

Extremely Small Amounts of B[a]P Residues
Remobilised in Long-term Contaminated Soils: A
Strong Case for Greater Focus on Readily
Available and not Total-extractable Fractions in
Risk Assessment

Anthony C. Umeh^{a, b}, Luchun Duan^{a, b}, Ravi Naidu^{a, b}, Kirk T. Semple^c*

^a Global Centre for Environmental Remediation (GCER), University of Newcastle,
Callaghan, NSW 2308, Australia

^b Co-operative Research Centre for Contamination Assessment and Remediation of the
Environment (CRC CARE), University of Newcastle, Callaghan, NSW 2308, Australia

^c Lancaster Environment Centre, Lancaster University, Lancaster, LA1 4YQ, United
Kingdom

ABSTRACT

There is a lack of understanding about the potential for remobilisation of polycyclic aromatic hydrocarbons (PAHs) residues in soils, specifically after the removal of readily available fractions, and the likelihood to cause harm to human and environmental health. Sequential solvent extractions, using butanol (BuOH), dichloromethane/acetone, and methanolic saponification were used to investigate the time-dependent remobilisation of B[a]P residues in aged soils, after removal of readily available or total-extractable fractions. After 120 d of aging, BuOH-remobilised B[a]P were small or extremely small ranging from 2.3 ± 0.1 mg/kg to 4.5 ± 0.5 mg/kg and from 0.9 ± 0.0 mg/kg to 1.0 ± 0.1 mg/kg, after removal of readily available and total-extractable fractions, respectively. After removal of readily available fractions, the remobilisation rates of B[a]P residues were constant over 5 re-equilibration times, as shown by first-order kinetics. The amounts of B[a]P remobilised significantly ($p < 0.05$) decreased with aging, particularly in hard organic carbon-rich soils. After 4 years of aging, BuOH- and total-remobilised B[a]P were generally $< 5\%$ of the initially spiked 50 mg/kg. Based on the findings of this study, the potential or significant potential for B[a]P NERs in soils to cause significant harm to human and environmental health are minimal.

Keywords: Polycyclic aromatic hydrocarbons, B[a]P, Nonextractable residues, Small Remobilisation, Release, Risk assessment

1. Introduction

The impacts and significance of nonextractable residues (NERs) of hydrophobic organic contaminants (HOCs), such as polycyclic aromatic hydrocarbons (PAHs), to human and environmental health risk assessments of contaminated land are gaining increasing attention within the scientific and regulatory communities¹⁻⁴. Specifically, whether NERs will be remobilised in soil and pose unacceptable levels of risks to human and environmental health remain uncertain.

Sixteen priority PAHs are of global concern as they are ubiquitous and persistent in soils, and are potentially toxic and/or carcinogenic. Benzo[a]pyrene (B[a]P) is classified as a Group 1 carcinogen and is a key risk-driver for many PAH-contaminated sites^{5,6}. The potential for B[a]P to persist in soils is predictable given its high Log K_{ow} , very low aqueous solubility and volatility, as well as recalcitrance to biodegradation^{5,7}. The persistence and sequestration of PAHs are key features governing their fate in soils and may affect the biological functions and productivity of soils. PAHs are subjected to various physical, biological, environmental loss and sequestration processes⁸, making it difficult to determine total PAH concentrations in soils, particularly for retrospective risk assessments. Non-exhaustive extractions of PAHs aged in soils often reveal that PAH desorption may be bi- or tri-phasic comprising a rapid initial loss phase, and then slow and/or very slow phases that remains constant over time under undisturbed environmental conditions⁹. Slow desorption of PAHs in soil is attributed to mass-transfer limitations resulting from PAH sequestration onto and into rubbery or glassy soil organic matter, clay-organic matter matrices and carbonaceous materials (such as black carbon), as well as diffusion into micro- and nano-pores^{10,11}. Detailed sequestration mechanisms are described elsewhere^{12,13}.

Highly sequestered PAH fractions in soils may result in the failure of remediation objectives, particularly where associated concentrations exceed regulatory guideline values¹⁴.¹⁵ Regulatory concerns regarding the potential risks of these sequestered residues to biological receptors may be justified^{16,17}. The highly sequestered PAH fraction is usually defined as that which is not extractable by exhaustive solvents¹⁸; hence, the term 'NERs'. Previous studies have reported that a fraction of PAH NERs may be extractable by methanolic saponification of pre-extracted soils¹⁹. Methanolic saponification cleaves ester or ether linkages within the soil matrix, thereby partially releasing entrapped or occluded PAH NERs²⁰. According to Kastner et al.¹⁸, entrapped or occluded NERs (parent compounds and metabolites), referred to as Type I NERs, are non-covalently bound to the soil matrix. This implies that although Type I NERs have low to high stability, they may be released or remobilisable at slow rates and small extents²¹. Type II NERs are covalently bound in soils, very stable, and less likely to be released. For PAHs, potential covalent binding of metabolites to the soil matrix may result in the formation of new compounds that are physico-chemically different from the original compound. Since Type II PAH NERs may be distinguishable from their parent compounds, their classification as NERs of original parent compounds may be arguable. Type III NERs are biogenic residues and bear no environmental relevance¹⁸. Biodegradation is not a substantial loss mechanism for hydrophobic and persistent high molecular weight (HMW) PAHs, such as B[a]P; hence, the bulk of total NER in soils from HMW PAHs may be Type I. A lack of understanding of soil-PAH interactions and the nature of PAH NERs in soil is a major uncertainty constraining risk-based approaches to contaminated land management, and may lead to over- or under-estimation of potential health risks that may be associated with exposure to PAH NERs in soils.

Many researchers regard highly sequestered HOC-fractions in soil as unimportant in risk assessments²², because associated desorption rates are slow or very slow and soil half-lives may last for decades^{21,23}. In contrast, a few studies have showed potential for release of PAH NERs^{19,24}; however, amounts released and associated implications for risk assessment are not usually considered²¹. In these previous studies, highly sequestered PAH NERs are often considered as residues following exhaustive extraction of soils. Exhaustive extractions of PAH contaminated soils, a common technique in traditional risk assessment approaches, yield total-extractable PAH concentrations considered to be overestimative of risks to human and ecological health^{22,25}. Risk-based approaches, which are based on bioavailability or bioaccessibility concepts, focus on the fractions that are readily available to pose risks to human and ecological health^{25,26}. Till date, there are no published studies on the potential remobilisation of PAH residues in soils resulting from treatments that remove only readily available fractions, rather than those that remove total-extractable fractions. Such studies are required to minimise the uncertainties associated with risk-based approaches to contaminated land management.

The objectives of this study are to investigate: the time-dependent remobilisation of B[a]P residues in soils aged for up to 4 years after the removal of readily available fractions (partial removal); the remobilisation kinetics after equilibrating soils from which readily available fractions have been removed; and to compare the amounts of PAHs remobilised after partial removal and the removal of total-extractable fractions (complete removal) in soils.

2. Experimental

2.1. Chemicals

Analytical grade B[a]P (> 96% purity) was obtained as crystalline solid from Sigma Aldrich Pty Ltd., Sydney, Australia. Also sourced from the same supplier were p-Terphenyl-d14, analytical grade acetone (Ace), acetonitrile (ACN), 1-butanol (BuOH, density = 0.81 g/mL, $\geq 99.4\%$), dichloromethane (DCM), methanol (MeOH, HPLC Grade), toluene (Tol, 99.8%), potassium hydroxide (KOH) and silica sand. Hexane (Hex, HPLC grade) was purchased from Fisher Scientific, Loughborough, UK.

2.2. Design

Three air-dried soils I (Kurosol), M (Ferrosol) and B (Black Vertosol) were collected from a depth of 5 – 20 cm from the Adelaide Hills, South Australia, Mount Tamborine and Beaudesert, Queensland respectively. Soils properties were presented previously²⁷. Soil I is sandy loam (10.7% Clay, 4.3% TOC), soil M is sandy-clay-loam (21.2% Clay, 7.4% TOC) and soil B is sandy-clay-loam (30.9% Clay, 3.5% TOC) (USDA textural classification). The contents (%) of soft- and hard-organic carbon (OC), and black carbon (BC)-rich materials were determined²¹. Air-dried soils were sieved through a 2 mm sieve, spiked with 50 mg/kg B[a]P, and wetted to their field moisture contents (25% to 40% on dry weight basis)²⁷. Only B[a]P spiked soils aged in the dark for 120 d and 4 years were used. Silica sand was concurrently spiked and used as a reference material to monitor spike recovery after aging. Soil subsamples (n = 18 or n = 8) were collected and dried at 37.5 °C after each aging time. For the removal of readily available B[a]P fractions (A in Fig. 1), subsamples of soils (n = 18) were subjected to a mild BuOH extraction following quality control and quality assurance procedures described previously²⁷. The resulting soil residues were then divided into 6 batches (n = 3) according to the design in Fig. 1.

