# 1 Grassland biodiversity restoration increases resistance of carbon fluxes to

2 drought

3 Andrew J. Cole<sup>1,2</sup>, Robert I. Griffiths<sup>3</sup>, Susan E. Ward<sup>2</sup>, Jeanette Whitaker<sup>1</sup>, Nicholas J. Ostle<sup>2</sup>, Richard

4 D. Bardgett<sup>4</sup>

<sup>5</sup> <sup>1</sup> Centre for Ecology & Hydrology, Lancaster Environment Centre, Library Avenue, Bailrigg, Lancaster

6 LA1 4AP, UK.

- <sup>7</sup> <sup>2</sup> Lancaster Environment Centre, Lancaster University, Lancaster, LA1 4YQ, UK.
- 8 <sup>3</sup> Centre for Ecology & Hydrology, Maclean Building, Benson Lane, Crowmarsh Gifford, Wallingford

9 OX10 8BB, UK.

- <sup>4</sup> School of Earth and Environmental Sciences, Michael Smith Building, The University of Manchester,
- 11 Oxford Road, Manchester M13 9PT, UK.
- 12
- 13 Correspondence: Andrew J. Cole, Centre for Ecology & Hydrology, Lancaster Environment Centre,
- 14 Library Avenue, Bailrigg, Lancaster LA1 4AP, UK (andjcole@gmail.com)

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16 Running headline: Grassland restoration increases resistance to drought

17

## 18 Abstract

19 1. Evidence suggests that the restoration of plant diversity in grasslands not only brings benefits 20 for biodiversity conservation, but also the delivery of ecosystem services. While biodiversity-21 function experiments show that greater plant diversity increases resistance of plant productivity 22 to climate extremes, it is not known whether real-world management options for grassland 23 restoration likewise stabilise ecosystem responses to extreme climate events. We used a long-term (23 year) field experiment in northern England to test the hypothesis that
 management aimed at biodiversity restoration increases the resistance and recovery of
 ecosystem carbon (C) fluxes to short-term summer drought. This was tested by measuring plant,
 soil and microbial responses to a simulated drought in experimental grassland plots where
 fertiliser application and seed addition have been managed to enhance plant species diversity.

- 3. The cessation of fertiliser application brought about small increases in plant species richness.
   Additionally, cessation of fertiliser application reduced overall plant productivity and promoted
   hemi-parasitic plants at the expense of grasses and forbs.
- Resistance of CO<sub>2</sub> fluxes to drought, measured as ecosystem respiration, was greater in non fertilised plots, as lower plant biomass reduced water demand, likely aided by proportionally
   more hemi-parasitic plants further reducing plant biomass. Additionally, legumes increased
   under drought, thereby contributing to overall resistance of plant productivity.
- 36 5. Recovery of soil microbial C and nitrogen was more rapid after rewetting than soil microbial
   37 community composition, irrespective of restoration treatment, suggesting high resilience of soil
   38 microbial communities to drought.
- Synthesis and applications. This study shows that while grassland diversity restoration
  management increases the resistance of carbon fluxes to drought, it also reduces agricultural
  yields, revealing a trade-off for land managers. Furthermore legumes, promoted through longterm restoration treatments, can help to maintain plant community productivity under drought
  by increasing their biomass. As such, grassland management strategies not only have
  consequences for ecosystem processes, but also the capacity to withstand extreme weather
  events.

Keywords: biodiversity, carbon cycling, drought, fertiliser, grassland restoration, seed addition, soil
microbial community.

### 48 Introduction

49 The restoration of plant diversity in grasslands, and the management practices required to bring it about, 50 has been a major focus of research (Smith et al. 2000; Carter & Blair 2012). In addition to increasing 51 plant diversity, grassland restoration can bring benefits for ecosystem services such as increasing soil 52 carbon (C) accumulation (De Deyn et al. 2011), plant productivity (Bullock et al. 2001) and nutrient 53 retention (Maron & Jefferies 2001). While biodiversity-function experiments show that greater plant 54 diversity can increase the resistance of plant productivity to climate extremes (Isbell et al. 2015), it is 55 not known whether real-world management options for the restoration of grassland plant diversity 56 likewise stabilise ecosystem responses to extreme climate events. One such extreme climate event is 57 drought, which is predicted to become more frequent and intense under climate change (IPCC 2013), 58 with potential to disrupt C and nitrogen (N) cycling in grasslands (Harper et al. 2005).

