1 Leaf nutrients not specific leaf area are consistent indicators of elevated nutrient

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Abstract

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Leaf traits are frequently measured in ecology to provide a 'common currency' for predicting how anthropogenic pressures impact ecosystem function. Here, we test whether leaf traits consistently respond to experimental treatments across 27 globally distributed grassland sites across four continents. We find specific leaf area (SLA; leaf area per unit mass), a commonly measured morphological trait to infer shifts between plant growth strategies, did not respond to up to four years of soil nutrient additions. Leaf nitrogen, phosphorus and potassium concentrations did increase in response to the addition of each respective soil nutrient. We found few significant changes in leaf traits when vertebrate herbivores were excluded in the short-term. Leaf nitrogen and potassium concentrations were positively correlated with species turnover, suggesting interspecific trait variation was a significant predictor of leaf nitrogen and potassium, but not of leaf phosphorus concentration. Climatic conditions and pre-treatment soil nutrient levels also accounted for significant amounts of variation in the leaf traits measured. Overall, we find that leaf morphological traits such as SLA are not appropriate indicators of plant response to anthropogenic perturbations in grasslands.

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Text: Biodiversity loss is accelerating at an alarming rate, particularly in grasslands due to eutrophication linked to agricultural intensification and industrial pollution¹, and altered trophic level interactions such as reduced consumption by native hervivores^{2,3}. These anthropogenic pressures also impact species composition, potentially selecting for species with particular traits, and thereby affecting ecosystem function^{4,5}. Functionally relevant traits, rather than species richness, have been increasingly used as a "common currency" to assess the consequences of biodiversity loss^{6,7} on ecosystem functioning ^{8,9}. Leaf traits are commonly used, and considered as

part of the 'Holy Grail'^{6,10} set of traits, to predict plant-animal interactions¹¹, community composition and ecosystem function in response to perturbations¹².

Ecology's focus on leaf traits is based on strong eco-physiological evidence that leaves represent important investment strategies for plant growth and survival. Plants invest photosynthate and mineral nutrients in the construction of leaves, which capture light to produce more photosynthate 13,14. Leaf traits such as specific leaf area (SLA) and leaf nutrient concentrations are typically used as comparative measures of how plants capitalize on these investments. SLA, measured as leaf area per unit mass, represents a trade-off between surface area for capturing photons and thickness related to structural adaptations for water conservation and herbivore defence.

Indeed, leaf traits correlate across a continuum of fast to slow returns-on-investment, known as the leaf economic spectrum (LES) 14-16.

Fast-growing species, which are adept at resource acquisition and tend to dominate in regions with high rainfall levels and soils where resource availability is not limiting, are hypothesized to have higher SLAs and leaf nutrient concentrations^{10,17}. High SLA is associated with lower costs of leaf construction, and higher rates of herbivory as tissue becomes more palatable⁶. Additionally, higher species turnover and palatability are also positively correlated with leaf nitrogen (N), phosphorus (P), and potassium (K) concentrations¹⁴⁻¹⁶. By contrast, slower-growing species, which exhibit resource conservation, are hypothesized to have lower SLAs and leaf nutrient concentrations¹⁴⁻¹⁷. As a result, slow-growing species are less palatable to herbivores, while having a longer leaf life span.

Trade-offs between leaf traits discovered in the LES were shaped over evolutionary timeframes as successful trait combinations are selected for and unfavourable combinations are selected against. LES relationships were built from

comparative relationships among leaves collected across biomes ranging from tundra to tropical forests¹⁴. However, the extent to which rapid changes in structuring forces such as soil nutrient availability and reduced herbivory result in predictable shifts in trait values within a biome, like grasslands, remains equivocal⁶. Indeed, in agriculture the growth-dilution effect postulates that leaf nutrient concentrations may not increase in response to fertiliser because increased plant growth outpaces nutrient accumulation in tissue¹⁸

SLA and leaf nutrient concentrations are commonly used as surrogate measures of broad-scale biogeographical differences¹². However, leaf trait responses of individual species are also influenced by short-term local-scale abiotic and biotic factors. Climatic and edaphic conditions interact with fertilization and changes in natural disturbance regimes to sculpt community composition and ultimately ecosystem functioning^{5,10,11,19,20}. Given the complex sets of interactions that may explain leaf trait responses to short-term environmental change, a modelling approach is necessary to discern interactions that may otherwise be missed when using traditional bi-variate analyses^{21,22}.