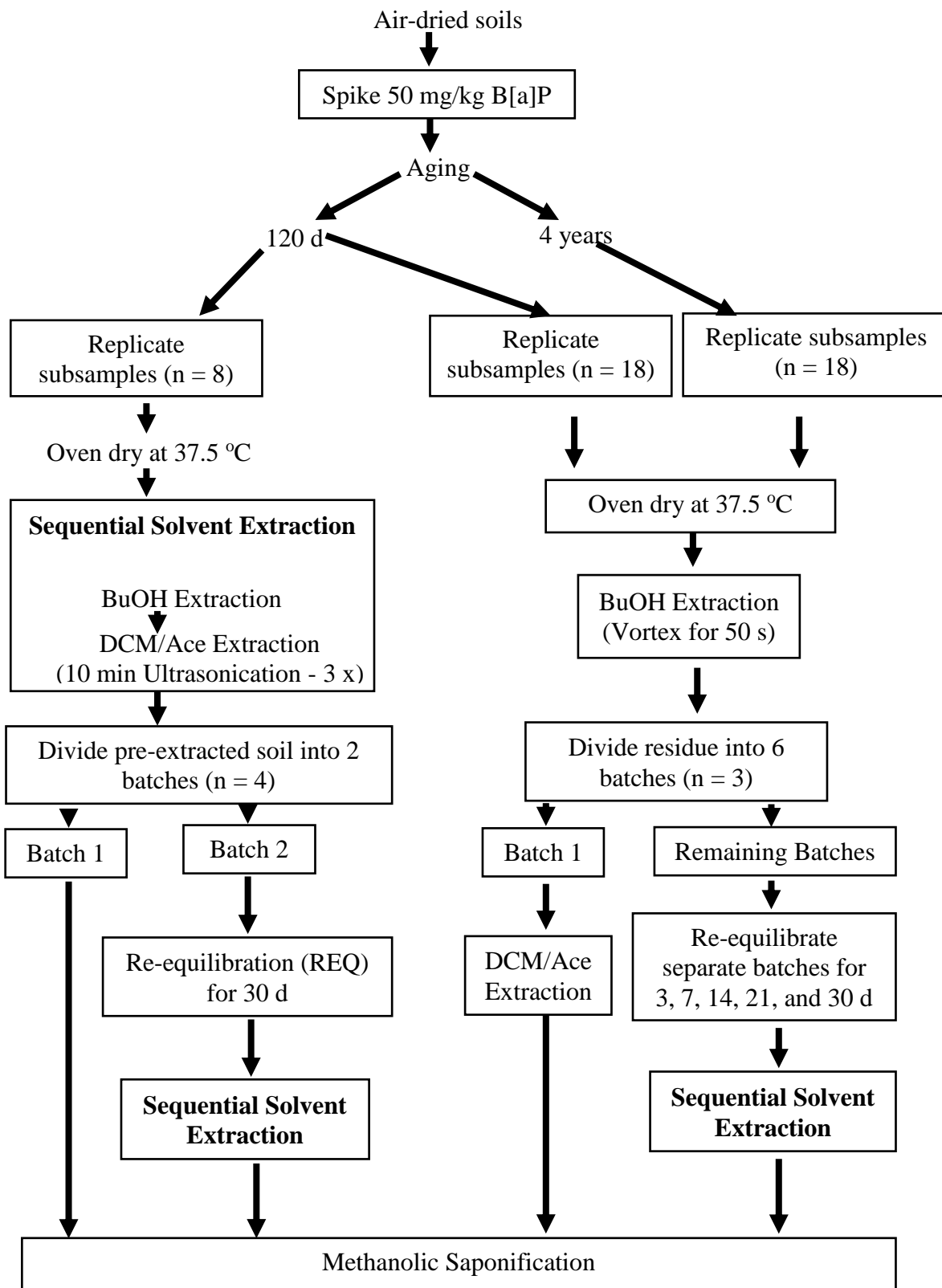


Fig. 1. Experimental design

The replicates did not show significant between-batch differences ($p > 0.05$) in their BuOH extractabilities (data not shown).

The first batch ($n = 3$) of the BuOH-extracted soil samples was subsequently extracted with DCM/Ace to remove B[a]P fractions that were not extractable by BuOH. The pre-extracted soils (NERs after DCM/Ace extraction) were further subjected to methanolic saponification (MeKOH). The remaining batches, i.e. 2, 3, 4, 5 and 6 (Fig. 1), were then kept in the dark to re-equilibrate at 25.5 ± 1.0 °C for 3, 7, 14, 21, and 30 d, respectively, for modelling of remobilisation kinetics.

For the removal of total-extractable B[a]P fractions (B in Fig. 1), only soils aged for 120 d were used. Subsamples ($n = 8$) were sequentially extracted with BuOH and DCM/Ace²⁷. The pre-extracted soils were each divided into 2 batches ($n = 4$). The first batches of soils were extracted with MeKOH, while the second batches of soils were kept in the fume hood overnight and rewetted to field moisture contents, and re-equilibrated in the dark for 30 d.

After re-equilibration (REQ) of soils (A and B in Fig. 1), soils were again sequentially extracted with BuOH and DCM/Ace, and MeKOH. Blank soils were also treated in a similar manner. All percentage extractability calculations were based on the initial amounts of B[a]P spiked into the soil (50 mg/kg), unless otherwise specified.

$$\text{Extractability (\%)} = \left(\frac{\text{Amount of B[a]P Extracted by Solvent (mg)}}{\text{Amount of B[a]P Spiked into Soil (mg)}} \right) \times 100\% \quad (1)$$

Remobilisation refers to the additional amounts of B[a]P extractable by BuOH, or by a combination of BuOH and DCM/Ace (total remobilisation), after REQ. The amounts of B[a]P released after the removal of readily available fractions were fitted to a first-order exponential decrease model as described previously²¹.

2.3. *Determination of Butanol-, Total-, and MeKOH-extractable B[a]P.*

Soils were extracted with 1-BuOH following a previously described procedure with some modifications²⁷. In the modified procedure, the butanolic supernatant was not decanted following centrifugation. Instead, a 2 mL aliquot of the supernatant was transferred into a clean glass vial, vacuum-concentrated, redissolved in ACN, and filtered through 0.45 µm PTFE syringe filters before transfer into 2 mL amber HPLC vials for analysis. After BuOH extraction, soils were extracted with 3 mL DCM/Ace (1:1, v/v)²⁷. Total-extractable B[a]P is the sum of BuOH- and DCM/Ace- extractable B[a]P. The other operational definitions are presented in the SI. After the sequential solvent extractions, NERs were then subjected to MeKOH by extracting soils with 10 mL MeOH/2 M KOH (14:1, v/v)²⁷.

2.4. *HPLC Analysis of B[a]P*

The concentrations of B[a]P were determined with an Agilent 1100 Series HPLC equipped with a fluorescence detector (excitation wavelength = 230 nm and emission wavelength = 460 nm) as in described previously²⁷.

2.5. *Data analysis*

Extractability data were statistically analysed with SPSS (IBM Corporation, version 24) and graphing was by Origin (Microcal Software Inc., Northampton, MA, USA, version 6). No data transformation was applied. There were 3 independent and 2 or more outcome variables. The independent variables included: soil type (3 nominal categories); aging time (2 nominal categories); and the procedures for removing extractable fractions (i.e. partial or complete). The outcome variables were BuOH, DCM/Ace, total, and MeKOH extractability (mg/kg or %). A 95% significance level was used. One-way ANOVA was used to test between-group differences with Tukey's post hoc test where equal variance is assumed and

Games Howell's test where equal variance is not assumed ²⁸. Student's *t* test was used for pairwise comparisons of independent samples.

