

1 Wheat's wild relatives vary in their response to nitrogen and ozone

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5

6 **Abstract**

7 The wild relatives of bread wheat (*Triticum aestivum* L.) are valued by plant breeders for
8 their genetic diversity. However, increasing levels of nitrogen (N) deposition and ground-
9 level ozone (O₃) threaten plant biodiversity in the Mediterranean and Near East, a hotspot
10 for many crop wild relatives. Knowledge of the effect of these air pollutants in combination is
11 still limited, but early indications are that effects vary depending on the level of pollutants,
12 and on the sensitivity of the species to N and O₃. This study examined the responses of four
13 important wheat wild relatives (*Aegilops tauschii*, *Aegilops speltoides*, *Triticum dicoccoides*,
14 and *Triticum monococcum*) and one modern wheat cultivar (*Triticum aestivum* 'Cadenza') to
15 treatments of N (equivalent to 50 kg ha⁻¹ yr⁻¹ ammonium nitrate) and O₃ (100 ppb for 21
16 days), alone and in combination. Measurements included root, shoot and seed biomass, and
17 electrolyte ratios. The O₃ sensitivity of *Ae. tauschii* and *T. 'Cadenza'* were exacerbated by
18 the addition of N, whilst *Ae. speltoides* was found to be nitrophilous, with N ameliorating the
19 negative effect of O₃. Both *T. 'Cadenza'* and *T. dicoccoides* produced immature seed heads,
20 with the cultivar's seed head biomass reduced in response to O₃ and N+O₃ whilst that of *T.*
21 *dicoccoides* was largely unaffected. These data suggest that all four wild relatives are likely
22 to be affected when N and O₃ air pollutants co-occur, and their *in situ* populations may
23 therefore be at risk. Equally, the results of this study can inform use of their beneficial traits
24 by wheat breeders, and alert them to the inadvertent inclusion of N and O₃ sensitivity.

25

26 **Keywords**

27 Crop wild relative; *Triticum aestivum* L.; *Triticum dicoccoides*; *Aegilops tauschii*; *Aegilops*
28 *speltoides*; air pollution.

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29 Introduction

30 Wheat provides 20% of the global calorie and protein intake consumed by the current
31 population of 7.5 billion people, with the majority supplied by just one species, *Triticum*
32 *aestivum* L., bread wheat (Shiferaw et al., 2013). Whilst the green revolution delivered higher
33 yields it also led to a substantial reduction in the genetic diversity found in wheat cultivars,
34 making them more vulnerable to pest and disease attack and less adaptable to abiotic stress
35 (The Royal Society, 2009). Wild populations of *Triticum* and *Aegilops* species which form the
36 close wild relatives of wheat, have retained their wide genetic diversity and, as a result, are
37 particularly useful to plant breeders tasked with both increasing wheat yields and broadening
38 the narrow genetic profiles of wheat cultivars to improve resilience (Brozynska et al., 2015,
39 Dempewolf et al., 2017). These species continue to grow in the eastern Mediterranean and
40 Near East, a region containing a high proportion of important crop wild relatives (Castañeda-
41 Álvarez et al., 2016). However, this region is also a convergence zone for global air
42 pollutants, especially reactive nitrogen (N) and ozone (O₃) (Lelieveld et al., 2002), both of
43 which are known to affect plant biodiversity (Stevens et al., 2004).

44 Many of wheat's close wild relatives have been extensively used for wheat improvement,
45 mostly to improve pest and disease resistance (Maxted and Kell, 2009). Whilst the *ex situ*
46 collections of wheat germplasm conserved in gene-banks around the world are considered
47 to be well stocked (Castañeda-Álvarez et al., 2016), the importance of maintaining the wild
48 populations *in situ* is now recognised as a high priority, both because the genetic diversity of
49 the *ex situ* accessions is comparatively limited (Sasanuma et al., 2002) but also because,
50 unlike the stored seed, wild populations continue to evolve and adapt to global change
51 (Maxted et al., 2010, Castañeda-Álvarez et al., 2016). However, crop wild relative
52 populations in the Mediterranean are under threat, with agricultural intensification, over-
53 grazing, habitat loss, and climate change all contributing to a loss of genetic diversity (Jarvis
54 et al., 2008, Stolton et al., 2008, Haruntyunyan et al., 2010, Redden et al., 2015). The
55 additional threat from air pollution may be equally important, and affect species even in
56 protected areas.