In a global experimental test, we quantified how leaf traits in grasslands change in response to the addition of soil nutrients (i.e., N, P and K) and the exclusion of vertebrate herbivores. We sampled leaf traits from the Nutrient Network (NutNet)²³ cross-continental distributed experiment established at 27 sites (Fig. 1, Supplementary Table 1). This experimental network allowed us to test how commonly measured leaf traits respond to environmental change across grasslands. At the majority of sites, we sampled leaf traits after three to four years of treatment (five sites after two years and 22 of the 27 sites after three to four years; see Supplementary Table 1 for detailed information on each site).

At each site, three blocks of ten 5 m x 5 m plots were established, and two experiments initiated: 1) a full factorial nutrient addition experiment, including the addition of all factorial combinations of N, P and $K_{+\mu}$, where the subscript $+\mu$ refers to the inclusion of ten other micronutrients in the first application year as part of the K addition treatment (see Borer et al.²³ and Methods for more detail), and 2) a combination full nutrient addition (NPK $_{+\mu}$ addition) and herbivore exclusion experiment where fences were built to exclude vertebrate herbivores that were larger in weight than 50 g (for more details see Methods).

Relative cover was visually estimated before the experiment began and prior to the leaf harvest period, when leaf traits were collected from the three to five most dominant species in each plot. Overall, 243 species were sampled across the 27 sites, including grasses, forbs and legumes, and 2664 leaf samples were measured for leaf area, leaf dry weight, and leaf N, P and K concentrations²⁴. Overall the sampled species accounted for 26% of the total vegetation cover at the time when leaves were collected. The effect sizes of the mean leaf trait values for all species in response to the experimental treatments were estimated using multilevel regression models in a hierarchical Bayesian framework using integrated nested Laplace approximation²⁵, where the random effect structure included block nested in site nested in species. SLA values were log-transformed to meet assumptions of normality in the multilevel regression model.

Results and discussion

We found that SLA did not increase consistently with the treatments. We did, however, find evidence of a small but significant increase in SLA in the NP (mean $log(SLA) = 8.79 \text{ mm}^2/g$) and NPK fertiliser treatments (mean $log(SLA) = 8.81 \text{ mm}^2/g$) compared to the control (mean $log(SLA) = 8.69 \text{ mm}^2/g$), suggesting

simultaneous increases in availability of N and P may be necessary to find consistent increases in SLA in grasslands (Fig. 2a)²⁶. When we considered the variation explained by the random effects in the model, SLA showed the highest variability of any of the measured leaf traits at the site level (Fig. 3: ~75% of the variation in SLA in response to treatments was explained among sites), suggesting variation in SLA may be explained by other local abiotic and biotic factors not included in these models. These results provide a new mechanistic understanding of previous NutNet studies, which found that plant aboveground biomass increased in response to nutrient enrichment and fencing treatments, with the highest increase being recorded in the fencing treatments after just three years ^{27,28}. Our results indicate this increase in plant biomass is not explained by an increase in SLA, but instead may be explained by the number of leaves, stems and other structural elements produced.

N, P and K leaf concentrations increased significantly when the corresponding nutrients were applied as fertiliser (Fig. 2). Previous NutNet studies have found multiple-nutrient constraints on aboveground net primary production, including increased vegetation cover and biomass²⁹. Leaf N concentration also increased in leaves with PK_{+μ} fertilization (Fig. 2b), a likely reflection of the increased availability of N in soils³⁰ and the importance of other nutrient limitations for increasing plant N uptake. Leaf P showed the opposite trend to leaf N and decreased in concentration when either N or NK_{+μ} were applied as fertiliser (Fig. 2c). This trend likely reflects the limited availability of phosphate to plants, because of its high affinity to soil particles³¹, as otherwise we may have found an increase in Leaf P when limitations were lifted by the addition of other essential nutrients²⁶. Leaf K concentration showed the highest variation associated with 'species' random effects (~60%, Fig. 3). The

fencing treatment did not significantly alter leaf nutrient concentrations only when soil nutrient addition was combined with the fencing treatment (Fig. 2).