3. Results and discussion

3.1. Spike recovery and data variability

Spike recovery of B[a]P from silica sand was $> 95\%$ throughout the study, showing that B[a]P losses during laboratory procedures were minimal. Total extractability (i.e. BuOH+DCM/Ace extractability) of B[a]P 2 d (day 0) after spiking ranged from 78.5% (soil B) to 95.9% (soil M) and agreed with a previous study that used same soils^{27, 29}. After MeKOH, complete mass balance ($\geq 100\%$) was obtained, except in soil B where the mass balance was $88.9 \pm 8.5\%$. At 120 d of aging, total extractability ranged from $53.2 \pm 2.2\%$ (soil B) to $80.0 \pm 9.9\%$ (soil M), whereas it ranged from 63% (soil B) to 86% (soil M) after MeKOH. Average standard deviation in B[a]P extractability was generally below 5%. Decreases in the recovery of PAHs due to the effects of aging and sequestration in soils, and variations due to soil properties, are well known²⁹⁻³¹. Other studies have reported comparable decreases in total extractability from 100 to 80% over 200 d of aging using Soxhlet extraction³², and 83 to 73% after 170 d of aging using a DCM-soxtec extraction technique²⁰. In sterile conditions, ¹⁴C-B[a]P in aged soil was not completely recovered²⁰. The decrease in B[a]P extractability with aging was attributed to sequestration effects, although minimal losses from biodegradation may not be disregarded.

3.2. Extractability and aging of B[a]P in soils prior to re-equilibration

All soils showed substantial decrease ($p < 0.05$) in B[a]P extractability after aging. Butanol extractability has been used to assess PAH bioaccessibility in soils for human and ecological health risk assessments^{29, 33-35}. Prior to REQ (day 0), BuOH extractability ranged from 16.2 ± 0.8 mg/kg (soil B) to 26.4 ± 1.0 mg/kg (soil I) at 120 d of aging (Fig. 2A), and 0.28 ± 0.03 mg/kg (soil B), 2.97 ± 0.38 mg/kg (soil I) and 3.02 ± 0.21 mg/kg (soil M) at 4 years of aging. Soil B had significantly lower ($p < 0.05$) B[a]P extractability than the other

soils whose extractabilities were similar ($p > 0.05$). The decreases in B[a]P extractability through aging indicate that B[a]P became progressively sequestered in the soil matrix^{36, 37}. Hard OC and clay-sized particle fractions in soils can sequester PAHs^{29, 32, 38-40}, through physical adsorption of PAHs to surfaces and occlusion or entrapment in pores^{10, 13, 29, 41}. In this study, the fractions of hard OC relative to TOC (hard OC/TOC) in soils (0.45 – 0.58), as well as the contents of very fine-textured particles (10.7 – 30.9%) may account for the differences in B[a]P extractability by BuOH. This is indicated by the strong negative relationships between BuOH-extractable B[a]P and hard OC/TOC, and with clay content of soils ($R^2 > 0.9$) at 120 d and 4 years of aging in agreement with previous studies²¹. These strong relationships need to be validated using a wider range of soils.

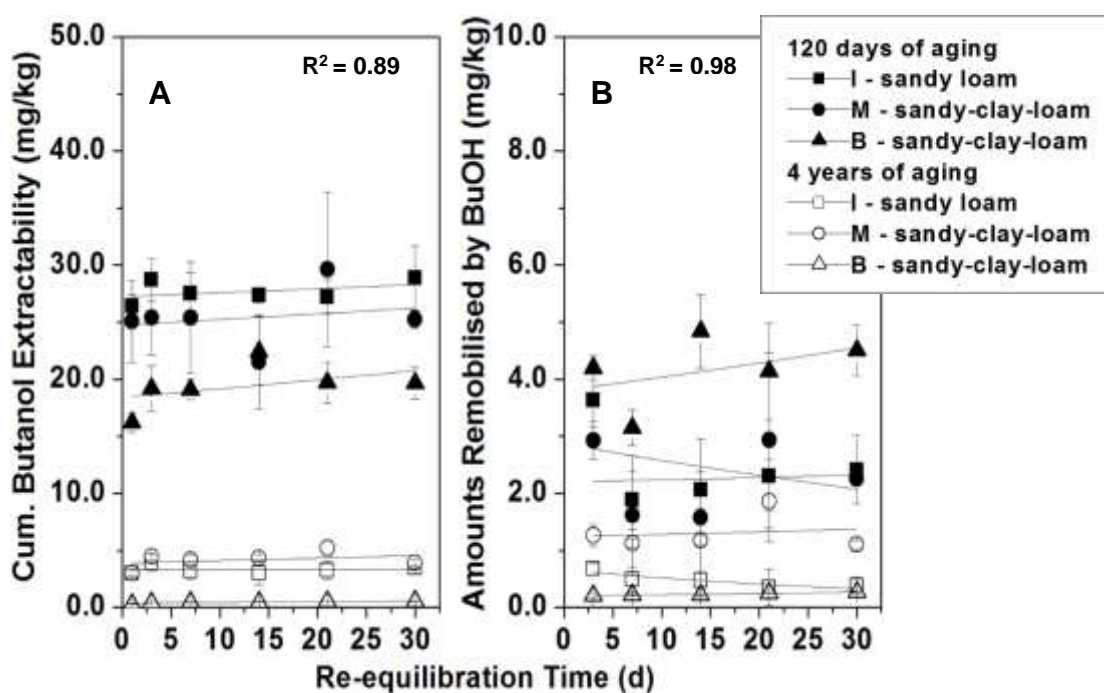


Fig. 2. Exponential curve-fitting of the cumulative amounts of butanol-extractable B[a]P following re-equilibration (A) and the amounts of butanol-remobilised B[a]P after each re-equilibration time (B). Values are means ($n = 3$) \pm standard deviations. Values are means ($n = 3$) \pm standard deviations. Note the differences in the Y-axis.

As with BuOH extractability, DCM/Ace-extractable B[a]P and total extractability (BuOH + DCM/Ace) decreased significantly ($p < 0.05$) after fresh spiking to 120 d of aging, and to 4 years of aging (Fig. 3), whereas B[a]P NERs increased (Fig. 3). The 4 year aged soils had a decrease in the cumulative total extractability of B[a]P ranging from 81% (soil M) to 97% (soil B) of the total-extractable B[a]P at 0 d of aging (Fig. 3). Methanolic saponification released detectable amounts of highly sequestered B[a]P NERs in the pre-extracted soils (Fig. 3). Although soil B contained the largest amount of B[a]P NERs after aging (Fig. 3), the fraction (%) of B[a]P NERs in pre-extracted soils released by MeKOH was consistently smaller in Soil B than in the other soils (Fig. 3), indicating stronger B[a]P sequestration by soil B. Soil B had large amounts of hard OC/TOC fractions and clay, and may be expected to strongly sequester B[a]P compared to the other soils^{10,42}. Strong sequestration of PAHs in soils lead to very low PAH extractability^{20,30}, particularly in long-term aged soils²³. The low total extractability from soil B, 2 d after spiking, further indicates rapid B[a]P sequestration and that the extents of sequestration is dependent on soil properties^{29,32,43}.

3.3. *Remobilisation of B[a]P after the removal of readily available fractions in soils*

The aim of BuOH extraction in the sequential extraction scheme was to partially, rather than completely, remove the extractable B[a]P fractions as in risk-based approaches to contaminated land management^{22,26,44}. The amounts of B[a]P that were then re-extractable or remobilised after the removal of readily available fractions in soils are illustrated in Fig. 2. Cumulative BuOH extractability of B[a]P slightly increased after 2 d of REQ at 120 d and 4 years of aging (Fig. 2A), however, the increases were generally not significant ($p > 0.05$). Cumulative total extractability of B[a]P after 2 d of REQ were not significantly different ($p > 0.05$) from total extractability prior to REQ (Fig. 3). After 2 d of REQ (Fig. 2), cumulative BuOH extractability of B[a]P also remained unchanged ($p > 0.05$), indicating that the amounts of B[a]P remobilised by BuOH plateaued quickly. The constancy of BuOH-remobilised B[a]P

after 2 d of REQ was reflected by the curves fitted with the first-order kinetic model (Fig. 2). The constancy in B[a]P remobilisation further showed that the rates and extents of decrease in the amounts of B[a]P remobilised were unchanged from 2 to 30 d of REQ, further reflecting the stability of the sequestered B[a]P.