57 Over the past 100 years the burning of fossil fuels and increasing use of artificial fertilisers
58 for agriculture have led to a doubling in the total fixation of N globally and a tripling of levels
59 within Europe (Sutton and van Grinsven, 2011). With ongoing industrialisation and
60 agricultural intensification in many parts of the world, levels of reactive N are expected to
61 continue to increase (Sutton and Bleeker, 2013), with predictions that N deposition across
62 the Mediterranean region, currently in the range of 12 – 45 kg N ha⁻¹ yr⁻¹, could triple by
63 2050 (Galloway et al., 2004, Ochoa-Hueso and Stevens, 2015). Anthropogenic emission
64 sources also contribute precursor gases, nitrogen oxides, methane, carbon monoxide and
65 non-methane volatile organic compounds, which react with sunlight to form O₃. Global O₃
66 daytime growing-season averages reach 30-50 ppb in the northern hemisphere (Sutton and
67 van Grinsven, 2011) with occasional short term peak O₃ episodes of 100-200 ppb (The
68 Royal Society, 2008). Some of the highest background levels of O₃ occur in the
69 Mediterranean and Middle East, especially in summer when air streams from Europe, Asia
70 and Africa converge on the region (Zanis et al., 2014). O₃ levels of 60-100 ppb are expected
71 to occur in parts of the eastern Mediterranean and Middle East by 2050 (Lelieveld et al.,
72 2014).

73 O₃ is toxic to plants, some being more sensitive than others (Ashmore, 2005, Mills et al.,
74 2016). Wheat is one of the most O₃ sensitive crops, with losses of global wheat yield
75 estimated at 9.9% in the northern hemisphere [MILLS ET AL 2018] (Van Dingenen et al.,
76 2009) and it is suggested that until global O₃ pollution can be controlled the only adaptation

77 available is the development of O₃ tolerant cultivars (Teixeira et al., 2011). Previous research
78 assessed the O₃ sensitivity of several of the genome donors of modern bread wheat, with
79 plants grown for 21 days and exposed to 100 ppb O₃ in open top chambers (Biswas et al.,
80 2008a). *Ae. tauschii* and *T. monococcum* were found to be most sensitive to O₃, with
81 reductions in shoot and root biomass, and rates of photosynthesis. The DD genome of *Ae.*
82 *tauschii* was proposed as the primary source of wheat's O₃ sensitivity (Biswas et al., 2008a).
83 Wild plants are known to be as sensitive to O₃ as crops (The Royal Society, 2008) and a
84 recent study of the effect of O₃ on Natura 2000 grassland habitats in Europe revealed that
85 areas in the Mediterranean are most at risk as a result of high levels of O₃ flux in the region
86 (Harmens and Mills, 2016). Separate studies have confirmed changes in species
87 composition due to a loss of O₃ sensitive species (Calvete-Sogo et al., 2016).

88 In the base rich soils of the Mediterranean, N deposition tends to lead to eutrophication,
89 supporting the growth of nitrophilous species (Bobbink et al., 2010) with a subsequent
90 impact on community composition. In Mediterranean grasslands, species richness has been
91 found to decline in response to N deposition, with perennial grasses replacing forb species,
92 especially those adapted to low N environments such as legumes (Bonanomi et al., 2013). In
93 the Mediterranean and Near East, N deposition is already exceeding critical loads for some
94 species (Phoenix et al., 2006), and climate change is predicted to increase other abiotic
95 stress factors through increased temperatures and water shortages (Bonanomi et al., 2013,
96 Intergovernmental Panel on Climate Change, 2013).