59 In the face of environmental perturbations such as drought, management is required to maintain the stability of ecosystem functions and services (Oliver et al. 2015; Donohue et al. 2016). Although 60 61 experimental studies show that greater species richness can increase the resistance of plant productivity to drought (Isbell et al. 2015), it is not known whether real-world grassland biodiversity restoration has 62 63 a similar effect, and whether impacts of drought on plant productivity are less in restored than 64 unrestored, species-poor grassland (Carter & Blair 2012). However, plant species introduced through 65 restoration are likely to have differing physiological adaptations to survive drought than those already 66 present on unrestored grassland (Hoekstra et al. 2014). For example, as drought can limit N availability 67 to grassland plants, in part through reduced N mobility in soil, the presence of legume species (with N-68 fixing rhizobial associations) may increase community resistant to drought (Hofer et al. 2016). 69 Although research has revealed restoration treatments which successfully increase plant diversity, it 70 remains to be seen whether there are also benefits of successful biodiversity restoration treatments in 71 terms of buffering grassland responses to drought.

72 Although drought can have direct effects on soil microbial communities, studies show that the response of plants to drought can also bring about indirect effects belowground (Bloor & Bardgett 2012). Specific 73 74 groups of microbes (e.g. fungi, bacteria and archaea) respond differently to drought (Schimel, Balser & 75 Wallenstein 2007), influencing the flow of C from plant roots into the soil (Fuchslueger et al. 2014). 76 Further, recent studies show that grassland management can alter the resistance of resilience of C 77 cycling processes to drought by changing patterns of plant C allocation and C transfer to the microbial 78 community (Karlowsky et al. 2018). This suggests grassland biodiversity restoration could likewise 79 alter the resistance and resilience of soil microbial processes to drought, for example through the 80 promotion of drought-tolerant groups such as fungi or gram-positive bacteria (Smith et al. 2008), which 81 may in turn promote soil C and N retention (De Vries et al. 2012b). However, it remains unclear whether 82 changes in the soil microbial community associated with biodiversity restoration, will in turn affect how 83 grassland C and N cycles respond to drought.

84 The goal of this study was to test whether long-term management treatments used to restore grassland plant diversity confer greater resistance and recovery of C and N cycling processes and microbial 85 86 communities to drought. This was achieved using a 23-year grassland biodiversity restoration 87 experiment at Colt Park Meadows, northern England, which has successfully brought about increased 88 plant diversity through a combination of mixed species plant seed addition and cessation of annual 89 inorganic fertiliser application, alongside shifts in the abundance of plant functional groups (Smith et 90 al. 2000, 2008; De Deyn et al. 2011). Additionally, the combined treatments of cessation of fertiliser 91 application and seed addition, has been associated with concurrent changes in the soil microbial 92 community, in particular an increase in the abundance of fungi relative to bacteria (Smith et al. 2008). 93 Such changes in microbial community composition have been linked to greater nutrient retention 94 (Bardgett & McAlister 1999; De Vries et al. 2012a) and an increase in soil C accumulation when 95 combined with legume addition (De Deyn et al. 2011).

We used rain-out shelters on selected treatments to test the hypothesis that the resistance and recoveryof C and N cycling to summer drought is enhanced by restored plant diversity. Specifically, we

98 hypothesised that: (H1) drought would reduce plant productivity, ecosystem respiration, and microbial 99 biomass and have a greater negative effect on soil bacteria than fungi; and (H2) the resistance and 100 recovery of C and N cycling processes to drought would be greater in treatments subject to long-term 101 fertiliser cessation and seed addition. This treatment combination has been associated with lower plant 102 productivity, which reduces water demand, and a greater abundance of fungi relative to bacteria in soil, 103 which has been shown to buffer effects of drought events on soil processes (De Vries *et al.* 2012b).

### 104 Materials and methods

#### 105 Experimental system

106 The study was conducted on selected plots of a long-term (23-year) grassland diversity restoration 107 experiment at Colt Park meadows, Ingleborough National Nature Reserve, northern England (latitude 108 54°12'N, longitude 2°21'W; Bardgett & McAlister 1999; Smith et al. 2000; De Deyn et al. 2011). The 109 experiment was set up in 1990 on agriculturally improved Lolium perenne-Cynosorus cristatus 110 grassland on brown earth soils over limestone bedrock, in order to identify optimal management 111 strategies for the restoration of botanical diversity (Smith et al. 2000). Onto this long term restoration 112 experiment, we superimposed short-term drought treatments on a subset of 12 plots (3m x 3m) in 3 replicate blocks from factorial treatment combinations, namely cessation or continued application of 113 114 fertiliser and with or without seed addition. These treatments were selected because the combination of 115 seed addition and cessation of inorganic fertiliser addition has resulted in the greatest increase in plant 116 diversity (Smith et al. 2008; De Deyn et al. 2011).