Our findings of an increase in leaf nutrient concentrations in response to the fertiliser treatments could be explained by intraspecific trait variation (increases shown by the same species over time) and by interspecific changes in dominant species following the application of treatments. After treatment initiation, changes in dominant species were observed at some study sites, whereas little change was observed at other sites. This difference is important because increases in leaf nutrient concentrations could be explained by two mechanisms: 1. current species increase their uptake of nutrients (i.e. intraspecific trait variation)³² and 2. new species are recruited into the dominant class (i.e. interspecific trait variation) as the increased nutrient availability favours their growth and establishment³³. Therefore, we evaluated the effects of temporal species turnover on leaf trait responses. We estimated temporal species turnover using Bray Curtis dissimilarity for the three to five most dominant species in each plot comparing pretreatment species composition with composition when the leaf traits were measured, two to four years later.

Given the global extent of our study sites and the high amounts of variation in leaf traits found at the site level, particularly for SLA (Fig. 3), we also evaluated the effects of climatic conditions and pre-treatment soil nutrient levels. We used structural equation models to examine the influence of these additional possible drivers (see supplementary material for details on model development including Supplementary Fig. 1 to 3). Because we did not find evidence of a leaf trait response to the fencing treatments, we did not further evaluate these treatments, only the nutrient addition treatments. Overall, the R² values for each of the leaf nutrient trait response variables were high, indicating a strong explanatory power of the models;

leaf K had the highest R^2 value and SLA the lowest (leaf N, R^2 = 0.53; leaf P, R^2 = 0.32; leaf K, R^2 = 0.55; SLA, R^2 = 0.11).

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All leaf traits varied with climatic and edaphic conditions (Fig. 4 and Supplementary Fig. 4). The nutrient addition treatments explained considerable amounts of variation in the leaf nutrient contents but not in SLA. Species temporal turnover was positively correlated with leaf nitrogen and potassium contents, but significant correlations were not found with the leaf phosphorus content or SLA. This result shows that a portion of the increase in the leaf nitrogen and potassium contents was explained by interspecific variation, suggesting some selection effect of the addition of these nutrients on species composition; whereas the positive response of leaf phosphorus was explained by intraspecific trait variation. These findings corroborate other studies that have also found considerable amounts of variation in leaf chemical traits are explained by intraspecific variation³². The duration of the nutrient addition treatments (represented as year in Fig 4 and Supplementary Fig. 4) was also positively correlated with species temporal turnover, suggesting that sites with longer treatment durations had higher species turnover. Co-variances among the leaf nutrient contents were high in the structural equation model, but SLA showed the lowest co-variation with all leaf nutrient contents (Supplementary Table 2).

Before trait-based ecological studies can scale the responses of leaf traits from individuals to communities and ecosystems¹⁰, a more definitive understanding of when, where and how to interpret changes in plant trait values is needed. This includes how to match plant traits to appropriate environmental conditions depending on the characteristics of specific ecosystems. This necessitates testing plant trait responses in experimental studies, particularly in relation to local and short-term environmental changes or disturbances⁶. We found using a global common

experimental test of leaf trait responses, that leaf nutrient concentrations responded consistently to short-term nutrient additions, and this response is explained by both changes in dominant species and the ability of current dominant species to take up more nutrients when available. The SLA of the dominant species did not increase consistently in response to short-term nutrient addition treatments. Our findings corroborate a recent meta-analysis that found higher intraspecific variation in leaf nutrients than in morphological traits such as SLA³². Based on these findings, if species composition within treatment plots continues to turn over, we may find a clearer response in SLA.

Contrary to expectations, we found little evidence of a consistent short-term increase in SLA or leaf nutrient concentrations to reduced vertebrate herbivory (fencing treatment). The lack of consistent response to the fencing treatment might be due to variation in vertebrate herbivore pressure at these globally distributed grassland sites. The majority of previous studies that have found a consistent increase in SLA and leaf nutrient concentrations with the exclusion of vertebrate herbivores focused on the impacts of cattle and sheep^{5,35-37}, whose grazing pressure tends to be higher and known for selectivity of plant tissue for increased palatability and nutrition³⁸. Here, only eight of our 27 grasslands included a recent or current history of domestic grazing. Other studies that have excluded wild herbivores have found the strongest increases in SLA and leaf nutrient concentrations, when invertebrate herbivores were also excluded^{11,27,39}; where in this experiment we only excluded vertebrate herbivores.

Our findings have implications for how leaf traits are used to infer responses to local-scale environmental perturbations within grassland ecosystems. SLA should be interpreted carefully when used as a predictor of functional response to environmental change within grasslands. SLA has been found to be a reliable

indicator of plant resource utilization strategies at biogeographical-scales ¹⁹. However, a global-scale experimental test demonstrated that SLA is not a consistent indicator of the short-term response of plants to increased soil nutrients or the exclusion of vertebrate herbivores.