97 Despite both N deposition and O₃ pollution frequently occurring together, relatively little
98 research has been performed on the effect on plants of the two pollutants in combination
99 (Ochoa-Hueso et al., 2011, Sánchez-Martín et al., 2017). The research that has been done
100 to date suggests that they do not necessarily replicate the effect of the individual pollutants
101 and the outcome of interactions can differ depending on a variety of factors, including the
102 plant's sensitivity to each pollutant (Mills et al., 2016). This study examined the responses of
103 four of wheat's close wild relatives and one wheat cultivar at an early stage of development
104 to N fertilisation and O₃ exposure, both separately and in combination. The study aimed to
105 test the following hypotheses: firstly, that the growth of each species would be positively
106 affected by N and negatively affected by O₃; and secondly, that there would be an interaction
107 between N and O₃ resulting in N either ameliorating or exacerbating the effect of O₃
108 depending on the species.

109 **Materials and methods**

110

111 *Plant establishment*

112 Two primary and two secondary wild relatives of wheat, along with one wheat cultivar, were
113 selected for the trial (Table 1). *Ae. tauschii*, *T. dicoccoides* and a close relative of *Ae.*
114 *speltoides* are the likely genome donors of bread wheat (Marcussen et al., 2014). *T.*
115 *monococcum* is one of the earliest forms of cultivated wheat (Matsuoka, 2011). All are
116 annual species. De-husked seeds were germinated in an incubator set at 23.5 °C and sown
117 into 1l plastic pots containing 650 g of Levington's John Innes No.1, a low nutrient seedling
118 compost, with two plants per pot. Treatments were replicated four times. All plants were
119 grown on in a temperature-controlled glasshouse (20.5 ± 2°C, relative humidity 51 ± 13 %)
120 under natural daylight for 23 days (25 April – 18 May 2016), with pots rotated randomly
121 every 3 - 4 days. Throughout the trial, all pots were manually watered, daily.

122 *Nitrogen and ozone treatment*

123 Prior to the start of O₃ exposure, 50 kg ha⁻¹ yr⁻¹ (25 ml) of ammonium nitrate solution was
124 added to N treatment pots in two split doses to reduce leaching potential, on the 21st and
125 23rd days after planting (DAP). An equivalent amount of water was added to the remaining
126 pots. At 24 DAP all pots were moved to closed top-lit chambers measuring 0.7 m x 0.7 m
127 (24-h photoperiod, 200 ± 14 μmol m⁻² s⁻¹, 24 ± 1.0 °C, 35.0 ± 6.4% humidity) in a
128 temperature controlled glasshouse. Pots were divided by treatment, with 20 pots placed in
129 each chamber. All pots were rotated and randomised within the chamber every two days and
130 between chambers every four days, with two out of the three available air or O₃ chambers
131 occupied at any one time. All chambers were supplied with charcoal filtered air from the
132 glasshouse, with half the chambers ventilated continuously with O₃ using a *Triogen* lab O₃
133 generator throughout the treatment period of 21 days (19 May – 9 June 2016). The mean O₃
134 level was 32 ± 4 ppb in the air chambers and 113 ± 22 ppb across all three O₃-ventilated
135 chambers, with a range of means from 104 ± 20 ppb to 126 ± 25 ppb, monitored using an
136 *Aeroqual v500* portable indoor air quality monitor. The length of trial and experimental
137 conditions were similar to previous studies assessing the effect of O₃ on wheat cultivars and
138 wheat's wild relatives (Biswas et al., 2008a).

139 *Post-harvest measurements*

140 Measurements of electrolyte leakage were obtained using a 2 cm tissue sample taken from
141 one of the two plants in each pot at the point of harvest. Samples were placed in 1 ml of RO
142 water, incubated at room temperature and shaken for 24 h. Conductivity was measured,
143 after 24 h of shaking, and then again following freezing and autoclaving using a *Horiba B-*
144 *173* conductivity meter.