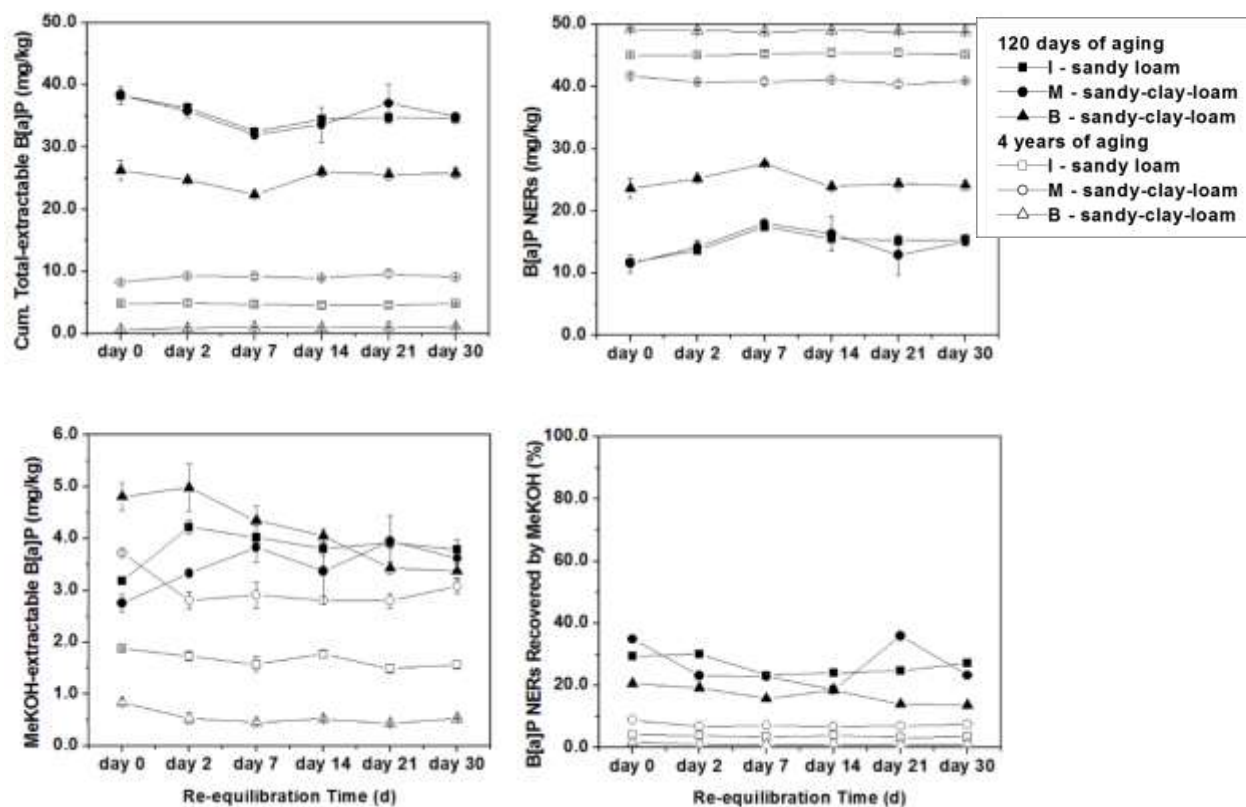


Fig. 3. Temporal changes in B[a]P extractability and sequestration before and after re-equilibration. Cum. is cumulative. NER is nonextractable. MeKOH is methanolic saponification. Values are means ($n = 3$) \pm standard deviations. Note the differences in the Y-axis.

Specifically, BuOH-remobilised B[a]P after 30 d of REQ in soils at 120 d of aging were 4.5 ± 0.5 mg/kg, 2.4 ± 0.6 mg/kg and 2.3 ± 0.1 mg/kg in soils B, I and M, respectively (Fig. 2B). Although the amounts of B[a]P remobilised by BuOH after 120 d in soils were below 9% of the initially spiked concentrations (50 mg/kg) or below 13% of the residue after BuOH extraction prior to REQ, the effects of hard OC fractions and clay content on B[a]P

remobilisation appear anomalously obscured. This anomaly may not be attributed to changes in hard OC- or clay-induced interactions on B[a]P extractability given that hard OC/TOC fractions and clay contents of soils showed negative relationships with B[a]P extractability by BuOH before REQ. Rather, the anomaly may be attributed to the non-exhaustive nature of the BuOH extraction²⁰ as it may underestimate the rapidly desorbing or readily available B[a]P fractions in soils⁴⁵. The extent to which the readily available B[a]P fractions in soil B (16.2 ± 0.8 mg/kg) were removed by BuOH prior to REQ may be less efficient than the extents in the other soils (25.1 ± 3.6 , and 26.4 ± 1.0 mg/kg in soils M and I, respectively). After REQ from 2 to 30 d, the readily available B[a]P fractions in all soils had equilibrated in the residual butanolic solvent (1 mL). Hence, subsequent re-extraction of soil B with BuOH after REQ efficiently captured both the readily available B[a]P that was not extractable prior to REQ and remobilised B[a]P, which may have resulted in the greater B[a]P extractability (remobilisation).

Sorption and desorption of PAHs may be underestimated during extraction of soils that may have not been equilibrated efficiently⁴⁶. In this study, the subsequent DCM/Ace-extractable B[a]P after 30 d of REQ was smaller ($p < 0.05$) in soil B (5.5 ± 0.4 mg/kg) compared to soils I (6.6 ± 0.3 mg/kg) and M (9.2 ± 0.7 mg/kg). Hence, total-remobilised B[a]P after 30 d of REQ ranged from 29 (soil B) – 43% (soil M) of the residues after BuOH extraction prior to REQ. Cumulative BuOH extractability and total B[a]P extractability after REQ at 120 d of aging were also significantly smaller ($p < 0.05$) in soil B than in soils I and M (Figures 2 and 3). In addition, B[a]P NERs in soils recovered by MeKOH before and after REQ at 120 d of aging was smaller ($p < 0.05$) in soil B than in the other soils (Fig. 3), further reflecting stronger B[a]P sequestration of B[a]P in soil B. Further, the fraction of DCM-Ace extractable B[a]P relative to the residues obtained following BuOH extraction was again smaller in soil B (13.6%) than in soils I (28.9%) and M (35.4%) prior to REQ. This suggested that the potential

for B[a]P remobilisation in the sandy-clay-loam soil B was low compared to the other soils. Although the sandy-clay-loam soil M had the largest TOC contents, its fraction of soft OC relative to TOC content (0.58) was larger than in soils I (0.55) and B (0.42). Considering that B[a]P sequestration in soils with large amounts of soft OC are weak^{10,13,32}, the large amounts of extractable B[a]P before REQ and potentially remobilisable B[a]P after REQ in soil M were reasonable. Soils B and I also contain expandable clays (i.e. montmorillonite)²⁹; this may contribute to stronger B[a]P sequestration resulting in reduced B[a]P extractability and potential for remobilisation⁴⁷. Hence, the influence of the quantity and quality of TOC, and clay, in soils should be considered towards understanding the extractability and potential for remobilisation of B[a]P NERs.

At 4 years of aging, the amounts of B[a]P remobilised by BuOH in soils were extremely small ($p < 0.01$) compared to the amounts remobilised after 120 d of aging (Fig. 2). Butanol-remobilised B[a]P were 1.1 ± 0.1 (soil M), 0.4 ± 0.2 (soil I), and 0.3 ± 0.0 mg/kg (soil B). This suggests that only 3% or less of initially spiked B[a]P (50 mg/kg), or of residues after BuOH extraction before REQ, was again re-extractable by BuOH. Total-remobilised B[a]P after 30 d of REQ were 6.0 ± 0.2 mg/kg, 1.9 ± 0.2 mg/kg, and 0.9 ± 0.07 mg/kg in soils M, I and B, respectively. This further suggests that approximately 2 to 13% of initially spiked B[a]P (50 mg/kg), or of residues after BuOH extraction conducted before REQ, was re-extractable. In addition, the fractions of B[a]P NERs recovered by MeKOH after 30 d REQ were again smaller in soil B (1.1%) than in soils I (3.5%) or M (7.5%), as with before REQ (Fig. 3). The extremely small amounts of extractable- and remobilised-B[a]P observed in all 3 soils after 4 years of aging, particularly in soil B, reflect the influence of aging and soil properties on B[a]P sequestration and remobilisation in soils. Slow repartitioning of sequestered B[a]P residues during the re-equilibration periods could explain the small amounts of B[a]P NERs remobilised

in the soils, and further described below. The extents of potential remobilisation of B[a]P may be expected to further decrease or remain constant in soils after extended aging periods ³⁰.