Broad-scale biogeographical trait relationships, such as the worldwide leaf economic spectrum¹⁴, do not necessarily correlate as plant functional responses to short-term disturbance and changing abiotic conditions. Our results show that changes in individual traits, in the same species or because of species turnover, do not necessarily represent a 'common currency' for comparing ecosystem-level responses in grasslands to anthropogenic perturbations. When it comes to dominant plant species, leaf nutrients are responsive to elevated soil nutrients, even across sites characterized by very different climatic and edaphic conditions, and are potentially more consistent plant functional response traits than SLA, particularly in the short-term.

Methods

286 Network of experimental sites

The 27 study sites are part of the Nutrient Network, a cooperative globally distributed experiment (Fig. 1 and Table S1 in Supporting Information, http://www.nutnet.org/). Each experimental site had a randomized block design, and at most sites, three replicate blocks divided of ten 5 m x 5 m plots were established, resulting in a total of 30 plots per site.

We quantified climatic variables (mean annual temperature, mean annual precipitation, temperature variation which is a measure of seasonality (calculate as the standard deviation * 100), precipitation variation which is a measure of seasonality (calculated as the coefficient of variation) for each site using modelled values sourced

from the WorldClim Global Climate database (version 1.4;

http://www.worldclim.org). The sites included in this study represented a wide range of climatic conditions with mean annual temperatures ranging from 0.3 °C (alpine grassland in Switzerland) to 18.4 °C (semi-arid C₄ perennial grassland in Australia) and mean annual precipitation ranging from 262 mm (shrub steppe in the USA) to 1898 mm (montane grassland in the USA).

Nutrient addition experiment

In this experiment, we established a set of nutrient addition treatments that included a full factorial combination of three essential plant macronutrients (N, P, $K_{+\mu}$), including a control. The following rates of nutrients, obtained from the same chemical sources, were applied at all sites: 10 g N m⁻² yr⁻¹ as timed-release urea, 10 g P m⁻² yr⁻¹ as triple super phosphate, and 10 g K m⁻² yr⁻¹ as potassium sulphate plus a once-off addition (100 g m⁻² yr⁻¹) of macro- and micro-nutrients (i.e., Fe, S, Mg, Mn, Cu, Zn, B, Mo, Ca). At all sites, N, P, and K fertilisers were applied annually, whereas micro-nutrients were applied once at the start of the study to avoid toxicity and only in treatments that included K. Sites entered the NutNet in different years (2007-2014) and usually measured leaf traits after 3-4 years of nutrient addition (Table S2). Note that ammonium nitrate was used in 2007 at some sites before switching to urea because of increasing difficulty in sourcing ammonium nitrate globally. At a subset of these sites, we tested whether this one-year addition of ammonium nitrate would influence the outcomes of the plant community responses and found no significant effect of nitrogen source²³.

To quantify soil nutrients during the pre-treatment year, we first removed the litter and vegetation from the soil surface and then collected two soil cores (2.5 cm in diameter and 10 cm deep) from each plot. The plot subsamples were composited,

homogenized, and air-dried. The Ecosystems Analysis Laboratory at the University of Nebraska assayed the soils to determine C (%) and N (%) using dry combustion GC analysis (COSTECH ESC 4010 Elemental Analyzer, Costech Analytical Technologies, Valencia, California, USA). Extractable soil P and K and soil pH were assayed at A&L Analytical Laboratory (Memphis, TN). Soil pH was measured using a 1:1 soil to water slurry.

Nutrient addition and herbivore exclusion experiment

The vertebrate herbivore exclusion treatment was established by fencing two plots within each of the blocks. We designed the fences to exclude large aboveground mammalian herbivores, including ungulates, across a diverse range of grasslands characterized by different herbivores²³. At most sites, the height of the fences was 180 cm, and the fence design included wire mesh (1-cm holes) across the first 90 cm in addition to a 30-cm outward-facing flange stapled to the ground to exclude burrowing animals; climbing and subterranean animals could potentially have accessed these plots.

Cover sampling within treatment plots

At peak biomass, species areal cover was visually estimated using a modified Daubenmire method⁴⁰, where cover is estimated to the nearest 1% within one 1-m² sub-plot in each plot. Cover was estimated independently for each species, so the total summed cover may have exceeded 100% for multilayer canopies. In the year when leaf traits were measured at each site (usually after three years of treatment), we used the cover data to identify the top three to five species (although the eight most dominant species were sampled at one site) in each plot to measure leaf traits. We chose to identify the most dominant species in each plot rather than across each site

because we wanted to capture the full range of spatial variation in composition and responses to the treatments, including species turnover.

