments in mapping plant functional trait distributions across most
tropical areas.

148 Satellite imagery with high spectral, spatial and temporal resolution is particularly needed in the 149 wet tropics (Asner et al., 2017), where clear days can be infrequent and several images may be required 150 to construct a cloud-free composite. The Copernicus mission from the European Space Agency's (ESA; 151 www.esa.int) aids in the improvement in this area. The Sentinel-2 multispectral imager satellites are part 152 of the Copernicus programme, which has the potential to provide new opportunities to evaluate canopy 153 traits remotely. Sentinel-2 has 13 spectral channels covering the visible, near-infrared, and short-wave 154 infrared, a spatial resolution of 10 m for visible and near-infrared, 20 m for short-wave infrared, revisit 155 period of 5 days and it provides open data availability. The improved spectral sampling (13 bands, 10 156 excluding the 60 m atmospheric bands) and fine spatial resolution of the Sentinel-2 images have the 157 potential to elucidate leaf chemistry, morphology, photosynthesis and water content at the pixel-level, 158 although this remains largely untested. Multispectral sensors do not provide the rich information available 159 from hyperspectral sensors, which have been used in numerous studies to map functional traits at small 160 spatial extents (Townsend et al., 2003, Laurin et al., 2016, Asner et al., 2015, Martin, M. E. et al., 2008). 161 However, high resolution open-access hyperspectral imagery is not currently available from space. 162 Although Landsat images have been used to predict a few functional traits at a local scale (Wallis et al., 163 2019), the extended spectral, spatial and temporal capabilities of the state-of-the-art sensors onboard the 164 Sentinel-2 satellites provide greater potential for mapping functional trait diversity in tropical forest 165 ecosystems at large extents.

Here, we employ a unique and large dataset of *in-situ* plant canopy traits and vegetation census data collected with a standardised protocol at multiple sites across the tropics to calibrate and validate Sentinel-2 imagery for predicting community leaf trait composition. The data provide 14 standardized measurements of *in-situ* collected plant functional traits, precisely geo-located and delineated individual tree crowns and vegetation censuses from Australia, South East Asia, Africa and South America to model

171 and predict functional trait composition at the pixel-level. We investigate how functional traits of tropical 172 forests vary within and between these different tropical regions and whether Sentinel-2 spectral data in 173 conjunction with climatic and soil information provide sufficient information to predict such pixel-level 174 trait composition in long-term vegetation plots across the tropics. We hypothesised that there would be 175 differences in trait variation among sites and regions given the range of climatic and soil conditions across 176 the tropics. Given the high spectral and spatial resolution of Sentinel-2 imagery we further hypothesised 177 that raw spectral bands and textural information will prove to be key predictors of functional trait 178 distributions across the tropics. The very high spatial resolution and local origin of the input plant traits 179 and census dataset, which represent traits adapted to local environments, plus the use of the Sentinel-2 180 data will allow us to accurately predict plant functional trait distributions that are potentially generalisable 181 across the tropical forest biome.

#### 182 **2. Methods**

#### 183 2.1 Vegetation plots

184 We collected vegetation census data from 47 permanent vegetation plots that are part of the Global 185 Ecosystems Monitoring network (GEM; www.gem.tropicalforests.ox.ac.uk). These plots encompass wet 186 tropical forests, seasonally dry tropical forests, and tropical forest-savanna transitional vegetation. The 187 sampled vegetation plots have an area ranging from 0.1 to 1 ha, with most (61%) being 1 ha. The plots 188 used are located across four tropical continents and specifically in the countries of Australia, Brazil, Gabon, 189 Ghana, Malaysian Borneo (from here onwards referred to as Malaysia) and Peru (Table 1). In each plot all 190 woody plant individuals with a diameter  $\geq$  10 cm at breast height (DBH) or above buttress roots were 191 measured and their exact geographic location was recorded (see the 'Individual tree crowns' section 192 below for more details). In two plots (NXV-01 and NXV-10) in Nova Xavantina, here onwards referred to 193 as Brazil-NX, the DBH was measured near ground level as is standard in savanna monitoring protocols.

194 2.2 Functional traits

195 We collected plant functional trait measurements from all woody plants located in each of the 47 196 vegetation plots mentioned above (Table 2). All traits were gathered from the GEM network and were 197 collected following a standardized methodology across plots. Forest inventory data were used to stratify 198 tree species by basal area dominance, a proxy for canopy area dominance. The tree species that 199 contributed most to basal area abundance were sampled with 3-5 replicate individuals per species, with 200 a goal of sampling 60-80% of basal area across the sampling region. Eighty percent of basal area was often 201 achieved in low diversity sites (e.g. montane or dry forests) but only around 60% was achieved in some 202 high diversity sites (lowland humid rainforests). For each selected tree a sun and a shade branch were 203 sampled and in each branch 3-5 leaves were used for trait measurements. We only included the sun 204 exposed branches in our analysis because we were interested in the branches that could potentially be 205 receiving direct sun radiation and thus show direct spectral reflectance. This represented a total sample 206 of 2434 individual trees across the tropics (Table 1). The plant functional traits collected were those 207 related to photosynthetic capacity at both saturating  $CO_2$  concentration (2000 ppm  $CO_2$ ; A<sub>max</sub>) and 208 ambient CO<sub>2</sub> concentration (400 ppm CO<sub>2</sub>; A<sub>sat</sub>); leaf chemistry (nitrogen, phosphorus, carbon, calcium, 209 potassium and magnesium content); and leaf morphological and structural traits (area, specific leaf area, 210 thickness, dry mass, fresh mass and water content). An overview of the methods for individual leaf 211 functional trait measurements is provided in the Supplementary Information (see full traits collection 212 protocol section). Further details of measurements for the Peruvian Andes campaign are given in Martin 213 et al. (2020) and Enquist et al. (2017), for the Malaysian campaign in Both et al. (2019), and for the Ghana 214 and Brazil campaigns in Oliveras et al. (2020) and Gvozdevaite et al. (2018).

Some individuals in the plots lacked functional trait values. To assign representative trait values to unsampled individuals we did the following: 1) individuals from which traits were measured kept their original trait information, 2) for individuals with no trait information we randomly sampled trait values from other individuals from the same species present in the same plot, 3) if the species was not sampled in the given plot then we randomly sampled an individual from the same species that had trait information in other plots from the same region (Table 1). This protocol for trait value allocation allowed us to work with the existing range of trait values at the species level and avoided to create average values per species
(Cadotte et al., 2011, Schneider et al., 2017). We did not assign trait values to the remaining individuals
belonging to species from which no trait collection was obtained at the regional level.

224 2.3 Individual tree crowns

225 Tree crown locations and structural attributes were recorded for each tree, where crown area and shape 226 were measured by direct crown field measurements in the case of plots in Malaysia and Peru (see protocol 227 below), or by means of regional level allometric equations developed by Shenkin et al. (2019) (all other 228 plots). In the latter case, the crown's shape was assumed to be circular. The direct field crown 229 measurements were as follows: all trees  $\geq$  10 cm DBH (i.e., 1.3 m from the ground) were mapped using a 230 ground-based Field-Map laser technology (IFER, Ltd., Jílové u Prahy, Czech Republic) (Hédl et al., 2009). 231 The Field-Map technology was based on a combination of Impulse 200 Standard laser rangefinder (with 232 in-built tilt sensor for measuring vertical angles), MapStar module II electronic compass (both Laser 233 Technology Inc., Colorado, USA), and the specialized mapping software Field-Map v. 11 (IFER, Czech 234 Republic). The technology was used to record spatial positions of tree stems in three-dimensional space 235 (x, y, z-coordinates) as well as to map individual horizontal projections of tree crowns in the plots. The 236 horizontal crown projection of every tree was obtained by measuring spatial positions (x and y-237 coordinates) of series of points (ranging from 5 to 30 points depending on the size of the crown) at the 238 boundary of a crown projected to the horizontal plane. The shape of crown projection was subsequently 239 smoothed using the "smooth contour line" feature of Field-Map software v. 11. Heights of all trees with 240 DBH ≥ 10 cm were measured by the Impulse and TruPulse 360 R laser rangefinders (both Laser Technology 241 Inc., Colorado, USA). Thus, each individual crown was accurately geolocated rendering information about 242 its shape and vertical and horizontal position.

243 2.4 Calculating pixel-level trait composition

We calculated the community weighted mean of each trait for each 10 × 10 m subplot (matching the highest pixel spatial resolution of the Sentinel-2 imagery) based on the mass ratio hypothesis, which

states that the most dominant species drive the ecosystem processes by means of their functional traits (Grime, 1998). We first geolocated the vegetation plot, with its already mapped tree crowns (see protocol above), to the Sentinel-2 imagery based on the corner coordinates of the plots. This is an important step as geolocation errors between the vegetation plot and the correct location in the satellite image could represent a large proportion of a given plot depending on the plots' area. Then for each of the traits, *t*, and pixels, *p*, we calculated their community level weighted mean (CWM) using the individual tree crown horizontal area as the weighting factor (Fig. 1) as follows:

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$$CWM_{tp} = \frac{\sum_{i=1}^{N} CA_{ip} \times t_{ip}}{CA_{p}}$$

254 (Eq. 1)

Where  $CA_{ip}$  is the crown area of individual *i* in pixel *p*,  $t_{ip}$  is the trait value of individual *i* in pixel *p*, *N* is the total number of individuals per pixel and  $CA_p$  is the crown area of pixel *p*. The crown contribution to the CWM was weighted by its proportional cover of the corresponding pixel. The total number of pixels used in our calculations are 403 for Australia, 449 for Brazil -NX (Nova Xavantina), 302 for Brazil -ST (Santarem), 464 for Gabon, 620 for Ghana, 976 for Malaysia and 1280 for Peru.



261 Figure 1. Diagram summarising the steps followed to assign trait values per Sentinel-2 pixel. 1) First the 262 vegetation plots are defined based on the GEM (Global Environmental Monitoring) dataset and 2) from 263 each vegetation plot the corner coordinates are extracted. 3) From each vegetation plot the XY position 264 of each stem  $\geq 10$  cm DBH is extracted and 4) the crown horizontal area is calculated based on the protocol 265 described in the methods section. 5) Then the Sentinel-2 imagery for the study area is processed to level 266 2A using the ESA SNAP toolbox and 6) the vegetation plot is overlaid in the Sentinel-2 image based on its 267 corner coordinates. In this last step (6) each pixel defines a 'subplot' which is the unit used to calculate 268 the trait community weighted mean based on the crown area of the trees that are contained by that pixel. 269 In 6) n refers to a given tree in a given pixel, trait i represents a given trait and x and y are values for that 270 trait. The image used as an example in step (1) was taken by Jesus Aguirre-Gutierrez over a vegetation 271 plot using a multispectral ALTUM camera mounted on an Inspire 1 drone."

