A predictive model of wheat grain yield based on canopy reflectance indices and the theoretical definition of yield potential

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ABSTRACT

Predicting crop yields through simple methods would greatly aid crop breeding programs and could also be deployed at farm level to inform best crop management practices. This research proposes a new method for predicting wheat grain yields, along the crop growth cycle, based on canopy cover and reflectance indices, the Yield_p Model. The model was evaluated by comparing grain yields measured at harvest with the outputs of the proposed model using phenotypic data collected for a wheat population grown under UK field conditions for the 2015 and 2016 seasons. Accumulated radiation (RAD), Normalized Difference Vegetation Index (NDVI), Photochemical Reflectance Index (PRI), Water Index (WI), Harvest Index (HI) and canopy cover indices were the components of the model. Results suggested that the biomass accumulation predicted by the model was responsive along the crop cycle and the grain yield predicted was correlated to measured grain yield (r = 0.59; p < 0.001 for 2015; r = 0.64, p < 0.001 for 2016). The model allowed early prediction of grain yield based on biomass accumulated at anthesis (r = 0.54; p < 0.001 for 2015; r = 0.48, p < 0.001 for 2016). Evaluation of the model components enabled an improved understanding of the main factors limiting yield formation along the crop cycle. The proposed Yield_p Model explores a new concept of yield modelling and can be the starting point for the development of cheap and robust, onfarm, yield prediction during the crop cycle.

Keywords: Crop breeding, Early yield prediction, Mathematical modelling, On-farm yield, Remote sensing, *Triticum aestivum*

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AUTHORS' CONTRIBUTIONS

JPP, MAJP, DF and ECS designed research. JPP and ECS performed research. JPP and NV analysed data. JPP, NV, JPRADB, SPL, MAJP, DF and ECS wrote the paper.

INTRODUCTION

The increased demand for food motivated by the growing world population (Godfray et al. 2010) and changes in food consumption patterns (Pingali 2006) has increased the pressure on the agricultural sector. Increasing food production in a sustainable way, i.e. with minimal to no increases in land and water and nutrients inputs, emerges as the main solution for ensuring food security in the near future (FAO 2002). Improving yields is the main cornerstone for improving food security, as yield increases are expected to result in up to 77% of the increases in food production by 2050 (Alexandratos and Bruinsma 2012), while reducing agricultural impacts on the environment (Tilman and Clark 2015). Understanding yield formation and the main limitations to this process, at the farm level, is crucial to the success of breeding programs aiming to improve yield potential and resilience (Reynolds and Langridge 2016).

Yield potential (Yp) is defined as the yield of a cultivar when grown in environments to which it is adapted, with nutrients and water non-limiting, and with pests, diseases, weeds, lodging, and other stresses effectively controlled (Evans and Fischer 1999), and is theoretically presented as: $Y_p = RAD \cdot \epsilon i \cdot \epsilon c \cdot \epsilon p$ (Long et al. 2015). The equation represents the energy transformation process from sunlight to grain matter and the efficiencies (ϵ) involved in this process. The radiative energy from sunlight (RAD) is intercepted by the canopy (ϵ i) and converted (ϵc) into chemical energy, which is stored in the form of biomass. This biomass is then partitioned (ϵp) to the organ of interest (e.g. the grain, for the wheat crop) (Monteith and Moss 1977). However, the yield potential is not delivered at the farm level, mainly due to non-ideal growth conditions (Hengsdijk and Langeveld 2009). The difference between the yield potential and the on-farm yield is defined as the yield gap (Lobell et al. 2009).

Crop breeding programs, in the past decades, have been focusing mainly in the increase of yield potential (as discussed by Pennacchi et al. 2018b; Fischer et al. 2014), but future strategies must maintain the efforts to increase the yield potential while decreasing the yield gap (Araus et al. 2008). In breeding programmes, more than understanding yield formation, detecting high-yielding cultivars early in the crop cycle is important, as it improves the capacity of discarding unwanted genetic materials in the early stages and focus on promising genotypes. This is also necessary due to the increased capacity of genetic improvement and generation of different genotypes (Marti et al. 2007; Prasad et al. 2007; Becker and Schmidhalter 2017). Early yield prediction is also important for defining farm practices such as

fertilization (Marti et al. 2007) and informing the grain market and insurance companies (Balaghi et al. 2008).