3.4. Comparison of B[a]P remobilisation in soils after the removal of the readily available and total-extractable fractions

The results obtained after 30 d of REQ at 120 d of aging are presented in Fig. 4. Our recent investigations revealed that B[a]P remobilised in soils at 200 d of aging, after removing total-extractable fractions, were very small (0.7 ± 0.0 to 3.3 ± 0.2 mg/kg) and decreased at very fast rates over time ²¹. In this study, the extractable amounts of B[a]P in soils after 4 years of aging before REQ remained very small (Figures 2 and 3), reinforcing the effects of aging on B[a]P extractability ²¹. Investigating the potential remobilisation of B[a]P after removing total-extractable fractions in soils aged for 4 years was not justified; hence, only the results for 120 d of aging and at 30 d of REQ are presented (Fig. 4).

Clearly, the amounts of B[a]P remobilised after the removal of the readily available fractions in soils were greater than those remobilised after the complete removal of extractable fractions (Fig. 4), particularly total-remobilisable B[a]P. This is reasonable since more highly sequestered fractions were removed by the exhaustive extraction prior to REQ,

compared to the mild extraction.

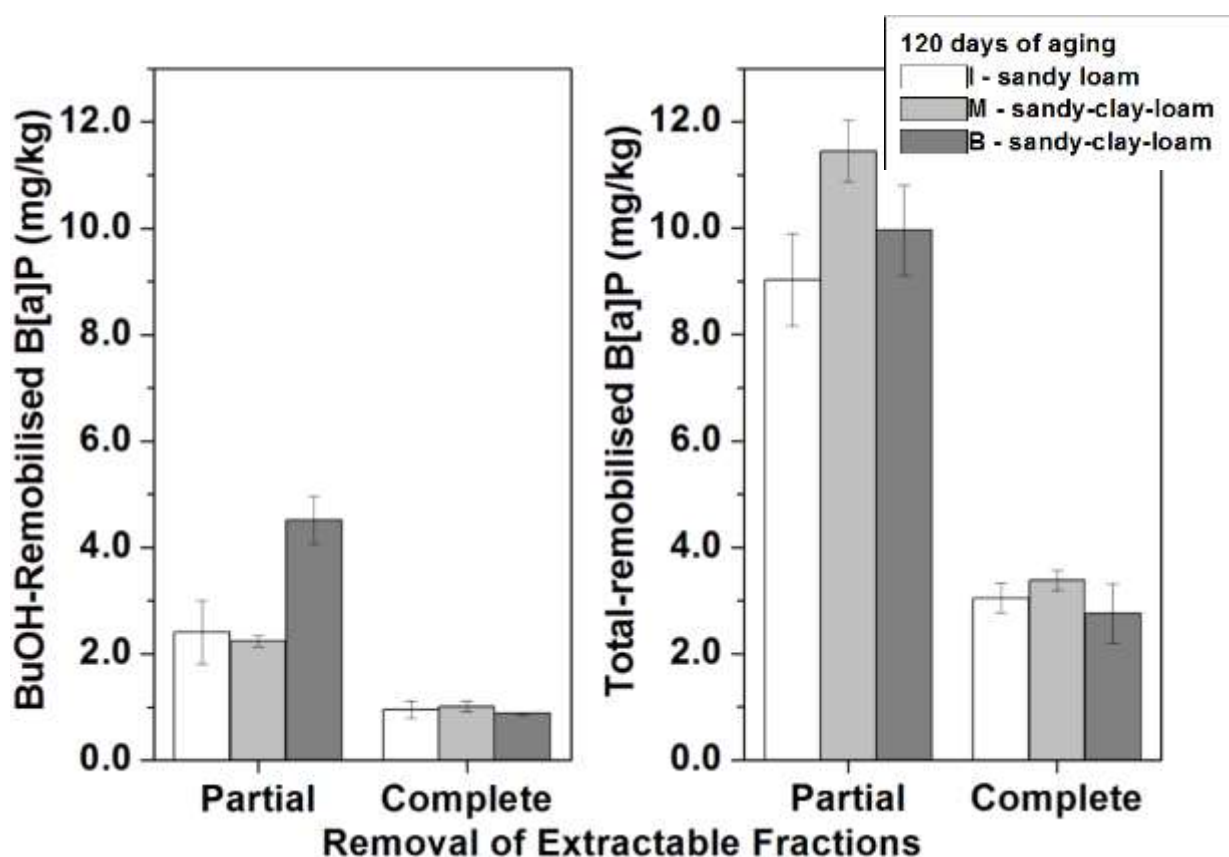


Fig. 4. Amounts of B[a]P remobilised following the removal of readily available and total-extractable B[a]P fractions in soils. Values are means ($n = 3$) \pm standard deviations.

However, the amounts of BuOH-remobilised B[a]P after the removal of readily available fractions in soils at 4 years of aging were similar ($p > 0.05$) to the amounts remobilised after removal of total-extractable fractions in soils at 120 d of aging.

Therefore, these results show that it is realistic to assess the risks of potential B[a]P remobilisation from long-term contaminated soils based on laboratory measurements of readily available concentrations as measured in this study, or the use of other non-exhaustive extractions^{44, 48}. Overall, structured monitoring for potential release of sequestered B[a]P from B[a]P-contaminated sites that are managed by risk-based approaches is suggested as good practice⁴⁹. This is important because the impacts of changing exposure scenarios and

changes in environmental conditions, such as pH, temperature, drying and wetting, freezing and thawing, and changes in land-uses on PAH remobilisation in long-term contaminated soils are not understood.

3.5. Proposed slow repartitioning of B[a]P residues following the removal of the readily available fractions

As illustrated in Fig. 5, B[a]P is weakly sequestered within easily accessible sites in soils (a), such as on clay surfaces and soft OC²⁰, during initial soil-B[a]P contact time and prior to aging and REQ ('I'). Hence, large amounts of B[a]P may be solvent-extractable ('b', 'c', and 'd' in 'I'). Over time ('a' in 'II'), B[a]P accesses more sequestration sites, diffusing into soft OC or may be adsorbed strongly to clay or organo-clay surfaces, and may be entrapped in pores or cracks and adsorbed to flat surfaces of (or collapsed pores within) hard OC or BC-rich materials through π - π interactions¹³. Hence, a mild BuOH extraction of soil removes only weakly sequestered B[a]P⁵⁰, whereas subsequent DCM/Ace extraction of soil ('b' in 'II') removes highly sequestered B[a]P. Harsh solvents penetrate soil pores causing them to swell, and may competitively displace sequestered B[a]P⁵¹. Harsh solvent mixtures, such as DCM/Ace, possess greater pore swelling and stronger competitive displacement capacities than BuOH⁵¹. Hence, total extractability will be greater than BuOH-extractable B[a]P²⁷. Methanolic saponification ('c' in II) partially releases more highly sequestered B[a]P from sites where they are physically entrapped¹⁸. The resulting residue after MeKOH extraction contain B[a]P that is very highly sequestered; this may be more associated with BC-rich materials in soils^{10,12}. We have previously reported a strong positive relationship between hard OC fractions and BC-rich fractions of these soils ($r = 0.98$, $p = 0.02$, $n = 4$), as well as between clay and hard OC/TOC ($r = 0.87$ and $p = 0.13$)²¹; however, contents of clay and hard OC were not multicollinear ($R^2 = 0.26$).