Before the long-term grassland restoration was initiated the meadow received regular additions of inorganic fertiliser (Smith *et al.* 2000). The fertiliser cessation treatment started in 1990 with the alternate treatment being continued fertiliser application (NPK 20:10:10; 25kg N ha<sup>-1</sup>) which has since been applied to plots by hand annually in spring (21 May in 2013), except in 2009 and 2010. Seed addition treatment also started in 1990 with seed of 19 species coming from locally collected and commercially bought seed. Since 1999 all plots have been cut for hay between mid-July and August 123 (16 July in 2013), and then grazed by sheep and cattle until hay production in May (Smith *et al.* 2000, 124 2003, 2008; De Deyn et al. 2011). The year prior to establishing the drought experiment, species 125 richness across  $4m^2$  was greatest in the fertiliser cessation treatment (26.5±1.4) and lowest with 126 continued fertiliser application (22.7±1.6; LRT=4.70, d.f.=1,5, P=0.030).

#### 127 Drought treatments

To investigate the effect of drought on C and N cycling in grasslands, we set up 3 levels of the drought 128 129 treatment in each of the 12 long-term experimental plots in June 2013 (Fig 1). The three treatments 130 were: ambient (no rain-out shelter), drought (rain-out shelter) and roofed control (rain-out shelter with 131 holes). Rain-out shelters were open sided, constructed of transparent corrugated PVC, 0.8mm thick 132 (Corolux, UK). Shelters were 90cm x 105cm with a height of 38cm-63cm, giving a sloped roof of 16 133 degrees. Roofed control shelters were identical to those used in the drought treatment, except they 134 contained holes to allow rainfall to reach the plot and were used to assess any artefacts of having rain-135 out shelters in place. The rain-out shelters were in place from 5 June-10 July 2013 (35 days) to match the length of 100-year drought events. For this, long-term precipitation data from Malham tarn was 136 137 fitted with Gumbel I distribution for consecutive days with <1mm precipitation during the primary 138 growth period (April-September), following Bloor & Bardgett (2012). The effect of rain-out shelters on 139 soil moisture was assessed using a ThetaProbe soil moisture meter (Delta-T, UK), and temperature was 140 measured using Hobo Pendant temperature loggers (Onset, USA) at 5cm depth for soil and 5cm above 141 soil surface for air temperature in ambient, control shelter and drought plots. Measurements of soil DOC 142 and DON, soil microbial community and vegetation C and N content were only made on the ambient 143 and drought treatment and not the roofed control treatment.

#### 144 *Plant community*

Aboveground plant biomass was harvested from all plots on 10 July, after the end of the drought treatment. The plant biomass was dried at 60°C for 48 hours and split by hand into plant functional types (PFT): grasses, forbs, legumes and hemi-parasitic plants. After ball milling, C and N content for

- 148 each PFT was measured on a Tru-spec CN analyser (Leco, St. Joesph, MI, USA). Vascular plant species
- surveys were carried out between 29 June and 4 July 2013 on the central 706cm<sup>2</sup> of each plot.

# 150 *CO*<sub>2</sub> *flux measurements*

Net ecosystem exchange (NEE) and ecosystem respiration were measured using static chambers following Ward *et al.* (2013), linked to an infra-red gas analyser (EGM4, PP Systems, UK), as used by De Deyn *et al.* (2011). Two minute headspace closures were used for NEE (transparent chambers) and ecosystem respiration (opaque chambers) between 10:15 and 16:30 hrs alongside measurements of photosynthetically active radiation (PAR), soil and air temperature. Two measurements were made before the drought (17, 24 May), six during the experimental drought (13, 20, 24, 27 June; 5, 8 July 2013) and five after shelter removal (19, 26, 30 July; 7, 16 August 2013).

#### 158 Soil microbial community

Soil was sampled at four time-points with three cores (2.4cm diameter, 10cm depth) taken from each subplot, bulked together and sieved (2mm). Sampling dates were 5 June (before drought), 10 July (during drought), 16 August (3 weeks after rewetting) and 4 November (3 months after rewetting). Before the drought treatment was imposed, samples were only taken from the 12 main plots to determine treatment effects of seed addition and fertiliser cessation. To quantify belowground biomass, roots were removed from soil cores sampled during the drought with roots sieved, washed and dried at 60°C for 48 hours before weighing.

The effect of grassland restoration and drought on bacterial and fungal community composition was assessed using the terminal restriction fragment length polymorphism (T-RFLP) method as detailed by Plassart *et al.* (2012). Genomic DNA was extracted from soil samples using the PowerSoil kit (MoBio, Carlsbad, US) and amplified using primers for bacterial 16SrRNA and fungal ITS genetic markers. In addition, broad-scale changes in soil microbial community composition were assessed by phospholipid fatty acid analysis (PLFA). Briefly, PLFA's were extracted from freeze-dried soil using a modified 172 Bligh-Dyer extraction and separated from other lipids using aminopropyl solid phase extraction 173 cartridge (Phenonenex, US; White *et al.* 1979). Gas chromatography was carried out on an Agilent 174 6890GC with fused silica capillary column (Agilent, US). Biomarkers were used for bacteria (i15:0, 175 a15:0, 15:0, i16:0, 17:0, i17:0, cy-17:0,  $18:1\omega7$  and cy-19:0) and saprotrophic fungi ( $18:2\omega6,9$ ) 176 (Bardgett, Hobbs & Frostegard 1996; Smith *et al.* 2008).