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#### 273 2.5 Sentinel-2 data, vegetation indices and canopy texture parameters

274 We gathered Sentinel-2 imagery that was closest in time and season to the sampling dates of functional 275 traits and vegetation census across the tropics for each of the study locations (Table S1). The Sentinel-2 276 imagery was first selected using the European Space Agency (ESA) ScienceHub (scihub.copernicus.eu) 277 choosing images with high pixel quality and low cloud cover (<10%). Atmospheric, radiometric and 278 topographic corrections were applied to the selected imagery (Level 1C) using the Sen2Cor algorithm in 279 the Sentinel SNAP toolbox (step.esa.int). Our overlapping imagery with the vegetation plots appeared free 280 of clouds and cirrus effects. The above-mentioned steps allowed us to obtain level 2A imagery with surface 281 reflectance values. We then resampled the 20 m bands to 10 m spatial resolution using bilinear 282 interpolation. The Sentinel-2 60 m resolution bands (B01, B09, B10) were not used as these are designed 283 for cirrus, water vapour and cloud detection (Table 3). Band 8A was not used as it covers an overlapping 284 spectral window with band 8 and has a lower spatial resolution. Since vegetation indices may increase 285 prediction accuracy when modelling community weighted traits (Wallis et al., 2019), we calculated three 286 of them (Table 3) which we hypothesised to inform trait distributions given their association with 287 chlorophyll and nutrient levels in the leaves and their use of the visible-to-red edge spectral bands.

Canopy structure may play an important role in separating different vegetation types and differences in canopy spectral composition. To characterize canopy structure, we calculated the Grey Level Co-Occurrence Matrix (GLCM) based texture features (Haralick et al., 1973). The desired texture metrics are computed from a grey tone matrix that is spatially dependent. The co-occurrence matrix 292 depends on the angular relationship and distance between two neighbouring pixels and depicts the 293 number of occurrences of the relationship between a pixel and its neighbour. After trials with smaller 294 windows size (5 x 5) we opted to use a 9 x 9 pixel kernel window which was sufficient to render enough 295 canopy contrast information during the modelling step (see section 2.7 below) without taking large 296 periods of time for its calculation. The texture results obtained with the used kernel window was highly 297 correlated to the smaller kernel window (Cor = 0.94, P = < 0.0001). Based on the GLCM we calculated two 298 variables that are least correlated with each other, the Entropy and Correlation, for each of the vegetation 299 indices. While Entropy measures the homogeneity level for a given area, the Correlation measures 300 probability of occurrence of the specified pixel pairs across the image (Haralick et al., 1973, Wallis et al., 301 2019). All remote sensing analyses related to the generation of vegetation indices and texture metrics 302 were carried out using the Sentinel SNAP toolbox (step.esa.int) and the R statistical environment (R 303 Development Core Team, 2014) with the 'Sen2R' package.

#### 304 2.6 Environmental and soil data

305 Climatic, topographic and soil characteristics may vary across regions and could at least partly determine 306 the region's vegetation and intrinsic trait composition. We obtained information on these three 307 components for each sampling location. The three components were grouped as belonging to 308 environmental (climate) or soil-terrain (texture, pH, cation exchange capacity and topography) drivers 309 (Table 3).

310 For climate and for each sampling location we gathered gridded data on the mean annual climatic 311 water deficit (MCWD), which is a metric of drought intensity and severity, mean annual maximum 312 temperature (MATmax), solar radiation (SRAD) and soil moisture (SM) (Table 3). All climatic data with a spatial resolution of  $\sim$ 4km were obtained from the TerraClimate gridded climate product (Abatzoglou et 313 314 al., 2018). To characterise the climatic conditions for each location we used a climatology of 30 years 315 (1986-2015) suggested by the World Meteorological Organization (WMO; as 316 www.wmo.int/pages/prog/wcp/ccl/faqs.php). We used the terrain slope to characterise the plot's

317 topography, as it has been shown that topography may shape the composition and structure of tropical 318 forests (Jucker et al., 2018) and may affect the vegetation spectral reflectance by modifying soil water and 319 nutrient availability. Terrain slope was calculated using a high-resolution digital elevation model, ~30m 320 pixel size at the equator, from the Shuttle Topography Mission (Farr et al., 2007). At most sites soil data 321 were sampled locally, and analysed to a standardised protocol in labs in either INPA, Manaus, Brazil or the 322 University of Leeds, UK, following the RAINFOR soil protocol (Quesada et al., 2012). From these data we 323 summarised plot level soil data averaged over the first 30 cm for texture (Sand% and Clay%), cation 324 exchange capacity (eCEC) and pH-H<sub>2</sub>O (pH). Plot level texture data were not available for plots in Australia 325 and the NXV-10 plots and were thus derived from the SoilGrids dataset at 250m pixel spatial resolution 326 for those plots only (Hengl et al., 2017).

#### 327 2.7 Comparing community level trait distributions across regions

We tested if and to what extent the community-level trait distributions differed among regions. We square-root transformed the trait value to improve normality and applied an analysis of variance (ANOVA). We then applied a Tukey's Honest Significant Difference (Tukey HSD) test to investigate the significance of the differences between the means of the community weighted mean (CWM) trait values among locations. The ANOVA and Tukey test were carried out using the 'stats' package for R (R Development Core Team, 2014).

334 2.7 Relating pixel-level trait composition to spectral reflectance, environment and soil conditions

We modelled the community weighted mean (CWM) of each trait at the pixel-level (10×10 m) as a function of the Sentinel-2 remote sensing, environmental and soil covariates (Table 3) using a 'spatial' version of the machine learning Random Forest (RF) algorithm (Breiman, 2001) named Geographic Random Forests (GRF) (Georganos et al., 2019). RF is a nonparametric algorithm that has been shown to be robust to overfitting and variable inputs thanks to the bagging process and its random feature selection (Hastie et al., 2009). Moreover, it has been extensively used to model and predict ecological and remote sensing 341 data within and across ecosystems (e.g. Asner et al., 2016, Van der Plas et al., 2018). In contrast to RF, 342 GRF disaggregates the underlying data in geographic space, in this case based on the spatial coordinates 343 of the Sentinel-2 pixels, building global and local sub-models (plot level), making the modelling framework 344 thus spatially explicit. The explicit inclusion of the spatial component (XY pixel location) in the models, 345 which are sequentially fitted with different sets of the training data (the bagging process) may contribute 346 to the observed reduced spatial autocorrelation of GRF in comparison to the common RF (Georganos et 347 al. 2019). In the GRF a global model is built as in other RF applications. However, GRF also generates a 348 local RF for each location, which includes a specified number of nearby observations, here defined by all 349 pixels in the vegetation plot (mostly 1 ha; Table 1), called 'neighbourhood', obtaining in this way metrics 350 of local and global model predictive power and variable importance. For model predictions, a fusion 351 between the global model (that uses more data) and local models (with low bias) can be applied, 352 weighting the contributions of the global and local models based on the parameters that increase the 353 predictive accuracy and decrease the model's Root Mean Square Error (RMSE). We used the spatial GRF 354 to fit a global model for each functional trait and also fit a specific model for each region (Australia, Brazil 355 -ST, Brazil -NX, Gabon, Ghana, Malaysia and Peru) using the SpatialML package in R.

356 We performed an extensive set of model optimization and regularization procedures to reduce 357 over-fitting. For the CWM models we selected the number of trees to fit by 10-fold cross-validation 358 analysis with number of trees ranging between 500 and 1500 and the number of variables randomly 359 sampled as candidates at each split (mtry) ranging between 1 to 10, using in the final model the 360 combination of terms that generated the lowest RMSE. All covariates included in the models had pairwise 361 Pearson correlation coefficients  $r \le 0.82$  (Table 3). For the final global and local models, we used 80% of 362 the data for model fitting and the remaining 20% for model evaluation. Variable importance for each model was computed as the decrease in node impurities from splitting on the variable, averaged over all 363 364 trees and derived from the Out of Bag (OOB) error. Then the resulting importance was standardised to a 365 0–1 scale for comparison purposes.

We carried out all analyses stated above with the full set of tree individuals present in each vegetation plot with functional traits assuming that the contribution of small individuals to the trait CWM value, and thus to the community reflectance at the pixel-level, would be minimal given the weighting factor used (i.e. the individual's crown area). However, to underpin this we carried out all analyses on two smaller datasets, one where the 25<sup>th</sup> and other where the 50<sup>th</sup> percentile of the smallest trees per region were removed. All analyses were carried out in the R statistical environment with the 'caret', 'tidyverse' and 'SpatialML' packages.

373 **3. Results** 

374 3.1 Variation in trait composition across tropical forests

Most leaf functional traits exhibited significant differences across the tropics (Fig. 2) including wide trait range variation within the same region (Fig. S8), with leaf fresh mass and leaf thickness being on average less variable among locations (Table S2).





**Figure 2. Comparison of trait distributions across tropical regions.** The boxplots are based on the pixellevel (10×10 m) community trait values for each trait and region (*n*=403 for Australia, 449 for Brazil-NX, 302 for Brazil-ST, 464 for Gabon, 620 for Ghana, 976 for Malaysia and 1280 for Peru). Horizontal lines in each boxplot show the median value and vertical lines are the whiskers that extend to the largest value or not further than 1.5 times the inter-quartile range. For some locations information for all traits was not available. For full details in significant differences in mean trait values among locations see Table S4. Brazil -NX: Nova Xavantina; Brazil -ST: Santarem.