To improve early yield prediction, remote sensing tools such as passive spectro-radiometers have been extensively used. Numerous studies have reported significant correlations between various spectral reflectance indices (SRIs) derived from spectral measurement and wheat grain yield (Raun et al. 2001; Babar et al. 2006; Marti et al. 2007; Balaghi et al. 2008; Peñuelas et al. 2011; Gizaw et al. 2016; Pradhan et al. 2017; Pennacchi et al. 2018a). However, their combinations in a mathematical model are not very frequent, although there are mentions in the literature (Raun et al. 2001; Singh et al. 2006; Montesinos-Lopez et al. 2017).

Certain SRIs reflect different canopy characteristics, which means they can be combined into a single model to fit the equation of theoretical definition of yield (Long et al. 2015). The NDVI (Normalized Difference Vegetation Index) is a vegetation index correlated to area cover by green tissue and relates to ɛi. The PRI (Photochemical Reflectance Index) is a pigment-related index correlated to dissipation of excess radiation and light conversion and thus relates to ɛc; HI (Harvest Index) is directly related to ɛp. Finally, WI (Water Index) correlates to the plant water status (Pietragalla et al. 2012). The use of WI in the model aims to include a term related to the general water status of the canopy and plant vigour. Although changes in water availability, and possible drought and heat stresses, will impact in NDVI and PRI, adding a component related to canopy water status may make the model more representative of the plant response and its potential impact to on-farm yield, mainly considering the high impact of drought in resource use efficiency and crop yield (Davies 2014). It may be true mainly considering the fact that water limitation is one of the key constraints of crop productivity (Dodd et al. 2011), and highly impacts yield (Jin et al. 2016; El-Hendawy et al. 2017).

Importantly, empirical models may not be particularly informative if their parameters have no intrinsic biological meaning (Adams et al. 2017). Simplicity and ease of application are also desirable characteristics of yield prediction models (Hoefsloot et al. 2012). Modelling crop yields in a single environment may prove challenging, mainly in conditions where there are no treatment factors. For instance, crop yield modelling for germplasm growing under different nitrogen concentrations or water regimes tends to present higher predictive power, as the additional factors may increase observation number and increase the spread of the data (Hernandez et al. 2015; Montesinos-Lopez et al. 2017). Working with germplasm in a single environment and in the absence of treatment levels relies solely on genetic variability.

In this study, the efficiency of multiple compositions of yield prediction models based on wheat canopy traits, such as reflectance parameters and canopy cover indices, was evaluated and compared to alternative statistical modelling methods. The model proposed was based on the yield formation equation and the underlying processes associated with the three efficiencies, ε_i , ε_c and ε_p (Long et al. 2015). It was applied to a wheat population grown in the field for two seasons. The dataset presented herein is part of a larger dataset already exploited for analysis of wheat yield drivers in the same population (Pennacchi et al. 2018a).

MATERIAL AND METHODS

Plant material and field experiments

A wheat population comprised of double-haploid lines (DHL) generated by Syngenta (Cambridge, UK) was used (Pennacchi et al. 2018a). The population, composed by the two parents and 119 lines, was grown at the Rothamsted Research farm, in Harpenden, UK, for two consecutive seasons, with the first sown in October 2014 and harvested in August 2015, and the second sown in October 2015 and harvested in August 2016. Each experiment was identified by the year of harvest and their details are presented in the sequence.

The 2015 experiment was planted at the Pastures field, in a Typical Batcombe soil (Avery and Catt 1995) after oilseed rape crop, in 2 x 1 m (2 m²) plots of 6 rows, with a sowing rate of 350 seeds m⁻², in three randomized blocks; sown on 20/10/2014 and harvested on 23/08/2015. The 2016 experiment was planted at the Delafield field, in a Batcombe soil (Avery and Catt 1995) after oilseed rape crop, in 4 x 1 m (4 m²) plots of 6 rows, with a sowing rate of 350 seeds m⁻², in three randomized blocks; sown on 12/10/2015 and harvested on 24/08/2016.

Application of fungicides, insecticides and herbicides, as well as fertilizers, followed Rothamsted farm practices.

Meteorological data

The meteorological data was acquired from the Rothamsted Meteorological Station at the Rothamsted Farm. The distance from the station to the experiments was, in a straight line: 1 km for the 2015 experiment and 1.6 km for the 2016 experiment. The daily radiation (MJ m⁻²) was recorded and the accumulated radiation (RAD) over a specific period was calculated as the sum of the daily value from the first to the last day in the considered period.

Crop development

The date at which half of the plants in a plot reached a given growth stage was monitored throughout the growing season using the Zadoks scale (Zadoks et al. 1974). The scale is based on scores relative to crop development stages: tillering, stem elongation, booting, flag leaf expansion, ear emergence, flowering, grain filling and maturation. The frequency of crop development monitoring depended on the crop stage, being less frequent when crop development was at early stages (from tillering (Z2) to booting (Z4) stages) and more frequent from booting (Z4) to dough development (Z8) as the crop development was faster. Leaf senescence was measured from anthesis to the end of the season using the wheat senescence scale (Pask and Pietragalla 2012).