177 Microbial biomass C and N were measured on 5g fresh soil subsamples using the chloroform 178 fumigation-incubation method (Brookes et al. 1985) in soil cores taken before, during and after 179 (August) the drought. One subsample was fumigated with chloroform for 16 hours before extraction 180 with 25mL 0.5M K<sub>2</sub>SO<sub>4</sub>. The resulting filtrate was analysed for microbial C using a TOC analyser 181 (5000A, Shimadzu, Milton Keynes, UK). For microbial N, filtrate was oxidised with  $K_2S_2O_8$  before 182 colorimetric analysis on an autoanalyser (Bran and Luebbe, Northampton, UK). Adjustment factors 183 were applied, using K<sub>c</sub>=0.35 for microbial C and k<sub>n</sub>=0.54 for microbial N (Bloor & Bardgett 2012). For 184 soil samples during and after (August) the drought, DOC and DON were extracted from 5g subsamples using 35mL of water with DOC analysed on a TOC analyser (5000A, Shimadzu, Milton Keynes, UK) 185 186 and DON extract oxidised with K<sub>2</sub>S<sub>2</sub>O<sub>8</sub> before colorimetric analysis (Bran and Luebbe, Northampton, UK). 187

# 188 Statistical analysis

To investigate responses of plant and soil measurements to drought, we used percentage change caused by drought and rewetting as indices of resistance and recovery, as widely used in previous studies (reviewed in Orwin & Wardle 2004). Specifically resistance and recovery were calculated as  $(P_0-C_0)/$  $C_0.100$ , where  $P_0$  is the drought treatment response and  $C_0$  is the control response. Where data were available, resistance and recovery were calculated using both the ambient and control shelter treatments as  $C_0$  to allow the effect of the rain-shelter to be investigated.

Linear mixed effects models (LME) were used to investigate plant and soil microbial responses to seed
addition, fertiliser and drought treatments. For each LME model the fixed effects were seed, fertiliser,

197 drought and all interactions. The random effect was split-plot nested within block to take account of the 198 experimental split-plot design. Where data included multiple measurements from a single plot, plot ID 199 was added as an additional nested random effect. Assumptions of normality and equal variances were 200 checked graphically and where necessary response variables were either logged or used varIdent weight 201 functions to improve model fit [e.g. varIdent(form=~1|fertiliser)], following Zuur et al. (2009). We 202 determined the significance of fixed effects by comparing models with and without the factor of interest 203 using likelihood ratio tests (LRT). To investigate changes in soil microbial community structure, the 204 relative abundance of T-RF peaks was assessed using between-sample Bray-Curtis dissimilarities and 205 non-metric multidimensional scaling with permutational multivariate analysis of variance. All statistical analysis was carried out in the R programming language 3.3.1 (R Core Development Team, 2016) using 206 207 the nlme package for mixed effect models (Pinheiro et al. 2013).

#### 208 **Results**

#### 209 Rain shelter effects on soil moisture and temperature

210 The drought treatment excluded 180.8mm of rainfall over 35 days with significant rewetting occurring 211 in late July, more than a week after rain-out shelters were removed (Fig 1b). The treatment intercepted 212 rainfall equivalent to 7.5% of average annual precipitation and reduced soil moisture from 58.8% to 213 33.3%, while the roofed control shelter also intercepted some rainfall and reduced soil moisture from 214 58.8% to 49.8% (Fig 1a, LRT=113.55, d.f.=2,10, P<0.0001). Additionally, fertiliser application reduced 215 soil moisture by 3% relative to cessation of fertiliser (LRT=5.68, d.f.=1,11, P=0.017). Two nearby sites 216 with long-term precipitation datasets suggest that 100-year extreme drought events in the primary 217 growing season equate to 27 and 34 days with <1mm rainfall (Bloor & Bardgett 2012), similar to the 218 35 days rain-out shelters were in place in this study. No effects of rain-out shelters were detected on 219 mean air temperature (LRT=2.45, d.f.=2,6, P=0.293) or soil temperature (LRT=1.72, d.f.=2,7, 220 *P*=0.424).