386

Leaf chemistry and photosynthetic capacity (A<sub>max</sub> and A<sub>sat</sub>) often showed significant differences

- 387 among locations (Table S2). Drier locations as in Nova Xavantina (Brazil -NX) displayed trait adaptations
- to seasonal rainfall and temperature with on average thicker and smaller (30 ± 0.05 mm and 56.2 ± 24.7
- 389 cm<sup>2</sup> respectively) leaves at the community level, with some of the highest community-level leaf nitrogen
- 390 concentration (2.2  $\pm$  0.3 %) and highest photosynthetic capacity (mean A<sub>max</sub>= 21.9  $\pm$  4.3  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, and
- 391  $A_{sat}$  = 8.3 ± 2.5 µmol m<sup>-2</sup> s<sup>-1</sup>). In contrast, wetter regions such as Malaysia displayed on average some of

the biggest (113.5 ± 55 cm<sup>2</sup>) and thinnest (0.25 ± 0.05 mm) leaves with high leaf water content (59.1 ± 5 %). The Peruvian altitudinal transect showed large variation in community-level traits values, which often overlapped with trait values from all other sampled locations across the tropics (Fig. 2). For most nutrients, leaf nutrient concentration was often highest in forests found in Ghana (e.g. K% = 0.97 ± 0.27 and Mg% = 0.33 ± 0.1) and Malaysia (K% = 1.05 ± 0.27 and Mg% = 0.27 ± 0.1). Australian forests showed on average some of the lowest community-level N (1.3 ± 0.21 %) and P (0.07 ± 0.01%) leaf concentrations.

#### 398 3.2 Pantropical and local community level functional trait models

The analyses carried out with the full dataset and the dataset where the 25<sup>th</sup> and 50<sup>th</sup> percentile of the smallest trees per region were removed gave similar results for the global ( $R^2 = 0.95$  and  $R^2 = 0.97$ respectively; Table S3) and local ( $R^2 = 0.81$  and  $R^2 = 0.80$  respectively; Table S4) models of plant trait distributions. Therefore, in the following we only present the results for the models carried out with the full vegetation dataset.

404 The accuracy of the pantropical prediction of functional traits ranged between a minimum of  $R^2$  = 0.26, for leaf fresh mass, and a maximum of  $R^2 = 0.70$  for leaf carbon content (C%) based on the out-of-405 406 sampled (testing) data across the tropics (Table 4). The predictive accuracies of leaf chemistry and 407 photosynthetic traits were often higher than for morphological and structural traits such as leaf dry mass  $(R^2 = 0.27)$  and leaf area  $(R^2 = 0.43)$  (Fig. 3). At the pantropical level, the highest prediction accuracy was 408 409 obtained for leaf thickness ( $R^2 = 0.52$ ) for morphological and structural traits, for leaf Ca (Ca%;  $R^2 = 0.64$ ) 410 and leaf K (K%;  $R^2$  = 0.63) for the chemical traits other than carbon. Leaf N and P concentrations were also 411 predicted with high accuracy ( $R^2$  = 0.59). Leaf photosynthetic capacity traits, A<sub>max</sub> and A<sub>sat</sub>, showed some 412 of the highest prediction accuracies ranging from  $R^2 = 0.55$  to 0.67, respectively. Model spatial predictions 413 for several traits and locations are shown in Fig. 4 and others can be seen in Fig. S1-Fig. S7.

414 Models built for each tropical region and trait uncovered marked differences in prediction 415 accuracy among them (Fig. 5; Table 5 and Table S5). Leaf area prediction accuracy ranged from  $R^2 = 0.04$ 416 (Brazil -ST) to 0.35 (Australia), and that of specific leaf area (SLA) ranged from  $R^2 = 0.06$  for Malaysia to 417 0.54 for Brazil -NX (Table S5). The local models showed a higher accuracy for predicting local level leaf chemical nutrients (up to  $R^2 = 0.68$ ), especially for P, Ca, and N concentrations in comparison to 418 419 morphological (e.g. leaf area and SLA) traits (Table 5; Fig. 5). Traits related to photosynthetic capacity 420 showed an overall better prediction accuracy than leaf area and SLA with prediction values ranging 421 between 0.36 (Peru) to 0.49 (Ghana) for A<sub>max</sub> and up to 0.52 for A<sub>sat</sub> (Brazil -NX; Fig. 5). On average the 422 highest prediction accuracy across regions for a given trait were reached for leaf P concentration ( $R^2$  = 423 0.47) and  $A_{max}$  ( $R^2 = 0.44$ ) and the locations with the highest average prediction accuracy across traits were 424 the Nova Xavantina savanna (Brazil -NX,  $R^2$ =0.40) and the Peru elevation gradient ( $R^2$  = 0.38; Table 5), both 425 sites encompassing strong gradients in vegetation morphology and structure.



Figure 3. Model predictions to the 20% test data from the general model which was fitted with 80% of the trait data from across the tropics. Grey dots are the observed against predicted trait values of the pixel-level (10 × 10 m) community weighted mean traits from the test dataset. The black line shows the 1:1 relationship between observed and predicted values. Model prediction accuracy is shown in the top left. Full model results are shown in Table 4.



432

### 433 Figure 4. Spatial predictions of trait distributions for a selected subset of plant traits and locations.

- 434 The map (middle) shows the locations of vegetation plots that were used during the modelling
- 435 framework. The spatial predictions (top and bottom rows) were obtained using the general models
- 436 (Table 4) for each of the traits and locations at a 10 × 10 m pixel resolution. The approximated location
- 437 of each vegetation plot used is shown as a white square within each spatial prediction map (for
- 438 visualisation purposes white squares are not scaled to the plot's real size). Spatial predictions for other
- 439 traits can be found in Fig. S1-S7.



440

Figure 5. Models predictions to the 20% test data from the regional models fitted with 80% of the trait data from each region across the tropics. Each colour represents an individual regional model and the coloured symbols are the observed against predicted trait values of the pixel-level (10 × 10 m) community weighted mean traits from the test dataset. The black line shows the 1:1 relationship between observed and predicted values. Model prediction accuracy is shown in Table 5. Full model results are shown in Table

446 S5. Brazil -NX: Nova Xavantina; Brazil -ST: Santarem.

447 3.3 Importance of spectral remote sensing, climatic and soil data for mapping trait distributions

448 We included Sentinel-2 band derived reflectance values, vegetation indices, their canopy texture 449 parameters, climatic and soil variables in the general trait models to predict community level traits at the 450 pixel-level (Table 3). The importance of these variables for predicting traits depended on the specific trait 451 being addressed (Fig. 6). In the global model, the remote sensing texture parameters were the first or 452 second major contributor for predicting nine of the functional traits across the tropics (Fig. 6 and Fig. S9). 453 Raw spectral variables were the second most important group for predicting four of such functional traits 454 but often lower in importance than the textural parameters. In the global model, soil and terrain factors were on average some of the most important for predicting photosynthetic traits and foliar P 455 456 concentration. On average, climatic variables were important for predicting 11 out of 14 functional traits 457 but their contribution was lower for predicting leaf dry and fresh mass and leaf water content (Fig. 6). 458 However, it is evident that a combination of textural, spectral, climatic and soil information is required to 459 obtain the best general model predictions across functional traits and no single variable appears as the 460 most important across all traits (Fig. S9).

461 The local models provided a site-specific view of the most important remote sensing derived 462 variables, environmental and soil conditions for deriving community level traits composition (Fig. S10). 463 Sentinel-2 remote sensing related variables were more important for detecting leaf morphology and 464 nutrient values than environmental and soil related variables in 88% of the trait models (in 75 out of 85 465 possible traits by region combinations). Eighty-one percent of the time (69 location by trait combinations) 466 the canopy texture parameters were more important than the raw spectral reflectance factors. In 5.9% 467 and 4.7% of the possible trait and region combinations, climatic or soil-topography related variables 468 respectively were the most important for detecting community traits (Fig. S10).



469

470 Figure 6. Group median variable importance of spectral remote sensing, environmental and soil 471 related variables for determining functional trait predictions in the global model. Variable importance 472 (Y axis) ranges from 0 (no importance) to 1 (highest importance) and represents the decrease in node 473 impurities from splitting on the variable, averaged over all trees and derived from the Out of Bag error, 474 the resulting value has been standardised to a 0-1 scale for comparison purposes. The spectral group (S2 475 -Spectral) contains the select raw bands from the Sentinel-2 and the vegetation indices; Texture 476 parameters (S2 -Texture) contain the Correlation and Entropy metrics from the grey level co-occurrence 477 matrix obtained from the vegetation indices; Climate contains all climatic variables; Soil-Terrain contains 478 all soil characteristics and slope. All variables are described in Table

- 479 4. Discussion
- To the best of our knowledge, this is the first study evaluating the ability of Sentinel-2 satellite data to
- 481 map plant functional traits across tropical ecosystems. Tropical forest trait mapping is fundamental for
- 482 understanding of plant responses to global change, and notably the plant functional traits we predict in
- this study are relevant to plant species responses to a changing environment (Both et al., 2019, Nunes et

al., 2019, Soudzilovskaia et al., 2013, Aguirre-Gutiérrez et al., 2019). We have demonstrated that accurate
pixel-level (10×10 m) predictions of tropical forest functional trait distributions across the tropics can be
generated by making use of extensive *in-situ* collected plant functional traits, geo-located canopy
structure, vegetation censuses and high spectral and spatial resolution remote sensing data from the
Sentinel-2 satellites.

489 4.1 Tropical forest trait distributions

490 Plant functional traits are characteristics that aid species to thrive in their environment or adapt to new 491 conditions. Given such adaptations to specific environments it might be expected that trait variation 492 would be higher in regions that encompass more varied environmental conditions (Enquist et al., 2015). 493 Environmental adaptation is exemplified by the strong variation in values for most traits in Peru and Malaysia. In Peru, the data represent a climatic and altitudinal gradient ranging from the lowland Amazon 494 495 in the Tambopata National Park at an elevation of 200-225 masl to plots in Acjanaco at above 3000 masl. 496 In Malaysia, the vegetation plots are distributed across a land-use gradient ranging from undisturbed to 497 heavily logged forests (Both et al., 2019). Environmental adaptation may be also shown by the observed 498 differences in trait distributions between different regions across the tropics (e.g. Australia and Gabon; 499 see also Fig. 2). The pixel-based community trait values in the Peruvian transect often extend across much 500 of the range in trait values observed in other locations (Fig. 2). We detected an overall significant 501 difference among locations in terms of morphological, chemical and photosynthetic traits (Table S2). This 502 wide variation in traits suggests the presence of local biotic and abiotic controls of trait distributions and 503 plant species adaptations that may differ among tropical regions. Such differences in trait composition 504 highlight the importance and the challenge of sampling as fully as possible the functional trait diversity 505 across different tropical forests. This is of pivotal importance when comparing forest responses to 506 changing environments across multiple regions. We thus suggest that further field trait survey campaigns 507 across the tropics are needed to improve pantropical trait predictions. As for the local biotic and abiotic 508 controls of trait distributions, for instance, it is widely known that African tropical forests are in general 509 less species diverse than their Asian and South American counterparts but that they have some of the

510 highest biomass carbon storage capacity per unit area (Sullivan et al., 2017). Tropical forests in West Africa 511 are in general drier in comparison to Amazonian tropical forests (Parmentier et al., 2007) and some African 512 regions such as Gabon have experienced increases in temperature and decreases in precipitation over the 513 last 30 years (Bush et al., 2020). Thus, such changes in climatic conditions as those observed in West 514 African tropical forest may also underlie variations in species composition and the locally observed 515 functional trait pool as shown in this study. It is also worth noting that two caveats of the community-516 weighted mean trait approach may account for part of the unexplained trait variation. First, it makes the 517 assumption of a unique functional optimum in a given environment, while multiple optimal strategies – 518 potentially corresponding to contrasting trait values – could coexist (Laughlin et al., 2018). Secondly, it 519 does not account for the dynamic nature of communities, so that a community weighted mean at a given 520 time point might not encompass the optimum at equilibrium (Laughlin et al., 2018).