Phenotyping

Measurements were taken over the crop growing season as described below. For canopy cover, horizontal photographs of the canopy from above, parallel to the soil surface of the plot, were taken weekly from March (Z2.4) to August (Senescence score 10, S10) using a digital camera. The pictures were analysed using the BreedPix open access software that outputs the area covered by green tissue as a percentage of the total area, i.e. the canopy cover (CC) (Casadesus et al. 2007). From this data, the following indices were calculated (Fig. 1):

Early vigour (EV): based on Rebetzke & Richards (1999) and calculated as the sum of the weekly single measurements of leaf cover from Z2.4 and the date when plots reached an average of 90% of area covered by leaves (90C) (Equation 1):

$$EV = (\sum_{Z2.4}^{90C} CC)$$
 (Equation 1).

Accumulated green area (AGA): calculated as the sum of the weekly single measurements of canopy cover from Z2.4 to Z6.5 (Equation 2):

$$AGA = \left(\sum_{Z2.4}^{Z6.5} \text{CC}\right) \qquad \text{(Equation 2)}.$$

Stay green (SG): based on Thomas & Smart (1993) and calculated as the sum of the weekly single measurements of canopy cover from Z6.5 to S10 (Equation 3):

$$SG = \left(\sum_{Z6.5}^{S10} \text{CC}\right) \qquad \text{(Equation 3)}.$$

Reflectance was measured 75 cm above the canopy using a HandySpec System (TEC5, Oberursel, Germany) spectroradiometer. From the reflectance measurements, the following

traits were calculated using the software accompanying the HandySpec System and according to Pietragalla et al. (2012):

Normalized Difference Vegetation Index (NDVI) (Equation 4):

NDVI =
$$(R_{900} - R_{680})/(R_{900} + R_{680})$$
 (Equation 4).

Photochemical Reflectance Index (PRI) (Equation 5):

 $PRI = (R_{530} - R_{570})/(R_{530} + R_{570})$ (Equation 5).

Water Index (WI) (Equation 6):

 $WI = (R_{970})/(R_{900})$ (Equation 6),

where R_{λ} is the reflectance measured at the wavelength λ (nm).

For the 2015 season, reflectance was measured at 3 time points: Z3.4, Z4.5 and Z6.5. For the 2016 season, the measurements were performed at 10 time points: Z3.7, Z5.7, Z6.5, Z7.1, Z7.5, Z7.8, Z7.9, Z9.4, Z9.7 and Z9.9.

The analysis of biomass at the end of the growing was performed at physiological maturity (Z9.9). The tillers in a 50 cm (2015) or 30 cm (2016) row were manually harvested from the third quarter (middle section) of the third row in each of the 6-row plots. The number of tillers and grains, as well as the straw, spike and grain dry mass and grain moisture were measured (Pask and Pietragalla 2012). Harvest index (HI) was calculated as the ratio between dry grain mass (GM) and above ground biomass (AGB, consisting of grain and straw mass), all measured in grams (g) and at 100% dry matter (Equation 7):

$$HI = GM/AGB$$
 (Equation 7).

For the final harvest of the experiments, plants were harvested using a Haldrup-C65 (Haldrup, Le Mans, France) plot combine. Grain mass for each plot was determined by the combine. Grain moisture was measured using a sub-sample of grains from each plot, at harvest time, and grain mass was normalized to 15% moisture content. Grain mass per plot was corrected for the sections harvested by hand and grain yield estimated in tons per hectare at 85% dry matter.

Grain yield modelling

Models were fit starting from the simplest and progressing towards increasing complexity with the inclusion of additional traits. The simplest model (Model 1) to predict wheat grain yield included NDVI, PRI and HI. PRI was embedded within the calculation of light use efficiency (LUE) using the conversion factor defined by Wu et al. (2015) (Equation 8):

LUE = (6.6.PRI) + 1.1 (Equation 8).

Thus, Model 1 was defined as:

Biomass_{M1} = \sum_{i}^{f} (RAD.NDVI.LUE) (Equation 9) Yield_{M1} = Biomass_{M1}.HI (Equation 10),

where $Biomass_{M1}$ is the predicted accumulated biomass, Yield_{M1} is the predicted yield, RAD is the sunlight radiation in the given crop growing period, LUE is the light use efficiency calculated from PRI, and HI is the harvest index; *i* and *f* are initial and final measurement points, respectively.