222 Cessation of fertiliser use brought about an average 1.2 species increase in plant species richness 223 (LRT=6.46, d.f.=1,10, P=0.011), while seed addition increased species richness, but only in the ambient 224 treatment which had no rain shelter (Fig 2a, drought x seed: LRT=8.59, d.f.=2,13, P=0.014). Total 225 aboveground biomass, harvested in July, was 49.4% lower in non-fertilised compared to fertilised plots 226 (LRT=18.45, d.f.=1,10, P<0.0001), while seed addition had no effect (Fig 2b, LRT=1.36, d.f.=1,10, 227 P=0.244). For specific plant functional types, grass and forb biomass was respectively 57.2% and 35.2% 228 lower in non-fertilised relative to fertilised plots (Fig 3, Grass: LRT=19.02, d.f.=1,10, P<0.0001; Forb: 229 LRT=9.43, d.f.=1,11, P=0.002), while hemi-parasitic plant biomass was nearly double (Fig 3d, 230 LRT=3.94, d.f.=1,10, P=0.047). As a consequence, the proportion of plant functional groups shifted in 231 non-fertilised plots, with proportionally less grass biomass and more legume and hemi-parasitic plant 232 biomass (Fig S1).

233 Drought, superimposed across the long-term grassland restoration treatments, did not reduce total 234 aboveground (Fig 2a, LRT=2.50, d.f.=2,9, P=0.286) or root biomass (Fig 3a, LRT=2.93, d.f.=2,9, 235 P=0.231). However, legume biomass increased under drought (Fig 2c, LRT=8.00, d.f.=2,11, P=0.018), 236 whereas for grasses drought had no impact, although forb biomass was marginally increased by drought 237 but only in plots with seed addition (Fig 3). Additionally, the hemi-parasitic plant species R. minor was more abundant in plots without than with seed addition, but primarily so under drought conditions (Fig 238 239 3e, drought x seed: LRT=6.43, d.f.=2,13, P=0.040). Furthermore, in grasslands with seed addition the 240 resistance of N content in shoot biomass was increased across all forbs, grasses and legumes (Fig 4a, 241 LRT=5.52, d.f.=1,12, P=0.019).

242  $CO_2$  fluxes

Ecosystem respiration increased with soil temperature (LRT=12.57, d.f.=1,77, P=0.0004), while continued fertiliser application also increased ecosystem respiration, with generally larger increases later in the growing season (Fig S2, date x fertiliser: LRT=11.56, d.f.=5,52, P=0.041). Furthermore, 246 grasslands with continued fertiliser application also had the greatest reduction in ecosystem respiration under drought compared with ambient and control shelter treatments (Fertiliser x drought: LRT=6.83, 247 d.f.=2,55, P=0.033), although the impact of drought was greatest in mid-June (Fig S2, date x drought: 248 LRT=20.52, d.f.=10,47, P=0.025). As such, the reduction in ecosystem respiration correlated negatively 249 250 with aboveground plant biomass compared to both ambient and control shelter treatments (Fig 6). 251 Consequently, resistance of ecosystem respiration to drought, expressed as percentage change, was 252 reduced by continued fertiliser application relative to ambient plots with no rain-shelter (LRT=3.86, 253 d.f.=1,16, P=0.049) and control shelters plots (Fig 5, LRT=5.41, d.f.=1,16, P=0.020).

254 After the hay cut, ecosystem respiration increased over time, particularly in fertilised plots (Date x 255 fertiliser: LRT=18.22, d.f.=4,45, P=0.001) and in those previously exposed to drought, although the increase occurred more quickly with seed addition (Fig S2, date x seed x drought: LRT=16.48, 256 257 d.f.=8,57, P=0.036). Recovery of ecosystem respiration, expressed as percentage increase, also suggested recovery was initially lower in grasslands with seed addition, but increased two to three 258 259 weeks after rewetting in early August, with a similar pattern when using either ambient treatment (Seed 260 x date: LRT=12.99, d.f.=4,19, P=0.011) or control shelter as reference treatment (Fig 5, seed x date: LRT=11.03, d.f.=4,19, P=0.026). 261

262 NEE increased with PAR (LRT=85.09, d.f.=1,84, P<0.0001), soil temperature (LRT=4.88, d.f.=1,84, 263 P=0.027) and fertiliser application, although this differed through the growing season (Fig S3, fertiliser 264 x date: LRT=21.06, d.f.=5,59, P=0.0008). Additionally, NEE varied slightly across drought treatments 265 depending on sampling date (Fig S3, date x drought: LRT=31.52, d.f.=10,54, P=0.0005). Resistance of 266 NEE to drought increased with fertiliser application on particular sampling dates, calculated as 267 percentage change relative to ambient plots with no rain-shelter (Fig S4a, fertiliser x date: LRT=13.74, 268 d.f.=5,22, P=0.017) and control shelters plots (Fig S4c, fertiliser x date: LRT=13.65, d.f.=5,22, 269 *P*=0.018).