521 Morphological and structural traits such as leaf area, fresh and dry mass, leaf thickness, SLA and 522 LWC, represent trade-offs between energy acquisition, consumption and survival and form a main part of 523 the global spectrum of plant functioning (Díaz et al., 2016). Besides investigating the predictability of such 524 plant structural traits, we further analysed the potential for predicting leaf chemistry (C, K, Mg, Ca, N, P) 525 and photosynthesis related traits (Amax and Asat). Mapping chemical and photosynthetic traits at a 526 pantropical scale has the potential for increasing our understanding of how photosynthetic capacity shifts 527 across tropical regions and on possible impacts of a changing environment on tropical forests productivity 528 (Guan et al., 2015, Mueller et al., 2014).

529 4.2 Sentinel-2 remote sensing for mapping community level trait distributions across the tropics

In their pioneering work with hyperspectral imagery and simulated multispectral Sentinel-2 data over Ghana, Laurin et al. (2016) demonstrated that Sentinel-2 imagery could be used to discriminate tropical forest types and map plant functional types. The authors argued that the full band set and vegetation indices derived from the Sentinel-2 would be advantageous for accurately mapping plant functional guilds in the tropics. By using functional trait data collected *in situ* across tropical forests and modelling at high 535 spatial resolution (pixel-level) we show that most of our global trait distribution models present a high 536 predictive power for most traits analysed, with prediction accuracy on the testing datasets being highest 537 for predicting leaf chemical and photosynthetic capacity traits. However, we also show that the local level 538 trait models produced less accurate predictions than the global models, probably as a result of the 539 narrower range of in plant trait values found at the local region in comparison to across the regions, 540 something also shown by Wallis et al. (2019). The prediction accuracy obtained from our models using 541 Sentinel-2 multispectral data is similar and in some cases higher than that shown by recent studies that 542 make use of hyperspectral imagery and other multispectral sensors to map functional traits (Martin et al., 543 2018, Asner et al., 2017, Asner et al., 2015). For instance our predictions on test data for leaf nitrogen, 544 phosphorus and carbon are comparable or higher than those obtained by other innovative studies in Malaysia ( $R^2 = 0.46$ , 0.44 and 0.48 respectively; Martin et al., 2018), Peru ( $R^2 = 0.48$ , 0.39 and 0.44; Asner 545 546 et al., 2015) and temperate forests ( $R^2$  = 0.55, 0.22, 0.46; Nunes et al., 2017), and closely related to those 547 obtained by Wallis et al. (2019) with other multispectral sensor for nitrogen and phosphorus ( $R^2 = 0.65$ and 0.65). Specially the work of Asner et al. (2017) has shown how such plant trait predictions (with its 548 549 inherent accuracies) can be used for other applications such as to guide biodiversity conservation actions. 550 In our approach we resample the 20m spatial resolution bands from the Sentinel-2 to 10m pixels as to 551 work with the highest spatial resolution available for most spectral bands. Such resampling could in 552 principle have an effect on the match between the tree canopies' reflectance signal and the spectral signal 553 from the Sentinel-2 pixel and could thus influence the textural parameters, by for instance, detecting 554 lower heterogeneity.

Some of the leaf chemistry we modelled can be directly related to the reflectance obtained from the Sentinel-2 remote sensor in the visible, infrared and red-edge regions which capture the leaf biogeochemistry (Ustin & Gamon, 2010). For instance, it has been shown how carbon and carbon containing metabolites peak in reflectance at around 550 nm (band 3 in the Senitnel-2) and at the lower part of the 702–715 nm (Ely et al., 2019), which would be depicted best by the red-edge band 5 in the Sentinel-2. such spectral behaviour captured by the Sentinel-2 bands contributed to the high prediction

561 accuracy of leaf carbon in our study. Our models show how Sentinel-2 imagery, and especially the canopy 562 texture parameters derived from it, can be especially useful for mapping traits related to leaf chemistry 563 (Fig. 2 and Fig. S9). Moreover, our high predictive accuracy for photosynthetic capacity ( $A_{max}$ ,  $A_{sat}$ ) is 564 consistent with studies carried out in other vegetation types (e.g. agroecosystems; Serbin et al., 2015) 565 where a strong association was shown between photosynthesis related traits and the red-edge spectral 566 region. Sentinel-2 has 3 bands over the red-edge spectral region (bands 5, 6, 7) and two over the near 567 infrared (bands 8 and 8a) with different bandwidths, which as shown by Shiklomanov et al. (2016) can be 568 advantageous for detecting foliar nutrients such as leaf N (Schlemmer et al., 2013), as small differences 569 in wavelength position in different bands may impact their capacity to retrieve canopy trait 570 characteristics. Moreover, the strong relationship between photosynthetic capacity and spectral 571 reflectance can be partly captured from the leaf N signal, as leaf N concentrations are strongly associated 572 with photosynthetic capacity (Reich, 2012, Vincent, 2001). The N reflectance signal is often best obtained 573 in wavebands centred between 440 and 570 nm (Ferwerda et al., 2005).

574 In this study we leverage evidence on covariation among traits to estimate and predict values of 575 traits that have no clear physical effects on spectral reflectance. There is ample evidence of the existence 576 of covariation among plant traits, as for instance between leaf N concentration, specific leaf area and leaf 577 longevity (Walker et al., 2017). Such covariation among traits may in principle also represent covariation 578 in the spectral reflectance patterns across vegetation types (Ma et al., 2019), especially if such individuals 579 vary in leaf structural tissue that drive energy scattering and reflectance (Ollinger, 2011). Such covariation 580 between traits can be helpful for mapping functional trait diversity across large spatial extents that include 581 diverse vegetation types (Townsend et al., 2003, Both et al., 2019). We show that the spectral reflectance, 582 image textural parameters (Entropy and Correlation), climate and soil, are highly relevant for modelling plant trait distributions across the tropics with high prediction accuracy. However, the canopy texture 583 584 parameters (Entropy and Correlation) are some of the most important for attaining high trait prediction 585 accuracies across plant functional traits (Sarker & Nichol, 2011, Wallis et al., 2019) and differences in

spectral, climatic and soil conditions between different regions are key components for improving model
 predictions across broad spatial extents.

588 Image texture parameters were derived from the vegetation indices that we calculated, which in 589 turn were derived from the raw spectral bands of the Sentinel-2. Thus, the texture metrics besides taking 590 advantage of the high spectral resolution of the sensor also take advantage of its high spatial resolution. 591 Although the raw spectral bands of the Sentinel-2 were not as important for predicting some functional 592 traits as image texture, it is relevant to consider that texture values tend to differ based on the spatial 593 resolution of the underlying data on which they are based. A larger pixel (e.g. Landsat's 30 × 30 m pixels) 594 may thus mask differences in the landscape that could in principle be captured by the Sentinel 10 × 10 m 595 resolution texture generated metrics. This therefore highlights the relevance of Sentinel-2 imagery for 596 functional plant functional trait predictions in comparison to others with lower spectral and spatial 597 resolution. Image texture parameters can help characterise the upper surface of the vegetation, which in 598 our study is composed of varied sets of functional trait characteristics that confer them different spectral 599 responses. When such spectral differences are analysed with grey level co-occurrence matrices, the 600 generated image texture parameters (e.g. entropy and correlation) can help differentiate contrasting 601 vegetation in the landscape. The role of texture parameters for modelling biomass and functional traits has also been recognised by other studies focusing not only on mapping functional traits along elevation 602 603 gradients but also for estimating standing biomass (Wallis et al., 2019). Moreover, such relevance of 604 texture parameters does not seem to be limited to the spatial resolution of the Sentinel-2 imagery as 605 shown when using high spatial resolution SPOT imagery for modelling forest aboveground biomass 606 (Hlatshwayo et al., 2019) and WorldView-3 for tree species identification (Ferreira et al., 2019), or lower 607 spatial resolution data as that from the Landsat (Wallis et al., 2019). Other added value of the Sentinel-2 in contrast to finer spatial resolution satellites (e.g. SPOT and WorldView-3) is its high revisit period, to 608 609 obtain cloud free imagery, and it's free availability. Moreover, soil properties can be informative when 610 modelling trait distributions across regions in the tropics as they partly drive the plant functional and 611 species compositional turnover (Prada et al., 2017, Asner et al., 2016). In our study different vegetation

plots appeared to be on soils with different parent materials resulting in varying cation exchange capacity,
 pH and soil texture, and thus including differences between sites contributes to increasing the prediction
 accuracy of trait distributions.