145 The plant not used for electrolyte tissue tests was harvested from each replicate pot with the
146 shoot cut at the surface of the substrate. Measurements of shoot height, fresh shoot
147 biomass, and leaf area (Transparent Belt Conveyor Li-3050A, *Li-Cor*, Lincoln, NE, USA)
148 were taken immediately. Shoot biomass was determined by drying at 60°C for 4-6 days until
149 a constant dry weight was reached. Root samples were retained *in situ* in pots and left to dry
150 for at least 3 days before roots were washed by wet sieving and then dried in drying cabinets
151 at 60°C for 4-6 days until a constant dry weight was reached. As the roots of the two plants
152 in each pot were intertwined the total root dry biomass was halved to derive the root dry
153 biomass data for each replicate and for the root:shoot ratio calculations. Two of the *Triticum*
154 species, the cultivar *T. 'Cadenza'* and *T. dicoccoides*, had formed immature seed heads by

155 the end of the trial period, though before any grain fill had occurred. These seedheads were
156 cut at the base and dry seed head biomass measurements taken.

157 Shoot C:N ratios were obtained by grinding in a mixer mill (*Retsch* MM400, Hann, Germany),
158 and analysed using a *Vario* EL III CHNOS Elemental Analyzer (Elementar Analysensysteme
159 GmbH, Germany).

160 *Statistical analysis*

161 The effects of the different treatments on the parameters measured were assessed using
162 Analysis of Variance (ANOVA). All data were checked for normality and log transformed
163 where necessary. 3-way ANOVAs of the effects of N and O₃ and their interaction across all
164 species were carried out for each variable. 2-way ANOVAs were carried out on the data for
165 individual species and treatments. All statistical analyses were carried out using IBM SPSS
166 statistics v22. Significance was assessed at $p < 0.05$.

167 **Results**

168 *Effects of nitrogen and ozone on growth parameters*

169 The height and leaf area of the five species (Fig. 1) were all significantly different to each
170 other ($p < 0.001$) under control conditions. There were no significant effects of the N or O₃
171 treatments on the height or leaf area of either *T. monococcum* or *Ae. tauschii*. N had a
172 significant positive effect on the height of *Ae. speltooides* ($p = 0.011$) and a strong trend
173 towards a positive interaction between N and O₃ ($p = 0.05$) whereby the O₃-only treatment
174 reduced the height of *Ae. speltooides* whilst the N+O₃ treatment increased it. The leaf area of
175 *T. 'Cadenza'* increased significantly in response to O₃ ($p = 0.026$), whilst there was a trend
176 towards a positive effect of N on the leaf area of *T. dicoccoides* ($p = 0.067$).

177 There were significant differences between species' shoot and root biomass and subsequent
178 root:shoot ratios ($p < 0.001$) under control conditions (Fig. 2). There were no significant
179 effects of the treatments on shoot biomass. There were trends toward a negative effect of O₃
180 on the shoot biomass of *Ae. tauschii* ($p = 0.057$), and an interaction between N and O₃
181 ($p = 0.067$) reducing the shoot biomass of *Ae. tauschii* further under the N+O₃ treatment.
182 There was also a small but non-significant positive response to N on the shoot biomass of
183 *Ae. speltooides* ($p = 0.073$). None of the treatments had a significant effect on the root biomass
184 of any species apart from *T. 'Cadenza'* where O₃ had a significant negative effect ($p = 0.037$).

185 There was a significant difference in the seed head biomass (Fig. 3) between the two
186 species which produced seed heads, *T. 'Cadenza'* and *T. dicoccoides* ($p < 0.001$). *T.*
187 *Cadenza's* seed head biomass was significantly reduced by O₃ ($p = 0.007$). There were no
188 significant effects of any of the treatments on the seed head biomass of *T. dicoccoides*.

189 There were significant differences in the C:N ratios between species under control conditions
190 (Fig. 4). There were no significant effects of N or O₃ alone, although the C:N ratios of *T.*
191 *'Cadenza'* ($p = 0.054$) and *T. monococcum* ($p = 0.061$) showed small but non-significant
192 reductions with the addition of N. However, across the majority of species the combined
193 N+O₃ treatment significantly reduced the C:N ratio ($p = 0.004$).