Leaf trait collection and trait analyses

For each species selected for leaf trait analysis in each plot, we randomly selected five fully developed leaves with little to no signs of herbivore damage from five mature individuals. Sampling followed the standardized protocols detailed by Cornelissen et al.²⁴. All leaves from each species in each plot were combined to measure leaf area. Depending on the resources available at each site, leaf area (mm²) was measured using various leaf area meters or using a flatbed scanner (Epson perfection V300) and image analysis software ImageJ; ⁴¹. Thereafter, all leaves were dried at 60 °C for 48 h and then weighed (dry weight; g). SLA was calculated as leaf area divided by dry weight. SLA was calculated for all five leaves collected from each species in each plot at every site.

Dried leaves were then ground, bulked per plot and per species and analysed for leaf nutrient concentrations. The leaf nitrogen content was determined using a LECO TruMac, which is based on a combustion technique that uses thermal conductivity relative to pure gas; the leaf nitrogen content is determined and is considered accurate to within 1%. The leaf potassium, and phosphorus concentrations were determined using laser ablation ICPMS after Duodu et al.⁴² with the following exceptions: the internal standard was not added but was measured C, the most abundant naturally occurring element was used, and no extra pulverizing was performed beyond that required for C and N analysis, which consisted of placing a sample and a 2-mm-diameter tungsten carbide ball inside 2-mm plastic centrifuge vials, followed by grinding for 15 min using a TissueLyser©. Leaves (approximately 0.2 g) were compressed in a hydraulic dye, which produced a pellet approximately 5 mm across

and 2 mm tall. These pellets were glued to a plastic tray in groups of ~100 and were placed inside the laser chamber. A New Wave 193-nm excimer laser with a True-line cell was connected to an Agilent 8800 ICPMS. The laser beam was 65 microns in diameter and was rastered across a length of approximately 500 microns for approximately 50 seconds, five times per sample with a 30-second washout or background between rasters. The laser fluence at the laser exit was approximately 2 J/cm², and the repetition rate was 7 Hz. The reference material was NIST NBS peach leaves⁴³, and NIST NBS spinach⁴⁴ was used as a monitoring standard; these were analysed every three samples (15 rasters) for moderately close sample-standard bracketing. The average and standard deviation of each element in each sample were calculated and reported after the method presented by Longerich et al.⁴⁵ using Iloite data reduction software.⁴⁶

382 Data analyses

Hierarchical Bayesian multilevel regression models

We developed multilevel regression models in a hierarchical Bayesian framework. All analyses were run using the integrated nested Laplace approximation (INLA²⁵) interfaced with the R statistical computing package (v. 3.3.2) ⁴⁷. The default priors in INLA were used for all analyses, which included the normal distribution specified as N (mean, precision), fixed effects: intercept = N (0,0), slopes = N (0,0.001), and variances modelled as log-precision with priors of log-gamma (1, 5e-5), which was specified as log-gamma (shape, inverse-scale). The random effect structure was constructed to reflect the design of the experiment, and its structure was fixed for all models, regardless of whether each component explained a significant source of variability.

We ran separate models for each of leaf trait (i.e., specific leaf area, leaf N, P and K concentrations), where y_{ijkl} denoted the response, and $\mathbf{x}_{jk} = (x_{1jk}, x_{2jk}, ..., x_{pjk})$ denoted the ith observation from the jth block at the kth site of the lth plant species (Fig. M1). Specific leaf area was log transformed to meet assumptions of normality. Models were constructed as follows:

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$$y_{iikl} \sim N(\mu_{ikl}, \sigma^2)$$
,

400 where
$$y_{ijkl} = \mu_{jkl} + u_l + v_{kl} + w_{jkl} + e_{ijkl}$$

401
$$\mu_{jkl} = \beta_0 + \beta_1 x_{1jk} + \beta_2 x_{2jkl} + ... + \beta_p x_{pjkl},$$

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$$u_l \sim N(0, \sigma^2_u),$$

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$$v_{kl} \sim N(0, \sigma^2_{\nu}),$$

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$$w_{ikl} \sim N(0, \sigma^2_w)$$
, and

405
$$e_{ijkl} \sim N(0, \sigma^2_e)$$
 such that $\sigma^2_u + \sigma^2_v + \sigma^2_w + \sigma^2_e = \sigma^2$,

where μ_{jkl} is the fixed effects associated with species l and block j at site k, β_0 is an estimate of the model intercept, and β_p represents the slope estimates for each linear predictor, i.e., x_{pjkl} . In addition, u_l is the random effect associated with the lth species, v_{kl} is the random effect associated with the kth site (within species l), w_{jkl} is the random effect associated with the jth block (within species l and site k), and e_{ijkl} is the residual error associated with the jth response of block j at site k for species l.