After natural rewetting, NEE increased with PAR (LRT=30.50, d.f.=1,66, P<0.0001) and increased over time, but varied with fertiliser cessation and seed addition (Fig S3, date x fertiliser x seed: LRT=11.04, d.f.=4,58, P=0.026). Recovery of NEE, expressed as percentage change after rewetting, varied across sampling dates (Fig S4b, LRT=12.21, d.f.=4,11, P=0.016), while when using the control shelter as the reference treatment, recovery was marginally slower in communities with seed addition, but only when combined with fertiliser application (Fig S4d, fertiliser x seed: LRT=4.12, d.f.=1,23, P=0.042).

### 277 Soil properties and microbial community

Microbial biomass C and N were partially reduced by drought (Fig S5), with reduced resistance in grasslands with seed addition (Fig 4b, microbial biomass C: LRT=12.47, d.f.=1,5, *P*=0.0004; microbial biomass N: LRT=4.79, d.f.=1,6, *P*=0.029). Three weeks after the drought, both microbial biomass C and N showed full recovery to levels in non-droughted plots (Fig S5, 4c). In contrast to soil DOC, soil DON was partially reduced by drought (Fig S6), with reduced resistance in grasslands without seed addition (Fig 4b, LRT=4.71, d.f.=1,5, *P*=0.030). However after rewetting, recovery of DON was almost 4 fold greater with continued fertiliser application (Fig 4c, LRT=6.07, d.f.=1,7, *P*=0.014).

Bacterial PLFAs did not change in response to drought or rewetting (Fig 7a, S5), while bacterial community composition, assessed by T-RFLP, was altered by drought (Fig S3, F=3.28, d.f.=1,12, P=0.010), although after rewetting in August the community differed with seed addition (Fig S3, F=3.04, d.f.=1,14, P=0.020). In contrast, fungal PLFA generally increased under drought (Fig S7f, fertiliser x seed x drought: LRT=4.26, d.f.=1,11, P=0.039), although after rewetting recovery was slower in grasslands with seed addition and fertiliser application (Fig 7b) but with no difference win recovery in November (Fig 7b).

#### 292 **Discussion**

293 The aim of this study was to investigate whether real-world management options for the restoration of 294 grassland plant diversity brought about changes in the capacity for plant and soil microbial communities 295 to resist and recover from drought. Differences in plant species richness across treatments were small, 296 but this study suggests grasslands most resistant to drought are likely to be those with greater 297 proportions of legumes and hemi-parasitic plants, but lower plant biomass. This indicates a trade-off 298 for management between resistance to drought and agricultural yield. Overall, this study suggests that 299 biodiversity restoration management through long-term fertiliser cessation and addition of key plant 300 species can have positive effects for resistance to summer drought events.

In this study, drought was superimposed on a 23-year grassland restoration experiment in which seed addition and fertiliser cessation have brought about the greatest increase in plant diversity (Smith *et al.* 2003, 2008). Following previous studies, fertiliser cessation and seed addition have continued to bring about generally small increases in species richness, alongside shifts in plant functional groups from grasses to legumes and hemi-parasitic plants and an increase in soil fungi relative to bacteria (Smith *et al.* 2000, 2003, 2008).

307 The impact of drought on grasslands subject to biodiversity restoration management was tested using 308 both a complete rain shelter and a control shelter with holes to test for any unanticipated effects of the 309 shelter (Vogel et al. 2013). The control shelter reduced soil moisture by 9% making it difficult to 310 separate unanticipated side-effects of the shelter from those caused by the reduced soil moisture. Even 311 so, any unanticipated artefacts of the shelter were most likely to be seen in CO<sub>2</sub> fluxes, which were 312 sensitive to PAR and soil temperature, and which can be impacted by shelters (Vogel et al. 2013). In 313 contrast, the reduction in ecosystem respiration under drought was similar irrespective of whether it 314 was relative to the ambient or control shelter treatment (Fig 6). This suggests that results are unlikely 315 to be impacted by unanticipated artefacts of the rain shelter, with overall conclusions robust to changing 316 how drought data are included in statistical models (Table S2, S3). Therefore the subset of 317 measurements that were not able to include the extra level of control are still likely to represent the 318 effect of drought rather than the shelter.