194 *Effects of nitrogen and ozone on ozone injury*

195 High variability in the electrolyte ratio data for most species meant there were no significant
196 differences between species (Fig. 4) and there were only non-significant differences in the
197 response to treatments. A significant statistical correlation was found between the
198 electrolyte ratio and root dry biomass ($r^2 = -.322$, 2-tailed significance $p = 0.004$), with an
199 increase in electrolyte leakage correlated with a reduction in root biomass.

200 **Discussion**

201 *Ozone sensitivity and nitrogen responsiveness*

202 Although *T. monococcum* showed some O₃ sensitivity, as has been found previously
203 (Biswas et al., 2008a), none of the treatments had any significant effects on this species. *Ae.*
204 *tauschii* and *Ae. speltooides* showed O₃ sensitivity but each responded differently to N and to
205 N+O₃. *Ae. tauschii* had reduced shoot and root biomass in O₃-only and N+O₃ treatments
206 (Fig.2), replicating similar responses to N+O₃ found in Mediterranean annual pastures
207 (Calvete-Sogo et al., 2016). These results support those from a previous study on the O₃
208 sensitivity of *Ae. tauschii* where it was found to have reduced shoot and root biomass, and,
209 as the donor of the DD genome, was proposed as the likely source of bread wheat's O₃
210 sensitivity (Biswas et al., 2008a). In addition, this new trial has shown that whilst *Ae. tauschii*
211 was largely unresponsive to the sole addition of N, N exacerbated the negative effect of O₃,
212 with further reductions in the shoot dry biomass of this species in response to the N+O₃
213 treatment; a response pattern also replicated in the wheat cultivar (Fig. 2).

214 The O₃ sensitivity of *Ae. speltooides* had not previously been assessed prior to this study. It
215 was found to be both less affected by O₃ and more responsive to N than *Ae. tauschii*, with
216 an ameliorating effect of N under the N+O₃ treatment on its height, leaf area, shoot biomass
217 and electrolyte ratio (Figs. 1, 2, & 4). These results suggest that if O₃ occurs with N
218 deposition, *Ae. speltooides* would be much less affected by O₃ than *Ae. tauschii*. In
219 Mediterranean annual pastures similar effects have been found, with N moderating the effect
220 of O₃ on biomass and yield, though not under high O₃ exposure (Calvete-Sogo et al., 2014).

221 Diploid *Aegilops* species, like *Ae. speltooides* and *Ae. tauschii*, are known to have a higher
222 frequency of stomata compared to triploid species (Kharazian, 2007), potentially leading to
223 higher levels of O₃ flux, especially with the large leaf area characteristic of *Ae. tauschii*.
224 Whilst rapid growth caused by N deposition confers an advantage on nitrophilous species,
225 rapid growth and high stomatal frequency is likely to be a disadvantage for plants also
226 subject to O₃ pollution, unless this is accompanied by rapid detoxification of O₃ (Ashmore,
227 2005). These factors may make fast-growing annual species which are not nitrophilous, such
228 as *Ae. tauschii*, potentially more vulnerable to O₃, especially when this rapid growth takes
229 place in spring when high O₃ levels can occur (Thwaites et al., 2006, Calvete-Sogo et al.,
230 2014, Calvete-Sogo et al., 2016).

231 N deposition is also known to have a variable effect on different species depending on their
232 sensitivity to the oxidised or reduced forms of N (Stevens et al., 2011, van den Berg et al.,
233 2016). In a study assessing the effect of ambient O₃ on the assimilation of both reduced and
234 oxidised forms of N in immature wheat plants, O₃ increased the incorporation of reduced N,
235 in the form of ammonium, whilst having no effect on the incorporation of oxidised N, in the
236 form of nitrate (Möcker et al., 1996). This suggests that, where N and O₃ co-occur, the
237 increase in reduced N levels, which results from O₃ exposure, could either be beneficial, or
238 detrimental, depending on the sensitivity of the species to reduced forms of N.