615 Although in the past it was thought not to be possible to map individual plant species or functional 616 traits (Price, 1994, Ustin & Gamon, 2010), the advent of remotely sensed data with high spectral, spatial 617 and temporal resolution has made it possible to extract information on the chemical and structural 618 composition of forest canopies even in highly biodiverse tropical forests. This has been demonstrated with 619 the use of hyperspectral sensors (Asner et al., 2017, Asner et al., 2015, Jetz et al., 2016), which often 620 collect hundreds of spectral bands at very high spatial and spectral resolutions but at relatively small 621 spatial extents and often without temporal replication. More research is needed to disentangle to what 622 extent hyperspectral data offers more information to that offered by the Sentinel-2 sensors for an 623 increased mapping accuracy of functional traits of tropical forests. As shown by Laurin et al. (Laurin et al., 624 2016), results obtained with simulated Sentinel-2 data are highly comparable to those obtained from 625 hyperspectral imagery for mapping forest types, dominant tree species and functional guilds. Being able 626 to monitor functional traits at high spatial and temporal resolution with multispectral data ranging from 627 the visible to the shortwave infrared across the tropics and with freely available data opens new 628 opportunities for understanding the effects of environmental changes on biodiversity at a local scale. This 629 is because functional traits play a major role in determining ecosystem productivity and functioning, e.g. 630 carbon capture (Díaz et al., 2019, Carmona et al., 2016). Moreover, spatially explicit models of functional 631 traits shift across the tropics can help decipher how ecosystem functioning varies even among tropical 632 areas, providing a cost-effective pathway to identifying regions of high conservation value and hence aid 633 in the creation of locally adequate biodiversity conservation policies. Overall, our findings are of relevance for informing biodiversity monitoring policies under ecosystem change as we show that accurate 634 635 predictions of relevant plant functional traits can be obtained in high biodiversity areas such as the tropics. 636 Our approach thus facilitates tracking possible shifts in trait distributions and composition across large 637 spatial extents as a response to environmental changes using the Sentinel-2 satellites.

#### 638 **5. Conclusions**

639 Tropical forest ecosystems are witnessing a rapid transformation as a result of changing environmental 640 conditions and direct human impacts (Lewis et al., 2015, Taubert et al., 2018, Aguirre-Gutiérrez et al., 641 2019). However, we cannot adequately understand or simulate tropical ecosystem responses to 642 environmental changes based solely on current ecosystem model approaches as these are unable to 643 capture the high diversity of plant ecosystem functions in the species-rich tropics. Neglect of functional 644 biodiversity can oversimplify the simulated response of an ecosystem to an environmental disturbance. 645 Here we show the high variation in functional traits that exists among tropical regions, which hints at the 646 different capabilities of such forests to respond to a changing environment. We demonstrate 647 opportunities for measuring the distribution of key functional traits across tropical forest ecosystems at 648 the pixel-level using the Sentinel-2 satellites, which if done across time could reveal areas where 649 functional shifts have occurred and likely where biodiversity conservation/amelioration measures are 650 needed. Although the Sentinel-2 satellites show high promise for this endeavour, our approach is limited 651 by the short time interval since they were launched (i.e. 2015) and the lower spectral resolution of 652 Sentinel-2 imagery in comparison to that derived from hyperspectral sensors. Methods and data products 653 are needed to track changes in functional composition in forest ecosystems across time and space. We 654 demonstrate a new approach to develop a rapid monitoring tool for capturing the effects of a changing 655 environment across the tropics. This new tool has the potential to contribute to a more robust and 656 evidence-based policy-making for conservation of tropical forest ecosystems.

#### 657 Authorship contribution statement

J.A.G. conceived the study, designed and carried out the analysis and wrote the first draft of the paper.
Y.M. conceived and implemented the GEM Network, obtained funding for most of the GEM traits field
campaigns and commented on earlier versions of the manuscript. S.R. advised on statistical and remote
sensing analysis and commented on earlier versions of the manuscript. All co-authors participated in or
coordinated vegetation, trait data and/or soil data collection or processed field data. The authors named

663 between S.A.B. and L.J.T.W. are listed alphabetically. All co-authors commented on and approved the 664 manuscript.

#### 665 **Declaration of competing interest**

The authors declare that they have no known competing financial interests or personal relationships thatcould have appeared to influence the work reported in this paper.

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#### References

- Abatzoglou, J.T., Dobrowski, S.Z., Parks, S.A., & Hegewisch, K.C. (2018). TerraClimate, a high-resolution
  global dataset of monthly climate and climatic water balance from 1958–2015. *Scientific data*, *5*, 170191
- Aguirre-Gutiérrez, J., Oliveras, I., Rifai, S., Fauset, S., Adu-Bredu, S., Affum-Baffoe, K., et al. (2019). Drier
- tropical forests are susceptible to functional changes in response to a long-term drought. *Ecology Letters, 22,* 855-865
- Ali, A.M., Darvishzadeh, R., & Skidmore, A.K. (2017). Retrieval of specific leaf area from landsat-8 surface
   reflectance data using statistical and physical models. *IEEE Journal of selected topics in applied earth observations and remote sensing*, *10*, 3529-3536
- Asner, G.P., Martin, R.E., Anderson, C.B., & Knapp, D.E. (2015). Quantifying forest canopy traits: Imaging
   spectroscopy versus field survey. *Remote Sensing of Environment, 158*, 15-27
- Asner, G.P., Knapp, D.E., Anderson, C.B., Martin, R.E., & Vaughn, N. (2016). Large-scale climatic and
   geophysical controls on the leaf economics spectrum. *Proceedings of the National Academy of Sciences*,
   *113*, E4043-E4051
- Asner, G.P., Martin, R.E., Knapp, D.E., Tupayachi, R., Anderson, C.B., Sinca, F., et al. (2017). Airborne
  laser-guided imaging spectroscopy to map forest trait diversity and guide conservation. *Science*, *355*,
  385-389
- 710 Barnes, E.M., Clarke, T.R., Richards, S.E., Colaizzi, P.D., Haberland, J., Kostrzewski, M., et al. (2000).
- Coincident detection of crop water stress, nitrogen status and canopy density using ground based
   multispectral data., 1619
- Beer, C., Reichstein, M., Tomelleri, E., Ciais, P., Jung, M., Carvalhais, N., et al. (2010). Terrestrial gross
  carbon dioxide uptake: global distribution and covariation with climate. *Science*, *329*, 834-838
- Both, S., Riutta, T., Paine, C.T., Elias, D.M., Cruz, R.S., Jain, A., et al. (2019). Logging and soil nutrients
  independently explain plant trait expression in tropical forests. *New Phytologist, 221*, 1853-1865
- 717 Breiman, L. (2001). Random forests. *Machine Learning*, 45, 5-32
- Bush, E.R., Jeffery, K., Bunnefeld, N., Tutin, C., Musgrave, R., Moussavou, G., et al. (2020). Rare ground
  data confirm significant warming and drying in western equatorial Africa. *PeerJ*, *8*, e8732
- Cadotte, M.W., Carscadden, K., & Mirotchnick, N. (2011). Beyond species: functional diversity and the
   maintenance of ecological processes and services. *Journal of Applied Ecology, 48*, 1079-1087
- 722 Carmona, C.P., de Bello, F., Mason, N.W., & Lepš, J. (2016). Traits without borders: integrating functional
- 723 diversity across scales. *Trends in ecology & evolution, 31, 382-394*

- Chave, J., Muller-Landau, H.C., Baker, T.R., Easdale, T.A., Steege, H.t., & Webb, C.O. (2006). Regional and
   phylogenetic variation of wood density across 2456 neotropical tree species. *Ecological Applications, 16*,
- 726 2356-2367
- 727 Clark, M.L. (2017). Comparison of simulated hyperspectral HyspIRI and multispectral Landsat 8 and
- Sentinel-2 imagery for multi-seasonal, regional land-cover mapping. *Remote Sensing of Environment*,
   200, 311-325
- 730 Daughtry, C., Walthall, C.L., Kim, M.S., De Colstoun, E.B., & McMurtrey Iii, J.E. (2000). Estimating corn
- reaf chlorophyll concentration from leaf and canopy reflectance. *Remote Sensing of Environment, 74,*229-239
- Díaz, S., Kattge, J., Cornelissen, J.H., Wright, I.J., Lavorel, S., Dray, S., et al. (2016). The global spectrum of
   plant form and function. *Nature*, *529*, 167-171
- Díaz, S., Settele, J., Brondízio, E., Ngo, H.T., Guèze, M., Agard, J., et al. (2019). Summary for policymakers
- of the global assessment report on biodiversity and ecosystem services of the Intergovernmental
   Science-Policy Platform on Biodiversity and Ecosystem Services.
- 738 <u>https://www.ipbes.net/sites/default/files/downloads/spm\_unedited\_advance\_for\_posting\_htn.pdf</u>, 739 ADVANCE UNEDITED VERSION –
- Díaz, S. & Cabido, M. (2001). Vive la différence: plant functional diversity matters to ecosystem
   processes. *Trends in ecology & evolution, 16*, 646-655
- 742 Ely, K.S., Burnett, A.C., Lieberman-Cribbin, W., Serbin, S.P., & Rogers, A. (2019). Spectroscopy can predict
- key leaf traits associated with source–sink balance and carbon–nitrogen status. *Journal of experimental botany, 70,* 1789-1799
- Find Enduist, B.J., Norberg, J., Bonser, S.P., Violle, C., Webb, C.T., Henderson, A., et al. (2015). Scaling from
  traits to ecosystems: developing a general trait driver theory via integrating trait-based and metabolic
  scaling theories. In Anonymous *Advances in ecological research* (pp. 249-318). : Elsevier
- Enquist, B.J., Bentley, L.P., Shenkin, A., Maitner, B., Savage, V., Michaletz, S., et al. (2017). Assessing
  trait-based scaling theory in tropical forests spanning a broad temperature gradient. *Global Ecology and Biogeography, 26*, 1357-1373
- Esquivel-Muelbert, A., Baker, T.R., Dexter, K.G., Lewis, S.L., Brienen, R.J., Feldpausch, T.R., et al. (2019).
  Compositional response of Amazon forests to climate change. *Global Change Biology, 25*, 39-56
- Farr, T.G., Rosen, P.A., Caro, E., Crippen, R., Duren, R., Hensley, S., et al. (2007). The shuttle radar
  topography mission. *Reviews of Geophysics*, 45
- Fell, M., & Ogle, K. (2018). Refinement of a theoretical trait space for North American trees via
  environmental filtering. *Ecological Monographs*, *88*, 372-384
- 757 Ferreira, M.P., Wagner, F.H., Aragão, L.E., Shimabukuro, Y.E., & de Souza Filho, Carlos Roberto. (2019).
- 758 Tree species classification in tropical forests using visible to shortwave infrared WorldView-3 images and
- texture analysis. *ISPRS journal of photogrammetry and remote sensing, 149*, 119-131
- Ferwerda, J.G., Skidmore, A.K., & Mutanga, O. (2005). Nitrogen detection with hyperspectral normalized
   ratio indices across multiple plant species. *International Journal of Remote Sensing, 26*, 4083-4095
- 762 Gallagher, R.V., Falster, D.S., Maitner, B.S., Salguero-Gómez, R., Vandvik, V., Pearse, W.D., et al. (2020).
- 763 Open Science principles for accelerating trait-based science across the Tree of Life. *Nature ecology* &
- 764 evolution, 4, 294-303