# 319 Drought and plant community

320 Contrary to the first hypothesis (H1), plant productivity was resistant to drought, however, legume 321 biomass increased under drought irrespective of grassland restoration treatment. The increase in 322 legumes under drought was only significant when compared to ambient (no shelter), and not the control 323 shelter. This suggests the reduction in soil moisture under the control shelter may have led to 324 intermediate legume biomass, although an unanticipated effect of the rain-shelter cannot be ruled out. 325 Although communities containing legumes can have greater reductions in productivity under drought 326 (Pfisterer & Schmid 2002), legumes may be more resistant to drought, as by accessing atmospheric N 327 they can avoid the N limitation imposed on plants by drought due to reduced mobility of nutrients 328 (Hofer et al. 2016). Additionally, both the two most common legume species in this study, T. repens 329 and T. pratense, can access water from lower soil depths during water scarcity (Grieu et al. 2001; 330 Hoekstra et al. 2014). Under severe drought, an increase in legumes may offset reductions in 331 productivity of other plant functional types under drought, thereby helping to maintain overall plant 332 productivity. In this long-term study T. pratense was initially introduced through seed addition (Smith et al. 2000) and promoted by cessation of fertiliser applications (Smith et al. 2003), which suggests that 333 through introducing and promoting legumes, grassland restoration may increase resistance of plant 334 335 productivity to drought.

### 336 Drought and $CO_2$ fluxes

As hypothesised (H1), drought reduced ecosystem respiration by 12.3% in grasslands with fertiliser addition but with minimal impact in grasslands with cessation of fertiliser. Additionally, this result was robust to the use of either ambient or control shelter treatments to calculate resistance indices. The greater resistance of ecosystem respiration to drought was likely due to low productivity grasslands being more resistant to drought, due to reduced water demand (Wang, Yu & Wang 2007). This suggests 342 a potential trade-off between agricultural yields and resistance to drought. In this long-term experiment, hay yields reduced as R. minor became abundant 6 years after initial seed addition (Smith et al. 2000), 343 344 suggesting hemi-parasitic plants may increase resistance of C cycling to drought through reducing plant productivity, in addition to enabling slow-growing species to establish (Joshi et al. 2000, Bardgett et al. 345 346 2006). These conclusions, however, will differ where grassland biodiversity restoration has been found to increase agricultural yields, for example in seeding experiments on ex-arable land (Bullock et al. 347 348 2001). This suggests there may be variability in the impact of grassland biodiversity restoration on 349 community resistance to drought, primarily due to the impact of restoration on agricultural yield.

### 350 Drought and soil microbial community

351 Drought reduced microbial biomass C and N in plots with seed addition, suggesting the drought 352 response was modulated by the plant community composition (Bloor & Bardgett 2012; De Vries et al. 353 2018). Additionally, the increase in fungal PLFAs under drought may point to a strong link between 354 plants and fungi under drought (Fuchslueger et al. 2014), potentially due to plants accessing water through the arbuscular mycorrhiza hyphal networks (Karlowsky et al. 2017). Unanticipated artefacts of 355 356 the rain shelter, however, cannot be ruled out, due to not having data on soil microbial communities 357 from the control shelter treatment. Nevertheless, our study suggests that the increase in the abundance 358 of soil fungi over two decades of biodiversity restoration has led to a soil microbial community that is 359 resistant to drought.

360 The soil microbial community showed high resilience after rewetting, irrespective of grassland 361 biodiversity restoration treatment. Three weeks after rewetting, microbial biomass C and N had fully 362 recovered, while the effect of drought on microbial community structure persisted. Additionally a large post-rewetting increase in soil DON followed similar increases in soil N availability post drought (Roy 363 364 et al. 2016). Recovery of the soil microbial community after drought can be variable, with different microbial groups responding differently (De Vries et al. 2012b; Fuchslueger et al. 2014; De Vries et al. 365 2018); however our data show that microbial biomass C and N can recover more quickly than microbial 366 367 community structure and this high resilience is irrespective of grassland restoration treatment.

369 This study suggests that the changes in plant and soil microbial communities brought about by drought 370 match those that aid restoration from species poor to species rich grassland. On this long-term 371 experiment, restoration was benefitted through introducing and promoting the hemi-parasitic species R. 372 *minor* and legumes, which were associated with higher abundance of fungi relative to bacteria in soil 373 (Smith et al. 2003; 2008). Similarly, in this study, grasslands exposed to drought had increased legume 374 biomass, the fungal-to-bacterial ratio and, in a subset of treatments, R. minor cover. Further work is 375 needed to assess if these shifts in one growing season have longer-term consequences for development of species rich communities. In contrast, on ex-arable grasslands, drought may impair restoration by 376 377 promoting ruderal species at the expense of species typical of old grasslands (Morecroft et al. 2004). 378 This study suggests, when restoring agriculturally improved species-poor grasslands the plants 379 promoted under drought can match those targeted by restoration treatments.