239 Other studies have shown that O₃ also affects the structure and functioning of microbial
240 communities in the rhizosphere (Zak et al., 2007, Li et al., 2013, Wu et al., 2016, Sánchez-
241 Martín et al., 2017). Following long term exposure to 60 ppb O₃, O₃ sensitive wheat cultivars
242 had significantly reduced microbial communities, including mycorrhizal fungi, compared to
243 O₃ tolerant cultivars. This led to significant reductions in both reduced and oxidised N cycling
244 soil processes, resulting in higher ammonia concentrations in the rhizosphere (Wu et al.,
245 2016). Microbial communities associated with O₃ tolerant cultivars also showed a preference

246 for easily degradable carbon sources whilst those associated with O₃ sensitive cultivars
247 preferred more complex ones (Bao et al., 2015). A separate study on the different microbial
248 communities associated with wheat and its wild relatives found that *Ae. tauschii* generates
249 high levels of mycorrhizal fungi in its rhizosphere, and its DD genome appears to control the
250 abundance of mycorrhizal fungi in both bread wheat, *T. aestivum*, and also synthetic wheat
251 (Turner et al., 2013, Poole, 2016). Further study is required to assess the genetic links
252 between wheat's wild relatives, its O₃ sensitivity and its mycorrhizal fungal communities, as
253 well as to determine whether wheat's wild relatives have a preference for reduced or
254 oxidised N.

255 *Seed head production*

256 Both the cultivar, *T. 'Cadenza'*, and the wild relative, *T. dicoccoides*, produced immature
257 seed heads within the 6-week time period of the trial, with this rapid generation of seed head
258 biomass made possible by the use of a continuous photoperiod, a system approved by some
259 wheat breeding programmes in order to speed up the research and breeding process
260 (Hickey et al., 2009, Watson et al., 2018). Although further studies would be needed to
261 assess responses under a natural lighting regime, both species were therefore exposed to
262 O₃ during the time they flowered and developed seed heads, the growth stages when wheat
263 is known to be at its most sensitive to O₃ (Soja et al., 2000, Ashmore, 2005). *T. 'Cadenza'*
264 produced less seed head biomass compared to *T. dicoccoides*, and this was also
265 significantly reduced by O₃ (-39%) and further reduced by N+O₃ (-52%), with both shoot and
266 root biomass, and root:shoot ratio also reduced (Fig. 2). In comparison the seed head
267 biomass of *T. dicoccoides* was not significantly affected by either O₃ (+11%) or N+O₃
268 (+0.5%) (Fig. 3). Of the five species, *T. dicoccoides* had the smallest leaf area (Fig. 1) and
269 its root biomass was the least affected by treatments. It has also been found to have lower
270 levels of mycorrhizal fungi compared to *Ae. tauschii* and *T. aestivum* (Poole, 2016). All these
271 characteristics may contribute towards its O₃ tolerance, and may be of interest to plant
272 breeders, especially as the O₃ sensitivity of wheat cultivars has been found to have
273 increased over time. Experimental studies on both immature (Barnes et al., 1990,
274 Velissariou et al., 1992, Biswas et al., 2008b) and mature wheat plants (Pleijel et al., 2006,
275 Harmens et al., 2017) have shown that highly bred genotypes from the late 20th century and
276 early 21st century are more O₃ sensitive compared to older ones. However, further study
277 would be required to assess the specific cause of the O₃ tolerance of *T. dicoccoides* found in
278 this trial.

279 None of the other three wild species developed seed heads within the time period available
280 for this trial, and longer trials growing all species to seed ripened stage would be needed for
281 a full assessment of their responses to O₃, especially after anthesis (Soja et al., 2000).
282 However, any negative effects of these pollutants on their growth may also affect their ability
283 to produce seed. O₃ pollution is known to affect both quantity and viability of seed in some
284 species (Leisner and Ainsworth, 2012), and can also lead to changes in the timing of
285 phenology (Sanz et al., 2011). In annual Mediterranean pastures many species store
286 photosynthates early in the season for use in the drought-prone summer periods when
287 flowering and seed production occur. If the O₃ exposure occurs at this early stage of plant
288 growth then later seed production could be affected (Sanz et al., 2011).