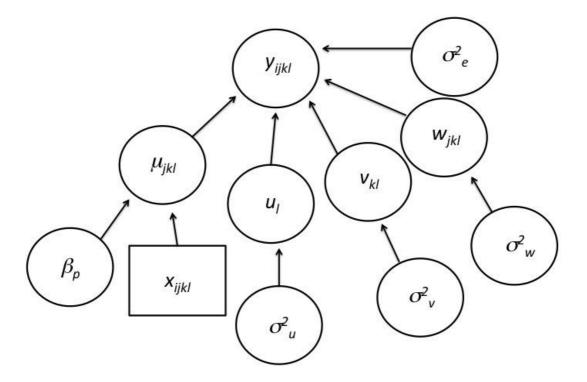


Fig. M1: Directed acyclic graph (DAG) used to represent the multilevel regression models in a hierarchical Bayesian framework for the overall model networks that were developed for both the nutrient addition experiment, and the nutrient addition and herbivore exclusion experiment.

Once a model was fit, residual plots were inspected for any potential relationships in the data that may not have been captured by the model (residuals were calculated as the observed value of the data minus the posterior mean prediction). Plots of the cross-validated probability integral transform (PIT⁴⁸) for each model were also inspected. PIT values provide estimates of the probability that the prediction is less than or equal to the corresponding observed data point, conditional on all other data. A histogram and normal quantile-quantile plot of these values were used to assess the calibration of out-of-sample predictions⁴⁹. If the residual and PIT plots were reasonable, then it was concluded that the model provided a satisfactory fit to the data.

Structural equation models

We began with an initial meta-model (Supplementary Fig. 2) based on a priori expert knowledge and the literature. To correct for the nested experimental design, we included a stratified independent design with blocks nested within sites as stratified variables. We used modification indices⁵⁰ to standardize our decisions of adding missing paths to the model. We used the "modindices" function in the lavaan package⁵⁰, which provides a list of all missing path regressions between two variables in the model, as well as the expected effect of the addition on the model data fit (Chisquare value). We used the modification indices in a stepwise approach, adding ecologically sound paths one at a time, until no modification indices were higher than 2. This incremental process led to the creation of 18 different models. We then scanned path regressions and pruned all non-significant ones (based on p < 0.05), generating a final 19th model. Among the 19 competing models, 13 had a significant model-data fit (estimated by maximum likelihood⁵⁰). To optimize the information-parsimony trade-off, we compared those 13 models using the Akaike information criterion⁵¹.

The selected best model had an AICc difference > 5 with respect to the closest model and an AICc weight of 0.77. To correct for the nested experimental design, we included a stratified independent design with blocks nested within sites as stratified variables. Using the lavaan.survey package, we extracted a robust test statistic (pseudo-maximum likelihood = 23.35, 32 model degrees of freedom, and P = 0.867), indicating a good model-data fit. All analyses were run using R 3.3.2.

Data availability: The data that support the findings of this study are available from the corresponding author upon request.

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467	References			
468	1	Rockstrom, J. et al. A safe operating space for humanity. Nature 461, 472-475		
469		(2009).		
470	2	Ripple, W. J. et al. Collapse of the world's largest herbivores. Scientific		
471		Advances 1, e1400103 (2015).		
		Advances 1, 61400103 (2013).		
472	3	Estes, J. A. et al. Trophic downgrading of plant earth. Science 333, 301-306		
472473	3			
	3	Estes, J. A. et al. Trophic downgrading of plant earth. Science 333, 301-306		
473		Estes, J. A. et al. Trophic downgrading of plant earth. Science 333, 301-306 (2011).		