- 765 Georganos, S., Grippa, T., Gadiaga, A.N., Linard, C., Lennert, M., Vanhuysse, S., et al. (2019).
- Geographical Random Forests: A Spatial Extension of the Random Forest Algorithm to Address Spatial
   Heterogeneity in Remote Sensing and Population Modelling. *Geocarto International*, 1-12
- Grime, J.P. (1998). Benefits of plant diversity to ecosystems: immediate, filter and founder effects.
   *Journal of Ecology, 86*, 902-910
- Guan, K., Pan, M., Li, H., Wolf, A., Wu, J., Medvigy, D., et al. (2015). Photosynthetic seasonality of global
   tropical forests constrained by hydroclimate. *Nature Geoscience*, *8*, 284
- 772 Gvozdevaite, A., Oliveras, I., Domingues, T.F., Peprah, T., Boakye, M., Afriyie, L., et al. (2018). Leaf-level
- photosynthetic capacity dynamics in relation to soil and foliar nutrients along forest–savanna
- boundaries in Ghana and Brazil. *Tree physiology, 38*, 1912-1925
- Haralick, R.M., Shanmugam, K., & Dinstein, I.H. (1973). Textural features for image classification. *IEEE transactions on systems, man, and cybernetics*, 610-621
- Hastie, T., Tibshirani, R., & Friedman, J. (2009). *The elements of statistical learning, 2nd edn New York.*
- 778 Hawthorne, W.D. (1995). Ecological profiles of Ghanaian forest trees. Tropical forestry papers
- Hédl, R., Svátek, M., Dančák, M., Rodzay, A.W., Salleh, A.B., & Kamariah, A.S. (2009). A new technique
- for inventory of permanent plots in tropical forests: a case study from lowland dipterocarp forest in
- Kuala Belalong, Brunei Darussalam. *Blumea-Biodiversity, Evolution and Biogeography of Plants, 54*, 124130
- Hengl, T., de Jesus, J.M., Heuvelink, G.B., Gonzalez, M.R., Kilibarda, M., Blagotić, A., et al. (2017).
  SoilGrids250m: Global gridded soil information based on machine learning. *PLoS one, 12*, e0169748
- Hlatshwayo, S.T., Mutanga, O., Lottering, R.T., Kiala, Z., & Ismail, R. (2019). Mapping forest aboveground
- biomass in the reforested Buffelsdraai landfill site using texture combinations computed from SPOT-6
   pan-sharpened imagery. *International Journal of Applied Earth Observation and Geoinformation, 74*, 65 77
- Huang, W., Ratkowsky, D.A., Hui, C., Wang, P., Su, J., & Shi, P. (2019). Leaf fresh weight versus dry
  weight: which is better for describing the scaling relationship between leaf biomass and leaf area for
- 791 broad-leaved plants? *Forests, 10,* 256
- Hubau, W., Lewis, S.L., Phillips, O.L., Affum-Baffoe, K., Beeckman, H., Cuní-Sanchez, A., et al. (2020).
  Asynchronous carbon sink saturation in African and Amazonian tropical forests. *Nature*, *579*, 80-87
- Jetz, W., Cavender-Bares, J., Pavlick, R., Schimel, D., Davis, F.W., Asner, G.P., et al. (2016). Monitoring
   plant functional diversity from space. *Nature Plants*, *2*, 16024
- Jucker, T., Bongalov, B., Burslem, D.F., Nilus, R., Dalponte, M., Lewis, S.L., et al. (2018). Topography
  shapes the structure, composition and function of tropical forest landscapes. *Ecology Letters, 21*, 9891000
- Juneau, K.J., & Tarasoff, C.S. (2012). Leaf area and water content changes after permanent and
   temporary storage. *PLoS One*, *7*, e42604
- Kattge, J., Bönisch, G., Díaz, S., Lavorel, S., Prentice, I.C., Leadley, P., et al. (2020). TRY plant trait
   database–enhanced coverage and open access. *Global Change Biology*
- Kissling, W.D., Walls, R., Bowser, A., Jones, M.O., Kattge, J., Agosti, D., et al. (2018). Towards global data products of Essential Biodiversity Variables on species traits. *Nature ecology & evolution, 2*, 1531-1540

- Laughlin, D.C., Strahan, R.T., Adler, P.B., & Moore, M.M. (2018). Survival rates indicate that correlations between community-weighted mean traits and environments can be unreliable estimates of the
- adaptive value of traits. *Ecology Letters, 21*, 411-421
- Laurin, G.V., Puletti, N., Hawthorne, W., Liesenberg, V., Corona, P., Papale, D., et al. (2016).
- 809 Discrimination of tropical forest types, dominant species, and mapping of functional guilds by
- 810 hyperspectral and simulated multispectral Sentinel-2 data. *Remote Sensing of Environment, 176,* 163-
- 811 176
- Lebrija-Trejos, E., Pérez-García, E.A., Meave, J.A., Bongers, F., & Poorter, L. (2010). Functional traits and environmental filtering drive community assembly in a species-rich tropical system. *Ecology*, *91*, 386-398
- Lewis, S.L., Edwards, D.P., & Galbraith, D. (2015). Increasing human dominance of tropical forests. *Science*, *349*, 827-832
- Lortie, C.J., Brooker, R.W., Choler, P., Kikvidze, Z., Michalet, R., Pugnaire, F.I., et al. (2004). Rethinking
  plant community theory. *Oikos*, *107*, 433-438
- 818 Ma, X., Mahecha, M.D., Migliavacca, M., van der Plas, F., Benavides, R., Ratcliffe, S., et al. (2019).
- 819 Inferring plant functional diversity from space: the potential of Sentinel-2. *Remote Sensing of* 820 *Environment, 233,* 111368
- Malhi, Y., Rowland, L., Aragao, L. E. O. C., & Fisher, R.A. (2018). New insights into the variability of the

tropical land carbon cycle from the El Nino of 2015/2016. *Philosophical transactions of the Royal Society* 

- of London.Series B, Biological sciences, 373, 10.1098/rstb.2017.0298
- Malhi, Y., Roberts, J.T., Betts, R.A., Killeen, T.J., Li, W., & Nobre, C.A. (2008). Climate change,
  deforestation, and the fate of the Amazon. *Science*, *319*, 169-172
- 826 Martin, M.E., Plourde, L.C., Ollinger, S.V., Smith, M., & McNeil, B.E. (2008). A generalizable method for
- remote sensing of canopy nitrogen across a wide range of forest ecosystems. *Remote Sensing of Environment, 112*, 3511-3519
- 829 Martin, R.E., Chadwick, K.D., Brodrick, P.G., Carranza-Jimenez, L., Vaughn, N.R., & Asner, G.P. (2018). An
- approach for foliar trait retrieval from airborne imaging spectroscopy of tropical forests. *Remote Sensing*, 10, 199
- Martin, R.E., Asner, G.P., Bentley, L.P., Shenkin, A., Salinas, N., Huaypar, K.Q., et al. (2020). Covariance of Sun and Shade Leaf Traits Along a Tropical Forest Elevation Gradient. *Frontiers in plant science, 10*, 1810
- McDowell, N., Allen, C.D., Anderson-Teixeira, K., Brando, P., Brienen, R., Chambers, J., et al. (2018).
  Drivers and mechanisms of tree mortality in moist tropical forests. *New Phytologist, 219*, 851-869
- Mueller, T., Dressler, G., Tucker, C., Pinzon, J., Leimgruber, P., Dubayah, R., et al. (2014). Human land-use
   practices lead to global long-term increases in photosynthetic capacity. *Remote Sensing*, *6*, 5717-5731
- 838 Naeem, S., Bunker, D.E., Hector, A., Loreau, M., & Perrings, C. (2009). Biodiversity, Ecosystem
- 839 Functioning, and Human Wellbeing: An Ecological and Economic Perspective. (pp. 388). : Oxford
- 840 University Press
- 841 Navarro, L.M., Fernández, N., Guerra, C., Guralnick, R., Kissling, W.D., Londoño, M.C., et al. (2017).
- Monitoring biodiversity change through effective global coordination. *Current opinion in environmental sustainability, 29*, 158-169
- Nunes, M., Davey, M., & Coomes, D. (2017). On the challenges of using field spectroscopy to measurethe impact of soil type on leaf traits.