289 *Implications for the conservation of wheat's wild relatives*

290 Due to the differing sensitivities of plant species, N deposition and O₃ lead to changes in the
291 composition of communities in Mediterranean-type ecosystems (Ochoa-Hueso et al., 2011,
292 Sanz et al., 2014, Calvete-Sogo et al., 2016). The results of this study suggest that the *in*

293 *situ* populations of all four wild species may be affected by either N or O₃, with varying
294 effects on each species when these pollutants co-occur. Further field study is required on
295 the plant communities which host wheat's wild relatives to assess how N deposition and O₃
296 pollution are affecting these species and their competitors. In addition, as legume species in
297 particular can be negatively affected by both N and O₃ (Sanz et al., 2014, Calvete-Sogo et
298 al., 2016) an assessment of the impact of N and O₃ on the crop wild relatives of important
299 leguminous crops such as pea, lentil and clover, also found in this Mediterranean and Near
300 East region would be beneficial.

301 *Implications for wheat breeding*

302 Whilst all the species in this study have adequate numbers of accessions within international
303 gene-banks (Castañeda-Álvarez et al., 2016), and as a result have been given low priority
304 status for *ex situ* collecting purposes (Global Crop Diversity Trust, 2016), their *in situ*
305 populations and genetic diversity remain important resources for plant breeding, both now
306 and in the future. As in previous studies (Biswas et al., 2008a), *Ae. tauschii* was shown to be
307 particularly O₃ sensitive, raising concerns that this trait could be transferred to some wheat
308 lines through the plant breeding process, with *Ae. tauschii* having been used, for example, to
309 improve disease resistance and drought tolerance (Global Crop Diversity Trust, 2016). In
310 addition, the creation of synthetic hexaploid wheat by combining *T. turgidum* L. with *Ae.*
311 *tauschii* is said to be a breakthrough in wheat breeding with the potential, as a result of its
312 improved genetic diversity, to increase both yields and resistance to biotic and abiotic
313 stresses (Li et al., 2014, Cox et al., 2017). It will be important, however, to ascertain whether
314 synthetic wheat carries *Ae. tauschii*'s O₃ sensitivity.

315 **Conclusions**

316 The *in situ* populations of crop wild relatives are of considerable economic value because
317 they have the potential to supply the genetic diversity and resilience needed in the most
318 important crops, including wheat. The potential impact of N deposition and O₃ on the
319 conservation of wheat's wild relatives is therefore of concern, and an understanding of their
320 N and O₃ sensitive traits is also important if they are to be used for breeding new wheat
321 lines. *Ae. tauschii*, previously identified as the likely source of wheat's O₃-sensitivity, along
322 with the cultivar *T. 'Cadenza'*, were found to be O₃-sensitive, and their O₃ sensitivity was
323 increased by the addition of N. *Ae. speltoides* proved to be nitrophilous, with N ameliorating
324 the negative impact of O₃. Whilst the cultivar had significantly reduced seed head biomass in
325 response to both O₃ and N+O₃ treatments, *T. dicoccoides* demonstrated O₃ tolerance in its
326 root, shoot and seed head biomass.

327 The *in situ* populations of all four wild relatives in this study are likely to be affected by N and
328 O₃, and, as N deposition and O₃ pollution affect species differently, the composition of the
329 plant communities in which these crop wild relatives reside are also likely to be affected.
330 Field studies would be needed to assess the extent of these changes and the implications
331 there may be for other crop wild relatives in this region, especially legumes which are known
332 to be particularly sensitive to both N and O₃. The results of this study can also inform the use
333 of these wild relatives in future plant breeding, not only to enable use of their beneficial traits,
334 but also to alert breeders to the inadvertent inclusion of N and O₃ sensitivity into wheat
335 cultivars.

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