476 5 McIntyre, S. The role of plant leaf attributes in linking land use to ecosystem 477 function in temperate grassy vegetation. Agriculture, Ecosystems and 478 Environment 128, 251-258 (2008). 479 6 Funk, J. L. et al. Revisiting the Holy Grail: using plant functional traits to 480 understand ecological processes. *Biological Reviews* **1153-1176**, 1-18 (2017). 481 7 Shipley, B., Vile, D. & Garnier, E. From plant traits to plant communities: A 482 statistical mechanistic approach to biodiversity. Science 314, 812-814, 483 doi:10.1126/science.1131344 (2006). 484 8 Messier, J., McGill, B. J. & Lechowicz, M. J. How do traits vary across 485 ecological scales? A case for trait-base ecology. *Ecology Letters* **13**, 838-848 486 (2010).487 9 Hooper, D. U. et al. Effects of biodiversity on ecosystem functioning: a 488 consensus of current knowledge. Ecological Monographs 75, 3-35 (2005). 489 Lavorel, S. & Garnier, E. Predicting changes in community composition and 10 490 ecosystem functioning from plant traits: revisiting the Holy Grail. Functional 491 Ecology 16, 545-556 (2002). 492 Firn, J., Schuetz, M., Nguyen, H. & Risch, A. C. Herbivores sculpt leaf traits 11 493 differently in grasslands depending on life form and land-use histories. 494 Ecology 98, 239-252 (2017). 495 12 Suding, K. N. et al. Scaling environmental change through the community-496 level: a trait-based response-and-effect framework for plants. *Global Change* 497 Biology 14, 1125-1140, doi:10.1111/j.1365-2486.2008.01557.x (2008). 498 13 Wright, I. J. et al. Assessing the generality of global leaf trait relationships. 499 New Phytologist 166, 485-496, doi:10.1111/j.1469-8137.2005.01349.x (2005).

500	14	Wright, I. J. et al. The worldwide leaf economics spectrum. Nature 428, 821-
501		827, doi:10.1038/nature02403 (2004).
502	15	Westoby, M. & Wright, I. J. Land-plant ecology on the basis of functional
503		traits. Trends in Ecology & Evolution 21, 261-268,
504		doi:10.1016/j.tree.2006.02.004 (2006).
505	16	Diaz, S. et al. The plant traits that drive ecosystems: Evidence from three
506		continents. Journal of Vegetation Science 15, 295-304 (2004).
507	17	Garnier, E. et al. Assessing the effects of land-use change on plant traits,
508		communities and ecosystem functioning in grasslands: a standardized
509		methodology and lessons from an application to 11 European sites. Annals of
510		Botany 99 , 967-985 (2007).
511	18	Jarrell, W. M. & Beverly, R. B. The dilution effect in plant nutrition studies.
512		Advances in Agonomy 34 , 197-224 (1981).
513	19	Dwyer, J. M., Hobbs, R. J. & Mayfield, M. M. Specific leaf area response to
514		environmental gradients through space and time. <i>Ecology</i> 95 , 399-410 (2014).
515	20	Leishman, M. R., Haslehurst, T., Ares, A. & Baruch, Z. Leaf trait relationships
516		of native and invasive plants: community- and global-scale comparisons. New
517		Phytologist 176, 635-643 (2007).
518	21	Grace, J. B. et al. Guidelines for a graph-theoretic implementation of
519		structural equation modeling. <i>Ecosphere</i> 3 , doi:10.1890/ES1812-00048.00041
520		(2012).
521	22	Grace, J. B. et al. Integrative modelling reveals mechanisms linking
522		productivity and plant species richness. Nature 529, 390 (2016).
523	23	Borer, E. T. et al. Finding generality in ecology: A model for globally
524		distributed experimens. <i>Methods in Ecology and Evolution</i> 5 , 65-73 (2014).

525	24	Cornelissen, J. H. C. et al. A handbook of protocols for standardised and easy
526		measurement of plant functional traits worldwide. Australian Journal of
527		Botany 51 , 335-380 (2003).
528	25	Rue, H., Martino, S. & Chopin, N. Approximate Bayesian inference for latent
529		Gaussian models by using integrated nested Laplace approximations (with
530		discussion). Journal of the Royal Statistical Society, Series B 71, 319-392
531		(2009).
532	26	Elser, J. J. et al. Global analysis of nitrogen and phoshorus limitation of
533		primary producers in freshwater, marine and terrestrial ecosystems. <i>Ecology</i>
534		Letters 10, 1135-1142 (2007).
535	27	Peeter, P. J. Correlations between leaf structural traits and the densities of
536		herbivorous insect guilds. Biological Journal of the Linnean Socity 77, 43-65
537		(2002).
538	28	Borer, E. T., Grace, J. B., Harpole, W. S., MacDougall, A. S. & Seabloom, E.
539		W. A decade of insights into grassland ecosystem responses to global
540		envirnmental change. Nature Ecology and Evolution 1, 1-8 (2017).
541	29	Fay, P. A. et al. Grassland productivity limited by multiple nutrients. Nature
542		plants 1, 1-5, doi:DOI:10.1038/NPLANTS.2015.80 (2015).
543	30	Galloway, J. N. et al. The Nitrogen Cascade. Bioscience 53, 341-356 (2003).
544	31	Lynch, J. P. & Brown, K. M. Topsoil foraging—an architectural adaptation of
545		plants to low phosphorus availability. Plant and Soil 237, 225-237 (2001).
546	32	Siefert, A. et al. A global meta-analysis of the relative extent of intraspecific
547		trait variation in plant communities. Ecology Letters doi:10.1111/ele/12508
548		(2015).