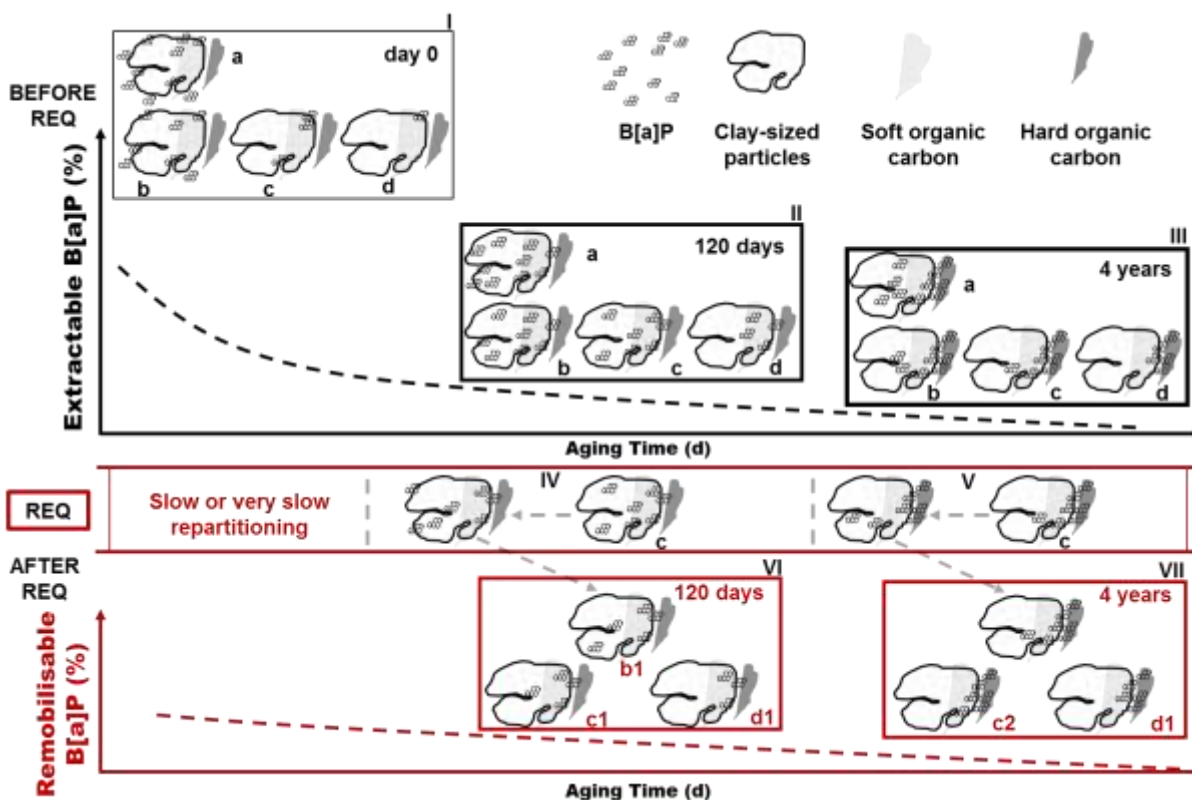


Fig. 5. Proposed slow repartitioning of B[a]P after the removal of readily available fractions. a is B[a]P-contaminated soil before aging; b is soil residue after mild BuOH extraction that removes the readily available fractions; c is NER after DCM/Ace extraction; and d is NER after methanolic saponification of soil.

These observations therefore support the strong B[a]P sequestration in soil B with large amounts of clay and hard OC/TOC fractions. Overall, the amounts of B[a]P extractable by each of the solvents decreased, whereas the non-extractable fraction increased over time due to increasing soil-B[a]P interactions (III).

Following the re-equilibration (IV and V) of the BuOH-extracted soil ('c' in 'II' and 'III'), sequestered B[a]P may repartition slowly into more readily available compartments in soils due to the concentration gradient between sequestered and readily available compartments²¹. Highly sequestered B[a]P repartition at slow rates (V)⁵². After REQ (VI

and VII), small amounts of B[a]P residues become re-extractable by BuOH (i.e. readily remobilisable B[a]P). Subsequently, DCM/Ace extraction of soil ('b1' in VI and VII) recovers B[a]P that is not readily remobilisable. Very small amounts of B[a]P residues ('c1' in VI and VII) may be further released by MeKOH, resulting in residues unlikely to be remobilised¹⁸.

After aging of 120 d and 4 years, strong positive relationships were observed between BuOH-remobilised B[a]P after REQ and DCM/Ace-extractable B[a]P prior to REQ ($r = 0.88$, $p = 0.02$), as well as with MeKOH-extractable B[a]P ($r = 0.78$, $p = 0.07$) (Fig. S1). Hence, total-remobilised B[a]P showed a stronger relationship with MeKOH-extractable B[a]P after REQ than before REQ (Fig. S2). These relationships suggest that B[a]P remobilisation may be due to repartitioning of sequestered B[a]P during REQ. Overall, the extremely small amounts of B[a]P remobilised, particularly following extended aging (4 years), indicates a likelihood for very low remobilisation extents in long-term aged soils.

3.6. Implications for contaminated land decision-making

Risk-based approaches to contaminated land management is gaining ground in a number of countries^{22, 53, 54}. However, not all regulators of contaminated lands agree to adopting risk-based approaches that rely on contaminant bioavailability or bioaccessibility due to associated uncertainties⁵⁵, such as the fate of contaminant residues in soils⁴. In this study, B[a]P remobilisation occurred in soils after 4 years of aging, but the amounts remobilised were extremely small (generally < 5% of initially spiked concentration, i.e. 50 mg/kg) and may be insignificant from a risk assessment perspective. For instance, considering the Australian health investigation levels (HILs) of at least 3 mg/kg B[a]P in soils⁵⁶, all contaminated soils used in this study passed the criteria based on the amounts of BuOH-remobilised B[a]P from soils after 4 years of aging. Generally, the HIL criteria was also passed where total-remobilised B[a]P was used in the soils. In this regard, the potential

risks to human and environmental health are minimal and acceptable. These observations are a strong case for greater focus on readily available B[a]P fractions, rather than total-extractable fractions, in the risk assessment of long-term B[a]P-contaminated soils.

4. Conclusions

To the best of our knowledge, this study is the first report of the extremely small remobilisation of B[a]P residues after the removal of readily available fractions in long-term aged soils, including associated remobilisation kinetics and associated implications for contaminated land decision-making. After removal of readily available B[a]P fractions, only small amounts of B[a]P residues were remobilised through slow intercompartmental repartitioning of sequestered fractions into readily available fractions. However, amounts of B[a]P that were remobilised decreased significantly over time. At each aging time, remobilisation kinetics of B[a]P residues through the re-equilibration times investigated remained unchanged indicating that the small amounts of B[a]P residues which were remobilised remained stable with time. After extended periods of aging, the amounts of B[a]P remobilised in soils after prior removal of readily available or total-extractable fractions were generally extremely small, indicating no significant potential to pose risks to human and environmental health. Therefore, it is reasonable to consider readily available fractions of B[a]P in the risk assessment of long-term contaminated soils, rather than total-extractable fractions.

Conflict of Interests

The authors declare no competing financial interest.

Acknowledgment

This work was supported by the joint support of the University of Newcastle (UoN), Australia through the UoN International Postgraduate Research Scholarships, and the Cooperative Research Centre for Contamination Assessment and Remediation of the Environment through the CRC CARE PhD Scholarships. AU acknowledges the kind assistance of Oluyoye Idowu from the Global Centre for Environmental Remediation, UoN during laboratory experiments.

Appendix A. Supplementary data

Supplementary material to this article can be found online.

References

1. Barraclough, D.; Kearney, T.; Croxford, A. Bound residues: Environmental solution or future problem? *Environ. Pollut.* **2005**, *133*, (1), 85-90; DOI 10.1016/j.envpol.2004.04.016.
2. *Significance of Bound Residues in Environmental Risk Assessment: Workshop Report No. 17*; European Centre for Ecotoxicology and Toxicology of Chemicals: Brussels, Belgium, 2010; http://www.ecetoc.org/wp-content/uploads/2014/08/ECETOC_WR_17._Significance_of_bound_residues_in_environmental_risk_assessment.pdf. Accessed 11/06/2018.
3. *Development of Interim Guidance for the Inclusion of Non-Extractable Residues in the Risk Assessment of Chemicals*: Brussels, Belgium, 2013; <http://www.ecetoc.org/wp-content/uploads/2014/08/ECETOC-TR-118-Development-of-interim-guidance-for-the-inclusion-of-non-extractable-residues-NER-in-the-risk-assessment-of-chemicals.pdf>. Accessed 11/06/2018.
4. Umeh, A. C.; Duan, L.; Naidu, R.; Semple, K. T. Residual hydrophobic organic contaminants in soil: Are they a barrier to risk-based approaches for managing contaminated land? *Environ. Int.* **2017**, *98*, 18-34; DOI <https://doi.org/10.1016/j.envint.2016.09.025>.
5. *Agents Classified by the IARC Monographs*; In Working Group on the Evaluation of Carcinogenic Risks to Humans: International Agency for Research on Cancer: Lyon, France, 2012; <https://monographs.iarc.fr/ENG/Monographs/vol100F/mono100F-14.pdf>. Accessed 05/06/2018.
6. Lemieux, C. L.; Long, A. S.; Lambert, I. B.; Lundstedt, S.; Tysklind, M.; White, P. A. Cancer risk assessment of polycyclic aromatic hydrocarbon contaminated soils determined using bioassay-derived levels of Benzo[a]pyrene equivalents. *Environ. Sci. Technol.* **2015**, *49*, (3), 1797-1805; DOI 10.1021/es504466b.