- 846 Nunes, M.H., Both, S., Bongalov, B., Brelsford, C., Khoury, S., Burslem, D.F., et al. (2019). Changes in leaf
- functional traits of rainforest canopy trees associated with an El Niño event in Borneo. *Environmental*
- 848 *Research Letters, 14, 085005*
- 849 Oliver, T.H., Heard, M.S., Isaac, N.J., Roy, D.B., Procter, D., Eigenbrod, F., et al. (2015). Biodiversity and 850 resilience of ecosystem functions. *Trends in Ecology & Evolution, 30*, 673-684
- 851 Oliveras, I., Bentley, L., Fyllas, N.M., Gvozdevaite, A., Shenkin, A.F., Prepah, T., et al. (2020). The
- influence of taxonomy and environment on leaf trait variation along tropical abiotic gradients. *Frontiers in Forests and Global Change, 3,* 18
- Ollinger, S.V. (2011). Sources of variability in canopy reflectance and the convergent properties of plants. *New Phytologist, 189*, 375-394
- Pacifici, M., Foden, W.B., Visconti, P., Watson, J.E., Butchart, S.H., Kovacs, K.M., et al. (2015). Assessing
  species vulnerability to climate change. *Nature Climate Change*, *5*, 215-224
- Parmentier, I., Malhi, Y., Senterre, B., Whittaker, R.J., ATDN, Alonso, A., et al. (2007). The odd man out?
  Might climate explain the lower tree α-diversity of African rain forests relative to Amazonian rain
  forests? *Journal of Ecology*, *95*, 1058-1071
- Prada, C.M., Morris, A., Andersen, K.M., Turner, B.L., Caballero, P., & Dalling, J.W. (2017). Soils and
- rainfall drive landscape-scale changes in the diversity and functional composition of tree communities in
   premontane tropical forest. *Journal of vegetation science, 28*, 859-870
- Price, J.C. (1994). How unique are spectral signatures? *Remote Sensing of Environment, 49*, 181-186
- Qi, J., Chehbouni, A., Huete, A.R., Kerr, Y.H., & Sorooshian, S. (1994). A modified soil adjusted vegetation
  index. *Remote Sensing of Environment*, 48, 119-126
- 867 Quesada, C.A., Phillips, O.L., Schwarz, M., Czimczik, C.I., Baker, T.R., Patiño, S., et al. (2012). Basin-wide
- variations in Amazon forest structure and function are mediated by both soils and climate.*Biogeosciences, 9*
- 870 R Development Core Team. (2014). R: A language and environment for statistical computing. R
- foundation for statistical computing, Vienna, Austria. ISBN 3-900051-07-0, URL <u>http://www.R-</u>
   project.org.
- Reich, P.B. (2012). Key canopy traits drive forest productivity. *Proceedings of the Royal Society B: Biological Sciences, 279,* 2128-2134
- Sarker, L.R., & Nichol, J.E. (2011). Improved forest biomass estimates using ALOS AVNIR-2 texture
  indices. *Remote Sensing of Environment*, *115*, 968-977
- 877 Schlemmer, M., Gitelson, A., Schepers, J., Ferguson, R., Peng, Y., Shanahan, J., et al. (2013). Remote
- estimation of nitrogen and chlorophyll contents in maize at leaf and canopy levels. *International Journal* of Applied Earth Observation and Geoinformation, 25, 47-54
- Schneider, F.D., Morsdorf, F., Schmid, B., Petchey, O.L., Hueni, A., Schimel, D.S., et al. (2017). Mapping
   functional diversity from remotely sensed morphological and physiological forest traits. *Nature*
- 882 *communications, 8,* 1441
- Serbin, S.P., Singh, A., Desai, A.R., Dubois, S.G., Jablonski, A.D., Kingdon, C.C., et al. (2015). Remotely
  estimating photosynthetic capacity, and its response to temperature, in vegetation canopies using
  imaging spectroscopy. *Remote Sensing of Environment, 167*, 78-87
- Shenkin, A., Bentley, L.P., Oliveras, I., Salinas, N., Adu-Bredu, S., Marimon, B.H., et al. (2019). The
  Influence of Ecosystem and Phylogeny on Tropical Tree Crown Size and Shape. *BioRxiv*, 789255

- Shiklomanov, A.N., Dietze, M.C., Viskari, T., Townsend, P.A., & Serbin, S.P. (2016). Quantifying the
  influences of spectral resolution on uncertainty in leaf trait estimates through a Bayesian approach to
  RTM inversion. *Remote Sensing of Environment, 183*, 226-238
- Soudzilovskaia, N.A., Elumeeva, T.G., Onipchenko, V.G., Shidakov, I.I., Salpagarova, F.S., Khubiev, A.B., et
  al. (2013). Functional traits predict relationship between plant abundance dynamic and long-term
  climate warming. *Proceedings of the National Academy of Sciences, 110*, 18180-18184
- 894 Sullivan, M.J., Talbot, J., Lewis, S.L., Phillips, O.L., Qie, L., Begne, S.K., et al. (2017). Diversity and carbon 895 storage across the tropical forest biome. *Scientific Reports*, *7*, 39102
- Szabó, L., Burai, P., Deák, B., Dyke, G.J., & Szabó, S. (2019). Assessing the efficiency of multispectral
- satellite and airborne hyperspectral images for land cover mapping in an aquatic environment with
   emphasis on the water caltrop (Trapa natans). *International Journal of Remote Sensing*, 40, 5192-5215
- Taubert, F., Fischer, R., Groeneveld, J., Lehmann, S., Müller, M.S., Rödig, E., et al. (2018). Global patterns
  of tropical forest fragmentation. *Nature*, *554*, 519
- 901 Townsend, P.A., Foster, J.R., Chastain, R.A., & Currie, W.S. (2003). Application of imaging spectroscopy to
- 902 mapping canopy nitrogen in the forests of the central Appalachian Mountains using Hyperion and
- 903 AVIRIS. *IEEE Transactions on Geoscience and Remote Sensing*, *41*, 1347-1354
- 904 Ustin, S.L., & Gamon, J.A. (2010). Remote sensing of plant functional types. *New Phytologist, 186*, 795905 816
- Van der Plas, F., Ratcliffe, S., Ruiz-Benito, P., Scherer-Lorenzen, M., Verheyen, K., Wirth, C., et al. (2018).
  Continental mapping of forest ecosystem functions reveals a high but unrealised potential for forest
  multifunctionality. *Ecology Letters*, 21, 31-42
- Vincent, G. (2001). Leaf photosynthetic capacity and nitrogen content adjustment to canopy openness in
   tropical forest tree seedlings. *Journal of Tropical Ecology*, *17*, 495-509
- Walker, A.P., McCormack, M.L., Messier, J., Myers-Smith, I.H., & Wullschleger, S.D. (2017). Trait
  covariance: the functional warp of plant diversity? *New Phytologist, 216*, 976-980
- Walker, A.P., Beckerman, A.P., Gu, L., Kattge, J., Cernusak, L.A., Domingues, T.F., et al. (2014). The
   relationship of leaf photosynthetic traits–Vcmax and Jmax–to leaf nitrogen, leaf phosphorus, and
- 915 specific leaf area: a meta-analysis and modeling study. *Ecology and Evolution, 4*, 3218-3235
- 916 Wallis, C.I., Homeier, J., Peña, J., Brandl, R., Farwig, N., & Bendix, J. (2019). Modeling tropical montane
- forest biomass, productivity and canopy traits with multispectral remote sensing data. *Remote Sensing* of Environment, 225, 77-92
- Wright, I.J., Reich, P.B., Westoby, M., Ackerly, D.D., Baruch, Z., Bongers, F., et al. (2004). The worldwide
  leaf economics spectrum. *Nature*, *428*, 821-827
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# 922 List of Figure Captions

923 Figure 1. Diagram summarising the steps followed to assign trait values per Sentinel-2 pixel. 1) First the 924 vegetation plots are defined based on the GEM (Global Environmental Monitoring) dataset and 2) from 925 each vegetation plot the corner coordinates are extracted. 3) From each vegetation plot the XY position 926 of each stem  $\geq 10$  cm DBH is extracted and 4) the crown horizontal area is calculated based on the protocol 927 described in the methods section. 5) Then the Sentinel-2 imagery for the study area is processed to level 928 2A using the ESA SNAP toolbox and 6) the vegetation plot is overlaid in the Sentinel-2 image based on its 929 corner coordinates. In this last step (6) each pixel defines a 'subplot' which is the unit used to calculate 930 the trait community weighted mean based on the crown area of the trees that are contained by that pixel. 931 In 6) n refers to a given tree in a given pixel, trait i represents a given trait and x and y are values for that 932 trait. The image used as an example in step (1) was taken by Jesus Aguirre-Gutierrez over a vegetation 933 plot using a multispectral ALTUM camera mounted on an Inspire 1 drone."

Figure 2. Comparison of trait distributions across tropical regions. The boxplots are based on the pixellevel (10×10 m) community trait values for each trait and region (*n*=403 for Australia, 449 for Brazil-NX,
302 for Brazil-ST, 464 for Gabon, 620 for Ghana, 976 for Malaysia and 1280 for Peru). Horizontal lines in
each boxplot show the median value and vertical lines are the whiskers that extend to the largest value
or not further than 1.5 times the inter-quartile range. For some locations information for all traits was not
available. For full details in significant differences in mean trait values among locations see Table S4. Brazil
-NX: Nova Xavantina; Brazil -ST: Santarem.

Figure 3. Model predictions to the 20% test data from the general model which was fitted with 80% of
the trait data from across the tropics. Grey dots are the observed against predicted trait values of the
pixel-level (10 × 10 m) community weighted mean traits from the test dataset. The black line shows the
1:1 relationship between observed and predicted values. Model prediction accuracy is shown in the top
left. Full model results are shown in Table 4.

## 946 Figure 4. Spatial predictions of trait distributions for a selected subset of plant traits and locations.

947 The map (middle) shows the locations of vegetation plots that were used during the modelling

948 framework. The spatial predictions (top and bottom rows) were obtained using the general models

949 (Table 4) for each of the traits and locations at a 10 × 10 m pixel resolution. The approximated location

950 of each vegetation plot used is shown as a white square within each spatial prediction map (for

visualisation purposes white squares are not scaled to the plot's real size). Spatial predictions for othertraits can be found in Fig. S1-S7.

# 953 Figure 5. Models predictions to the 20% test data from the regional models fitted with 80% of the trait

data from each region across the tropics. Each colour represents an individual regional model and the
coloured symbols are the observed against predicted trait values of the pixel-level (10 × 10 m) community
weighted mean traits from the test dataset. The black line shows the 1:1 relationship between observed
and predicted values. Model prediction accuracy is shown in Table 5. Full model results are shown in Table
S5. Brazil -NX: Nova Xavantina; Brazil -ST: Santarem.

# 959 Figure 6. Group median variable importance of spectral remote sensing, environmental and soil

960 related variables for determining functional trait predictions in the global model. Variable importance

961 (Y axis) ranges from 0 (no importance) to 1 (highest importance) and represents the decrease in node

- 962 impurities from splitting on the variable, averaged over all trees and derived from the Out of Bag error,
- the resulting value has been standardised to a 0-1 scale for comparison purposes. The spectral group (S2
- 964 -Spectral) contains the select raw bands from the Sentinel-2 and the vegetation indices; Texture
- 965 parameters (S2 -Texture) contain the Correlation and Entropy metrics from the grey level co-occurrence
- 966 matrix obtained from the vegetation indices; Climate contains all climatic variables; Soil-Terrain contains
- all soil characteristics and slope. All variables are described in Table 3.