549	33	Albert, C. H. et al. Intraspecific functional variabillity: extent, structure and
550		sources of variation. Journal of Ecology 98, 604-613 (2010).
551	34	Vitousek, P. M., Porder, S., Houlton, B. Z. & Chadwick, O. A. Terrestrial
552		phosphorus limitation: mechanisms, implications and nitrogen-phosphorus
553		interactions. Ecological Applications 20, 5-15 (2010).
554	35	Cingolani, A. M., Posse, G. & Collantes, M. B. Plant functional traits,
555		herbivore selectivity and response to sheep grazing in Patagonian steppe
556		grasslands. Journal of Applied Ecology 42, 50-59 (2005).
557	36	Firn, J., Prober, S. M. & Buckley, Y. M. Plastic traits of an exotic grass
558		contribute to its abundance but are not always favourable. PLoS One 7,
559		e35870 (2012).
560	37	Dorrough, J., Ash, J. & McIntyre, S. Plant responses to livestock grazing
561		frequency in an Australian temperate grassland. Ecography 27, 798-810
562		(2004).
563	38	Whalley, R. D. B. Grassland regeneration and reconstruction: the role of
564		grazing animals. Ecological Management and Restoration 6, 3-4 (2005).
565	39	Lind, E. M., Myron, E. P., Giaccai, J. & Parker, J. D. White-tailed deer alters
566		specialist and generalist insect herbivory through plant traits. <i>Environmental</i>
567		Entomology 41, 1409-1416 (2012).
568	40	Daubenmire, R. A canopy-coverage method of vegetation analysis. Northwest
569		Science 33 , 43-64 (1959).
570	41	Abramoff, M. D., Magalhaes, P. J. & Ram, S. J. "Image processing with
571		ImageJ. Biophotonics International 11, 36-42 (2004).
572	42	Duodu, G. O., Goonetilleke, A., Allen, C. & Ayoko, G. Determination of
573		refractive and volatile elements in sediment using laser ablation inductively

574		coupled plasma mass spectrometry. Analytica Chimica Acta 898, 19-27
575		(2015).
576	43	USA National Institute of Standards and Technology. (Gaithersburg, MD
577		20899, 2017).
578	44	USA National Institute of Standards and Technology. (Gaithersburg, MD
579		20899, 2014).
580	45	Longerich, H. P., Jackson, S. E. & Gunther, D. Laser Ablation Inductively
581		Coupled Plasma Mass Spectrometric Transient Signal Data Acquisition and
582		Analyte Concentration Calculation*. Journal of Analytical Atomic
583		Spectrometry 11, 899-904 (1996).
584	46	Paton, C. et al. Improved laser ablation U-Pb zircon geochronology through
585		robust downhole fractionation correction. Geochem. Geophys. Geosyst. 11,
586		doi:10.1029/2009GC002618 (2010).
587	47	R Development Core Team. R: language and environment for statistical
588		computing., (http://www.R-project.org/ , Vienna, Austrria, 2013).
589	48	Dawid, A. P. Statistical theory: The prequential approach. Journal of the
590		Royal Statistical Society, Series A 147, 278-292 (1984).
591	49	Czado, C., Gneiting, T. & Held, L. Predictive model assessment for count
592		data. Biometrics 65, 1254-1261 (2009).
593	50	Rosseel, Y. lavaan: An R package for structural equation modelling,. <i>Journal</i>
594		of Statistical Software 48, 1-36 (2012).
595	51	Burnham, K. P. & Anderson, D. R. Model selection and mult-model inference:
596		a practical information-theoretic (Springer, 2002).
597		
598		