7. Wilson, S. C.; Jones, K. C. Bioremediation of soil contaminated with polynuclear aromatic hydrocarbons (PAHs): A review. *Environ. Pollut.* **1993**, *81*, (3), 229-249; DOI [http://dx.doi.org/10.1016/0269-7491\(93\)90206-4](http://dx.doi.org/10.1016/0269-7491(93)90206-4).
8. Semple, K.; Morriss, A.; Paton, G. Bioavailability of hydrophobic organic contaminants in soils: Fundamental concepts and techniques for analysis. *Eur. J. Soil Sci.* **2003**, *54*, (4), 809-818; DOI <https://doi.org/10.1046/j.1351-0754.2003.0564.x>.
9. Rhodes, A. H.; McAllister, L. E.; Semple, K. T. Linking desorption kinetics to phenanthrene biodegradation in soil. *Environ. Pollut.* **2010**, *158*, (5), 1348-1353; DOI [10.1016/j.envpol.2010.01.008](https://doi.org/10.1016/j.envpol.2010.01.008).
10. Cornelissen, G.; Gustafsson, O.; Bucheli, T.; Jonker, M.; Koelmans, A.; Van Noort, P. Extensive sorption of organic compounds to black carbon, coal, and kerogen in sediments and soils: Mechanisms and consequences for distribution, bioaccumulation, and biodegradation. *Environ. Sci. Technol.* **2005**, *39*, (18), 6881-6895; DOI [10.1021/es050191b](https://doi.org/10.1021/es050191b).
11. Huang, W. L.; Ping, P. A.; Yu, Z. Q.; Fu, H. M. Effects of organic matter heterogeneity on sorption and desorption of organic contaminants by soils and sediments. *Appl. Geochem.* **2003**, *18*, (7), 955-972; DOI [10.1016/S0883-2927\(02\)00205-6](https://doi.org/10.1016/S0883-2927(02)00205-6).
12. Pignatello, J. J.; Xing, B. S. Mechanisms of slow sorption of organic chemicals to natural particles. *Environ. Sci. Technol.* **1996**, *30*, (1), 1-11; DOI [10.1021/es940683g](https://doi.org/10.1021/es940683g).
13. Luthy, R.; Aiken, G.; Brusseau, M.; Cunningham, S.; Gschwend, P.; Pignatello, J.; Reinhard, M.; Traina, S.; Weber, W.; Westall, J. Sequestration of hydrophobic organic contaminants by geosorbents. *Environ. Sci. Technol.* **1997**, *31*, (12), 3341-3347; DOI [10.1021/es970512m](https://doi.org/10.1021/es970512m).
14. Beckles, D. M.; Chen, W.; Hughes, J. B. Bioavailability of polycyclic aromatic hydrocarbons sequestered in sediment: Microbial study and model prediction. *Environ. Toxicol. Chem.* **2007**, *26*, (5), 878-883; DOI [Doi 10.1897/06-410r.1](https://doi.org/10.1897/06-410r.1).

15. Ortega-Calvo, J. J.; Tejeda-Agredano, M. C.; Jimenez-Sanchez, C.; Congiu, E.; Sungthong, R.; Niqui-Arroyo, J. L.; Cantos, M. Is it possible to increase bioavailability but not environmental risk of PAHs in bioremediation? *J. Hazard. Mater.* **2013**, *261*, 733-745; DOI 10.1016/j.jhazmat.2013.03.042.
16. Craven, A. Bound residues of organic compounds in the soil: The significance of pesticide persistence in soil and water: A European regulatory view. *Environ. Pollut.* **2000**, *108*, (1), 15-18; DOI 10.1016/S0269-7491(99)00198-0.
17. Craven, A.; Hoy, S. Pesticide persistence and bound residues in soil - regulatory significance. *Environ. Pollut.* **2005**, *133*, (1), 5-9; DOI 10.1016/j.envpol.2004.04.010.
18. Kastner, M.; Nowak, K. M.; Miltner, A.; Trapp, S.; Schaffer, A. Classification and modelling of nonextractable residue (NER) formation of xenobiotics in soil - A synthesis. *Crit. Rev. Env. Sci. Tec.* **2014**, *44*, (19), 2107-2171; DOI 10.1080/10643389.2013.828270.
19. Eschenbach, A.; Wienberg, R.; Mahro, B. Fate and stability of nonextractable residues of [(14)C]PAH in contaminated soils under environmental stress conditions. *Environ. Sci. Technol.* **1998**, *32*, (17), 2585-2590; DOI 10.1021/es9708272.
20. Northcott, G. L.; Jones, K. C. Partitioning, extractability, and formation of nonextractable PAH residues in soil. 1. Compound differences in aging and sequestration. *Environ. Sci. Technol.* **2001**, *35*, (6), 1103-1110; DOI 10.1021/es000071y.
21. Umeh, A. C.; Duan, L.; Naidu, R.; Semple, K. T. Time-dependent remobilisation of nonextractable benzo[a]pyrene residues in contrasting soils: Effects of aging, spiked concentration, and soil properties. *Environ. Sci. Technol.* **2018**, *52*, (21), 12295-12305; DOI 10.1021/acs.est.8b03008.
22. Ortega-Calvo, J.-J.; Harmsen, J.; Parsons, J. R.; Semple, K. T.; Aitken, M. D.; Ajao, C.; Eadsforth, C.; Galay-Burgos, M.; Naidu, R.; Oliver, R.; Peijnenburg, W. J. G. M.; Römbke,

- J.; Streck, G.; Versonnen, B. From bioavailability science to regulation of organic chemicals. *Environ. Sci. Technol.* **2015**, *49*, (17), 10255-10264; DOI 10.1021/acs.est.5b02412.
23. Doick, K. J.; Klingelmann, E.; Burauel, P.; Jones, K. C.; Semple, K. T. Long-term fate of polychlorinated biphenyls and polycyclic aromatic hydrocarbons in an agricultural soil. *Environ. Sci. Technol.* **2005**, *39*, (10), 3663-3670; DOI 10.1021/es048181i.
24. Gao, Y.; Hu, X.; Zhou, Z.; Zhang, W.; Wang, Y.; Sun, B. Phytoavailability and mechanism of bound PAH residues in field contaminated soils. *Environ. Pollut.* **2017**, *222*, 465-476; DOI <https://doi.org/10.1016/j.envpol.2016.11.076>.
25. Semple, K. T.; Doick, K. J.; Jones, K. C.; Burauel, P.; Craven, A.; Harms, H., Peer reviewed: Defining bioavailability and bioaccessibility of contaminated soil and sediment is complicated. *Environ. Sci. Technol.* **2004**, *38*, (12), 228A-231A; DOI 10.1021/es040548w.
26. Naidu, R.; Wong, M. H.; Nathanail, P. Bioavailability-the underlying basis for risk-based land management. *Environ. Sci. Pollut. R.* **2015**, *22*, (12), 8775-8778; DOI 10.1007/s11356-015-4295-z.
27. Umeh, A. C.; Duan, L.; Naidu, R.; Semple, K. T. Comparison of single- and sequential-solvent extractions of total extractable benzo[a]pyrene fractions in contrasting soils. *Anal. Chem.* **2018**, *90*, 19, 11703-11709; DOI 10.1021/acs.analchem.8b03387.
28. Field, A. P. *Discovering Statistics Using IBM SPSS Statistics: And Sex And Drugs And Rock 'N' Roll*. Sage: Los Angeles, U.S.A., 2013.
29. Duan, L.; Palanisami, T.; Liu, Y. J.; Dong, Z. M.; Mallavarapu, M.; Kuchel, T.; Semple, K. T.; Naidu, R. Effects of ageing and soil properties on the oral bioavailability of benzo[a]pyrene using a swine model. *Environ. Int.* **2014**, *70*, 192-202; DOI 10.1016/j.envint.2014.05.017.
30. Alexander, M. Aging, bioavailability, and overestimation of risk from environmental pollutants. *Environ. Sci. Technol.* **2000**, *34*, (20), 4259-4265; DOI 10.1021/es001069+.