380 Research shows that grassland restoration can increase soil C accumulation (De Deyn et al. 2011), 381 nutrient retention (Maron & Jefferies 2001), and resistance of plant productivity and C fluxes to drought 382 (Wang et al. 2007; this study). Yet these benefits, and those found here, were all found to be associated 383 with lower productivity grasslands or cessation of fertiliser application. This suggests multiple trade-384 offs between broader multifunctionality and agricultural yield when restoring grassland botanical 385 diversity. Grassland restoration is commonly supported by agri-environment schemes, however 386 payments are often linked to set estimates of income forgone rather than for the public goods they 387 deliver (Fraser 2009). In the context of grassland restoration, the actual income forgone will be greater 388 where restoration leads to low yields which may make payments less attractive to the land owner, but 389 confer additional benefits (e.g. Wang et al. 2007). In contrast, grassland restoration which increases 390 agricultural yields will have minimal actual income forgone (Bullock et al. 2007), and may appear more 391 attractive. This suggests a need to disentangle the trade-offs between the benefits of grassland 392 restoration to the public (e.g. C sequestration) and land manager (e.g. productivity resistant to drought), 393 and to what extent this may change under a future climate.

#### 394 Conclusions

395 This study set out to investigate if real-world grassland biodiversity restoration treatments influence 396 plant, soil and microbial responses to extreme climate events. Although increases in plant species 397 richness were small, this study suggests that grassland biodiversity restoration increases resistance of C 398 fluxes to drought, but that this is associated with lower plant biomass. This presents a trade-off for 399 management between resistance to drought and higher agricultural yields, and suggests Agri-400 environment schemes may need to account for such trade-offs when seeking to deliver public goods. 401 Additionally, grassland biodiversity restoration aiming to increase resistance to drought should increase 402 legumes and hemi-parasitic plants, both of which were also beneficial for restoration from species poor 403 to species rich grassland over the previous two decades. Overall, our results show that grassland 404 management strategies aimed at the restoration of biodiversity, not only have consequences for 405 ecosystem processes, but also their capacity to withstand extreme weather events. This has implications 406 for future grassland management given that extreme weather events are expected to increase in intensity 407 and frequency with climate change.

### 408 Authors' contributions

409 The drought experiment was initiated and designed by AJC, SEW, NO and RDB. Data collection and 410 analysis was carried out by AJC. All authors contributed to the writing and reviewing of the paper and 411 approve publication.

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### 419 Data accessibility

- 420 Data available via the Environmental Information Data Centre https://doi.org/10.5285/8a41b2a2-01d7-
- 421 409e-adf5-fba3f3770f29 (Cole *et al.* 2019).

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534 Figure 1. Soil moisture (a) and daily precipitation (b) through the experiment. -F represents fertiliser

cessation, +F represents continued fertiliser application. Central x-axis indicates soil sampling (circles) 535

and hay cut (triangle). Vertical dashed lines show installation and removal of rain-out shelters. 536



Figure 2. Plant community responses to biodiversity restoration treatments and drought; (a) species
richness; (b) shoot biomass; (c) legume biomass, for ambient (A), control shelter (cS) and drought (D)
treatments. The significance of main effects and interactions indicated by \*\*\*=P<0.001, \*\*=P<0.01,</li>

541 \*=P<0.05 and for panel (c) different letters represent significant differences between treatments.

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Figure 3. Plant community responses to restoration treatments and drought; (a) root biomass; (b) grass biomass; (c) forb biomass; (d) hemi-parasitic plant biomass; and (e) Rhinanthus minor % cover, for ambient (A), control shelter (cS) and drought (D) treatments. The significance of main effects and interactions indicated by \*\*\*=P<0.0001, \*\*\*=P<0.001, \*\*=P<0.01, \*=P<0.05. 



**Figure 4.** Resistance (a) vegetation N content to drought and resistance (b) and recovery (c) of soil microbial and organic C and N pools to drought. Assessed as percentage change where positive values show increases under drought or following re-wetting. –S represents no seed addition, +S represents seed addition, –F represents fertiliser cessation, +F represents continued fertiliser application. The significance of main effects and interactions indicated by \*\*\*=P<0.001, \*\*=P<0.01, \*=P<0.05



Figure 5. The resistance and recovery of ecosystem respiration to drought using the ambient treatment as reference treatment (a-b) and the control shelter treatment as reference treatment (c-d). The significance of main effects and interactions indicated by \*=P<0.05. 



**Figure 6.** Reduction in ecosystem respiration with drought averaged across sampling from 20 June to

568 5 July. Dashed horizontal line indicates the same ecosystem respiration in ambient and drought.



571 **Figure 7.** The resistance and recovery to drought of: (a) bacterial; and (b) fungal PLFAs, assessed as 572 percentage change where positive values show increases under drought or following re-wetting. -S573 represents no seed addition, +S represents seed addition, -F represents fertiliser cessation, +F represents 574 continued fertiliser application. The significance of main effects and interactions indicated by 575 \*=P<0.05.