	Spacios complad		Sizo	Centroid coo	ordinates	Date of collection		
Location	for traits	Plot code	(ha)	x	Y	Vegetation census	Traits	
		AEP-02	0.5	145.586	-17.146			
Australia	81	AEP-03	0.5	145.592	-17.088	2011	June-September	
Australia		DRO-01	0.9	145.430	-16.103	2011	2015	
		ROB-06	1	145.630	-17.121			
		ANK-01	1	-2.696	5.268	2012		
Ghana	63	ANK-03	1	-2.692	5.271	2015	October-March	
Glialia	05	BOB-01	1	-1.339	6.691	2015	2015/2016gramm	
		BOB-02	1	-1.319	6.704	2015		
		LPG-01	1	11.574	-0.174	2014	February-March	
Gabon	41	LPG-02	1	11.615	-0.216	2014	2017	
		MNG-04	1	9.324	0.577	2016	2017	
		NXV-01	1	-52.352	-14.708			
		NXV-02	1	-52.351	-14.701	2015		
		VCR-02	1	-52.168	-14.832			
		NXV-10-1	0.1	-52.353	-14.713			
		NXV-10-2	0.1	-52.352	-14.713			
		NXV-10-3	0.1	-52.351	-14.713			
Brazil -NX	64	NXV-10-4	0.1	-52.349	-14.713		March-May 2014	
		NXV-10-5	0.1	-52.346	-14.713	2014		
		NXV-10-6	0.1	-52.349	-14.712	2014		
		NXV-10-7	0.1	-52.348	-14.711			
		NXV-10-8	0.1	-52.347	-14.711			
		NXV-10-9	0.1	-52.347	-14.711			
		NXV-10-10	0.1	-52.346	-14.712			
Brazil -ST	136	261-10	0.25	-55.005	-3.019	2014		

Table 1. Collection details for vegetation plots and plant functional traits. A total of 2434 individual trees were sampled	ៅ for
functional traits.	

		261-9	0.25	-55.015	-3.040		
		363-6	0.25	-54.956	-3.337		August-September
		363-3	0.25	-54.963	-3.297		2015
		363-7	0.25	-54.961	-3.321		
		ESP-01	1	-71.595	-13.176		
		PAN-02	1	-71.263	-12.650		
		SPD-01	1	-71.542	-13.047	2012	
		SPD-02	1	-71.537	-13.049	2015	
Doru	150	TRU-04	1	-71.589	-13.106		April-November
Felu	139	WAY-01	1	-71.587	-13.191		2013
		ACJ-01	1	-71.632	-13.147	2014	
		PAN-03	1	-71.274	-12.638		
		TAM-05	1	-69.271	-12.830		
		TAM-06	1	-69.296	-12.839		
		SAF-01	1	4.732	117.619		
		SAF-02	1	4.739	117.617		
		SAF-03	1	4.691	117.588		
Malavaia	202	SAF-04	1	4.765	117.700	2010	July-December
walaysia	283	DAN-04	1	4.951	117.796	2016	2015
		DAN-05	1	4.953	117.793		
		MLA-01	1	4.747	116.970		
		MLA-02	1	4.754	116.950		

Brazil -NX: Nova Xavantina; Brazil -ST: Santarem; Malaysia: Malaysian Borneo.

Table 2. Description of plant functional traits collected across the tropics and their relevance under a changing environment.

Trait	Abbreviation	Units	Description	Relevance	References*		
Leaf area	Area	cm <sup>2</sup>	One-sided area of the leaf	Higher leaf area could result in higher levels of light capture and photosynthetic activity.			
Specific leaf area	SLA	m <sup>2</sup> g <sup>-1</sup>	One-sided area of a leaf divided by dry mass	Relevant for photosynthetic capacity, light capture, water loss, net assimilation rate, leaf life span.	_		
Leaf thickness	Thickness	mm	Thickness of a fresh leaf	Trade-off between decreasing water transpiration at the expense of higher construction investment and probably lower photosynthetic efficiency in thicker leaves.	ver 5		
Leaf nitrogen content	Ν	%			(Walker et al.,		
Leaf phosphorus content	Р	%		Nutrient relevant for metabolic reactions,	2014, Wright et al., 2004, Juneau & Tarasoff, 2012, Díaz et al 2016		
Leaf carbon content	С	%	Contant nor unit dry loof mass	including light capture, related to			
Leaf calcium content	Ca	%	Content per unit dry lear mass	Restricted availability of some nutrients may			
Leaf potassium content	К	%		limit plant carbon acquisition and growth.	Hawthorne, 1995,		
Leaf magnesium content	Mg	%			Chave et al.,		
Leaf water content	LWC	%	Amount of water in the leaf relative to its dry and fresh mass	Leaf mass is a proxy of leaf biomass investment which may vary depending on	2006, Huang et al., 2019)		
Leaf dry mass	Dry mass	g	Mass of a dry leaf	environmental conditions and phenology of			
Leaf fresh mass	Fresh mass	g	Mass of a fresh leaf	species. It has been considered relevant for photosynthetic potential.			
A <sub>max</sub>	A <sub>max</sub>	µmol m <sup>-2</sup> s <sup>-1</sup>	Light-saturated maximum rates of net photosynthesis at saturated CO <sub>2</sub> (2000 ppm CO <sub>2</sub> )	Indicate the maximum CO <sub>2</sub> assimilation and			
A <sub>sat</sub>	A <sub>sat</sub>	µmol m <sup>-2</sup> s <sup>-1</sup>	Light-saturated rates of net photosynthesis at ambient CO <sub>2</sub> concentration (2000 ppm CO <sub>2</sub> )	are thus indices of leaf photosynthetic capacity			

\*References are not exhaustive.

**Table 3. Spectral remote sensing, environmental and soil related variables used during the modelling protocol.** All climatic variables but slope were calculated using a climatology of 30 years (1986-2015). All soil variables were calculated for the top 30cm soil layer. Sentinel-2 band wavelengths (nm) are given in parenthesis after the band name.

Туре	Variable	Description	References			
RS	B2 (490), B3 (560), B4 (665), B8 (842)	Sentinel-2 bands with spatial resolution of 10m				
	B5 (705), B6 (740), B7 (783), B11 (1610), B12 (2190)	Sentinel-2 bands with spatial resolution of 20m	<u>www.esa.int</u>			
	MCARI	Modified Chlorophyll Absorption in Reflectance Index	(Daughtry et al., 2000)			
	MSAVI2	Modified Soil Adjusted Vegetation Index 2	(Qi et al., 1994)			
	NDRE	Normalized Difference Red edge Index	(Barnes et al., 2000)			
	Toyturo	Entropy, calculated for vegetation indices	(Haralick et al. 1973)			
		Correlation, calculated for vegetation indices				
	MCWD	Mean annual climatic water deficit				
Climato	MATmax	Mean maximum annual temperature	(Abatzoglau at al. 2018)			
Climate	SM	Soil moisture as a water balance indicator				
	SRAD	Downward Solar Radiation				
	eCEC	Cation Exchange Capacity (mmol <sup>+</sup> /kg <sup>-1</sup> )	Plot level soil data from			
	рН	Soil pH (H2O solution)	the Global Environmental			
SOII- Torrain	Clay (%)	Amount of clay (weight %)	Monitoring (GEM)			
renam	Sand (%)	Amount of sand (weight %)	database			
	Slope	Terrain slope (30 m resolution)	(Farr et al. 2007)			

Туре	Trait	MAE	RMSE	R <sup>2</sup>
	Area (cm <sup>2</sup> )	28.32	39.854	0.43
Morphologi	Dry mass (g)	0.349	0.48	0.27
cal and	Fresh mass (g)	0.799	1.075	0.26
structural	SLA (m <sup>2</sup> g <sup>-1</sup> )	0.001	0.001	0.50
	Thickness (mm)	0.034	0.046	0.52
	LWC (%)	3.718	4.886	0.36
	C (%)	1.237	1.615	0.70
	Ca (%)	0.14	0.204	0.64
Chemistry	K (%)	0.133	0.186	0.63
	Mg (%)	0.055	0.075	0.46
	N (%)	0.23	0.3	0.59
	P (%)	0.015	0.02	0.59
Photosynth	Amax (µmol m <sup>-2</sup> s <sup>-1</sup> )	2.89	3.937	0.67
etic	Asat (µmol m <sup>-2</sup> s <sup>-1</sup> )	1.297	1.734	0.55

Table 4. Statistical results on the test data (20% of full dataset) for theglobal trait distribution models. The prediction accuracy is shown by theR<sup>2</sup> score.

MAE: Mean Absolute Error; RMSE: Root mean square error.

				, , ,											
Location	P (%)	Amax (µmol m <sup>-2</sup> s <sup>-1</sup> )	Ca (%)	N (%)	Thickness (mm)	Asat (µmol m <sup>-2</sup> s <sup>-1</sup> )	Mg (%)	C (%)	SLA (m <sup>2</sup> g <sup>-1</sup> )	LWC (%)	K (%)	Dry mass (g)	Area (cm²)	Fresh mass (g)	Region mean R <sup>2</sup>
Australia	0.21	-	0.33	0.17	0.21	0.03	0.12	0.34	0.25	-	0.06	-	0.35	-	0.21
Brazil -NX	0.68	0.42	0.49	0.52	0.66	0.52	0.46	-	0.54	0.07	0.07	0.38	0.08	0.31	0.40
Brazil -ST	0.47	-	0.15	0.30	0.42	-	0.28	0.07	0.29	0.05	0.29	0.25	0.04	0.18	0.23
Gabon	0.60	-	0.39	0.50	0.23	-	0.52	0.22	0.15	0.38	0.24	0.22	0.11	0.11	0.31
Ghana	0.47	0.49	0.53	0.52	0.22	0.36	0.15	-	0.14	-	0.23	-	0.29	-	0.34
Malaysia	0.34	0.48	0.50	0.27	0.31	0.36	0.31	0.38	0.06	0.24	0.28	0.07	0.11	0.03	0.27
Peru	0.49	0.36	0.69	0.44	0.64	0.38	0.46	0.47	0.32	0.34	0.30	0.09	0.18	0.20	0.38
Trait mean R <sup>2</sup>	0.47	0.44	0.44	0.39	0.38	0.33	0.33	0.30	0.25	0.22	0.21	0.20	0.17	0.17	

Table 5. Prediction accuracy (R<sup>2</sup>) on the testing data among regions (shaded Region mean R<sup>2</sup> column) and functional traits (shaded Trait mean R<sup>2</sup> row). Not shaded values in the table show the prediction accuracy (R<sup>2</sup>) on the test data per region and trait.

976 - :no data available; Brazil -NX: Nova Xavantina; Brazil -ST: Santarem.

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