31. Hatzinger, P.; Alexander, M. Effect of ageing of chemicals in soil on their biodegradability and extractability. *Environ. Sci. Technol.* **1995**, *29*, (2), 537-545; DOI 10.1021/es00002a033.
32. Luo, L.; Lin, S.; Huang, H. L.; Zhang, S. Z. Relationships between aging of PAHs and soil properties. *Environ. Pollut.* **2012**, *170*, 177-182; DOI 10.1016/j.envpol.2012.07.003.
33. Liste, H.; Alexander, M. Butanol extraction to predict bioavailability of PAHs in soil. *Chemosphere* **2002**, *46*, (7), 1011-1017; DOI 10.1016/S0045-6535(99)00216-7.
34. Gomez-Eyles, J. L.; Collins, C. D.; Hodson, M. E. Relative proportions of polycyclic aromatic hydrocarbons differ between accumulation bioassays and chemical methods to predict bioavailability. *Environ. Pollut.* **2010**, *158*, (1), 278-284; DOI 10.1016/j.envpol.2009.07.012.
35. *PAH Interactions with Soil and Effects on Bioaccessibility and Bioavailability to Humans*; Final report Department of Defense Strategic Environmental Research and Development Program (SERDP) project ER-1743: Exponent, Inc., Boulder, U.S.A., 2017; [https://www.serdp-estcp.org/Program-Areas/Environmental-Restoration/Risk Assessment/ER-1743](https://www.serdp-estcp.org/Program-Areas/Environmental-Restoration/Risk-Assessment/ER-1743). Accessed 11/06/2018.
36. Duan, L.; Naidu, R.; Liu, Y. J.; Palanisami, T.; Dong, Z. M.; Mallavarapu, M.; Semple, K. T. Effect of ageing on benzo[a]pyrene extractability in contrasting soils. *J. Hazard. Mater.* **2015**, *296*, 175-184; DOI 10.1016/j.jhazmat.2015.04.050.
37. Macleod, C.; Semple, K. Influence of contact time on extractability and degradation of pyrene in soils. *Environ. Sci. Technol.* **2000**, *34*, (23), 4952-4957; DOI 10.1021/es000061x.
38. Rhodes, A.; Carlin, A.; Semple, K. Impact of black carbon in the extraction and mineralization of phenanthrene in soil. *Environ. Sci. Technol.* **2008**, *42*, (3), 740-745; DOI 10.1021/es071451n.

39. Rhodes, A.; Riding, M.; McAllister, L.; Lee, K.; Semple, K. Influence of Activated Charcoal on Desorption Kinetics and Biodegradation of Phenanthrene in Soil. *Environ. Sci. Technol.* **2012**, *46*, (22), 12445-12451; DOI 10.1021/es3025098.
40. Doick, K. J.; Burauel, P.; Jones, K. C.; Semple, K. T. Distribution of aged C-14-PCB and C-14-PAH residues in particle-size and humic fractions of an agricultural soil. *Environ. Sci. Technol.* **2005**, *39*, (17), 6575-6583; DOI 10.1021/es050523c.
41. Semple, K. T.; Riding, M. J.; McAllister, L. E.; Sopena-Vazquez, F.; Bending, G. D. Impact of black carbon on the bioaccessibility of organic contaminants in soil. *J. Hazard. Mater.* **2013**, *261*, (0), 808-816; DOI <http://dx.doi.org/10.1016/j.jhazmat.2013.03.032>.
42. Vogel, C.; Heister, K.; Buegger, F.; Tanuwidjaja, I.; Haug, S.; Schloter, M.; Kogel-Knabner, I. Clay mineral composition modifies decomposition and sequestration of organic carbon and nitrogen in fine soil fractions. *Biol. Fert. Soils* **2015**, *51*, (4), 427-442; DOI 10.1007/s00374-014-0987-7.
43. Macleod, C. J. A.; Semple, K. T. Sequential extraction of low concentrations of pyrene and formation of non-extractable residues in sterile and non-sterile soils. *Soil Biol. Biochem.* **2003**, *35*, (11), 1443-1450; 10.1016/S0038-0717(03)00238-4.
44. Cui, X.; Mayer, P.; Gan, J. Methods to assess bioavailability of hydrophobic organic contaminants: Principles, operations, and limitations. *Environ. Pollut.* **2013**, *172*, 223-234; DOI 10.1016/j.envpol.2012.09.013.
45. Swindell, A. L.; Reid, B. J. Comparison of selected non-exhaustive extraction techniques to assess PAH availability in dissimilar soils. *Chemosphere* **2006**, *62*, (7), 1126-1134; DOI 10.1016/j.chemosphere.2005.05.047.
46. Hwang, S. C.; Cutright, T. J. Evidence of underestimation in PAH sorption/desorption due to system nonequilibrium and interaction with soil constituents. *J. Environ. Sci. Heal. A* **2004**, *39*, (5), 1147-1162; DOI 10.1081/Ese-120030300.

47. Hwang, S. C.; Cutright, T. J. Preliminary exploration of the relationships between soil characteristics and PAH desorption and biodegradation. *Environ. Int.* **2004**, *29*, (7), 887-894; DOI 10.1016/S0160-4120(03)00053-9.
48. Riding, M. J.; Doick, K. J.; Martin, F. L.; Jones, K. C.; Semple, K. T. Chemical measures of bioavailability/bioaccessibility of PAHs in soil: Fundamentals to application. *J. Hazard. Mater.* **2013**, *261*, (0), 687-700; DOI <http://dx.doi.org/10.1016/j.jhazmat.2013.03.033>.
49. Latawiec, A.; Swindell, A.; Simmons, P.; Reid, B. Bringing bioavailability into contaminated land decision making: The way forward? *Crit. Rev. Env. Sci. Tec.* **2011**, *41*, (1), 52-77; DOI 10.1080/00102200802641780.
50. Kelsey, J.; Kottler, B.; Alexander, M. Selective chemical extractants to predict bioavailability of soil-aged organic chemicals. *Environ. Sci. Technol.* **1997**, *31*, (1), 214-217; DOI 10.1021/es960354j.
51. Jonker, M. T. O.; Koelmans, A. A. Extraction of polycyclic aromatic hydrocarbons from soot and sediment: Solvent evaluation and implications for sorption mechanism. *Environ. Sci. Technol.* **2002**, *36*, (19), 4107-4113; DOI 10.1021/es0505191.
52. Birdwell, J. E.; Thibodeaux, L. J. PAH repartitioning in field-contaminated sediment following removal of the labile chemical fraction. *Environ. Sci. Technol.* **2009**, *43*, (21), 8092-8097; DOI 10.1021/es9016798.
53. Naidu, R.; et al. Bioavailability: the underlying basis for risk based land management. In *Chemical Bioavailability in Terrestrial Environments*, Naidu, R., Ed.; Elsevier: Amsterdam 2008; pp 53 - 72.
54. Nathanail, C. P.; Ogden, R. C. Derivation of a site-specific assessment criterion for benzo[a]pyrene in red shale at a former coking works. *J. Environ. Sci. Heal. A* **2013**, *48*, (6), 594-603; DOI 10.1080/10934529.2013.731353.

55. Latawiec, A. E.; Simmons, P.; Reid, B. J. Decision-makers' perspectives on the use of bioaccessibility for risk-based regulation of contaminated land. *Environ. Int.* **2010**, *36*, (4), 383-389; DOI <http://dx.doi.org/10.1016/j.envint.2010.02.007>.

56. *The derivation of HILs for PAHs and total phenols*: National environment protection (assessment of site contamination) amendment measure; National Environment Protection Council (NEPC): Canberra, Australia, 2013; https://www.legislation.gov.au/Details/F2013C00288/Html/Volume_11. Accessed 15/06/2018.