A second model (Model 2) was built from Model 1 by incorporating a factor related to canopy water. The Water Status index (WS) was calculated as the inverse of WI (Equation 11). WI is negatively related to canopy water; therefore, higher WI corresponds to lower canopy water content:

WS = 1/WI (Equation 11).

The Model 2 was defined as:

Biomass_{M2} = \sum_{i}^{f} (RAD.NDVI.LUE.WS) (Equation 12), Yield_{M2} = Biomass_{M2}.HI (Equation 13).

A third model (Model 3) was built from Model 1, by using a correction factor for NDVI (NDVI_{cor}). NDVI_{cor} was calculated from NDVI multiplied by a factor related to canopy cover indices (Equation 14). The correction factor for each line was calculated by the chosen canopy cover index for the line divided by the average of the index for the population. The indices used for each of the periods in the season were: EV from Z3.4 to 90C, AGA from 90C to Z6.5 and SG from Z6.5 to Z9.9 (Fig. 1).

NDVI_{cor} = NDVI.k (Equation 14),

where *k* is the correction factor defined as (Equation 15):

k = [(EV₁/EV_p) from Z2.4 to 90C; (AGA₁/AGA_p) from 90C to Z6.5; (SG₁/SG_p) from Z6.5 to Z9.9]

where *I* and *p* are, respectively, the line and population mean values for the trait at each time point.

The Model 3 was defined as:

Biomass_{M3} = \sum_{i}^{f} (RAD.NDVI_{cor}.LUE) (Equation 16),

 $Yield_{M3} = Biomass_{M3}.HI$ (Equation 17).

The final model (Model 4) was the most complex, resulting from a combination of Models 2 and 3. The Model 4 was defined as:

$$Biomass_{M4} = \sum_{i}^{f} (RAD.NDVI_{cor}.LUE.WS)$$
(Equation 18),
Yield_{M4} = Biomass_{M4}.HI (Equation 19).

The Model 4, from here named Yield_p Model, can also be written as:

Yield_p =
$$[\sum_{i}^{f} (RAD.NDVI_{cor}.LUE.WS)]$$
.HI (Equation 20).

As the first measurements were made at stem elongation (Z3.4 for 2015 and Z3.7 for 2016), values for NDVI, PRI and WI were estimated for the early stages when crop growth restarted after winter (around Z2.4). NDVI at Z2.4 was estimated as 35% of the NDVI at Z3.4 (or Z3.7 for 2016) (based on unpublished data collected by the phenotyping group at Rothamsted Research), PRI and WI were estimated as the same values as at Z3.4 (or Z3.7 for 2016). Biomass accumulated at Z2.4 was incipient and considered as zero, being the biomass accumulation counted from this stage. For 2015, the final values for Z9.9 were also estimated, with NDVI as 0.13, LUE as 0.30 and WI as 1.1 (according to average measured values in 2016 season).

The unit of calculation of Yield_p was based on the components of the Equations 18 and 19. $NDVI_{cor}$ and WI were measured in percentage and have no unit. RAD was measured in MJ m⁻² and LUE was measured in g C MJ⁻² (grams of Carbon per square meter). As HI is also unitless, Biomass and Yield_p are given in g C m⁻² or converted to t C ha⁻¹.

Model Sensitivity evaluation

Each of the models was evaluated using the Pearson Product Moment (PPM) correlation coefficients (*r*) and the root mean square error (RMSE). The PPM coefficient represented the correlation between grain yield predicted by each model and measured grain yield. The same approach was used to evaluate the correlation between single traits and measured grain yield. The traits used in the model and the model output (predicted biomass and predicted yield at the end of the season) were compared to measured yield at each of the evaluation time points in 2015 and 2016. The RMSE is defined as the square root of the sum of the squared difference between predicted and measured yield for each observation (line). It was calculated for all the models as:

RMSE =
$$V[\sum_{1}^{n} (\text{Yield}_{p} - \text{Yield}_{m})^{2}]$$
 (Equation 21),

where n is the number of lines in the experiment (n = 121) and Yield_m the measured yield for each line.

The use of the two methods of evaluation (PPM and RMSE) allows an integrated and robust analysis of model fitting. The Pearson correlation coefficients (*r*) are related to the model capacity to distinguish the phenotypic response of the cultivars in terms of their grain yield. However, it does not account for the amplitude of the predicted values. The RMSE values accounted for the difference in amplitude between predicted and measured values. An ideal model would present high Pearson correlation coefficient and low RMSE values.

To evaluate the impact of ε i, ε c and ε p in the prediction of yield, each of the terms (NDVI_{cor}, LUE and HI) was individually taken out of Yield_p Model (Equation 20) and the output was compared to Yield_m using PPM correlation coefficient (r) and RMSE. These models are referred as Reduced Models (Yield_{pR1} to Yield_{pR3}).

To evaluate the impact of increasing the phenotyping frequency, especially after anthesis, a short model was run for 2016 using only the initial predicted time point (Z2.4), three middle time points (Z3.7, Z5.7 and Z6.5) and the last time point (Z9.9). The output was compared to measured grain yield using PPM correlation coefficient (r) and RMSE. This model is referred as Short Model and is only presented for 2016 season.

Multiple linear regression analysis

A multiple linear regression was independently fit for each of the seasons based in the same traits included in the previous presented model: NDVI, PRI, WI, HI, EV, AGA and SG. Initially, we performed the Forward and Backward Stepwise methods which resulted in a list of

parameters to be included in a further model. A final multiple linear regression was fit using a combination of multiple modelling methods (Accumulated, Pooled, Forward Selection, Backward Selection, Forward Stepwise and Backward Stepwise). The final multiple linear regression fitting process was based on fixed and random terms. Fixed terms (terms always present in the model) were the traits which were coincident in both, the Forward and Backward Stepwise methods; random terms (that can be present or absent from the model) were traits which were present or absent from the model) were traits which were present or absent from the model) were traits which were present in one of the methods but not in the other. Regression fitting was analysed based on the adjusted R-square (R²_{adj}) and the Mallows Cp coefficient. These models are referred as Multiple Linear Regression (MLR). The GenStat 17th Edition (VSN International Ltd., Hemel Hampstead, UK) was used for all the statistical analysis presented in this paper.

Data analysis

The method of residual maximum likelihood (REML) was used to evaluate spatial trends over the rows and columns in the field design by fitting a linear mixed model to each measured trait to test for any statistically significant (p < 0.05, Chi-squared test) variation. Predicted means from the model fitted to each trait were used in subsequent modelling.

RESULTS

The inclusion of canopy cover and plant water status indices improves the predictive power of grain yield models

Amongst the four tested approaches to describe the wheat yield from remote sense reflectance indices, from low to high complexity, Model 1 presented a correlation to measured yield of 0.50 and 0.57 for 2015 and 2016, respectively (Table 1). The use of canopy cover corrections to NDVI (NDVI_{cor}) generated an improvement in the fitting of the model from 0.50 (p < 0.001) to 0.57 (p < 0.001) in 2015 and from 0.57 (p < 0.001) to 0.60 (p < 0.001) in 2016 (comparison between Model 1 and 3, Table 1). The improvement was more evident in 2015, which suggests that inclusion of the canopy cover indices is most important when reflectance measurements were less frequent.

The use of the WS index (inverse of WI) as an indicator of canopy water status also improved the correlation between predicted and measured yield. For 2015, including WS increased the fitting from 0.50 (p < 0.001) to 0.52 (p < 0.001), and for 2016 from 0.57 (p < 0.001) to 0.62 (p < 0.001) (comparison between Model 1 and 2, Table 1).

The combined insertion of both NDVI_{cor} and WS improved the correlation between the predicted and measured yields to 0.59 (p < 0.001) in 2015 and to 0.64 (p < 0.001) in 2016 (comparison between Model 1 and Yield_p Model, Table 1).

The inclusion of NDVI_{cor} did not change the RMSE greatly as it did not affect the amplitude of the predicted values from Model 1 to Model 3. On the other hand, the inclusion of WS had a greater impact in the RMSE as it changed not just the predicted value for each line, but also the general amplitude of the model output. For 2015, the inclusion of WS caused a reduction of the amplitude between the predicted and measured values, with a reduction of RMSE from 1.66 to 0.97. For 2016, the opposite happened with the RMSE changing from 0.42 to 1.75 (Table 1). This contrasting response is associated with the underestimation of predicted yield in 2015 and overestimation in 2016 (Fig. 2). Overall, the Yield_p Model showed an improved fitting for both years, while it presented an increased RMSE in 2016 (Table 1). The Yield_p Model will be used as the basis for the further analysis presented in the manuscript, unless otherwise stated.

The wheat yield prediction model enhances the understanding of grain yield formation

The proposed Yield_p Model presented higher correlation to measured grain yield than any other single trait along the crop cycle in both seasons (Table 2). For the 2015 season, the predicted Yield_p and predicted Biomass presented, respectively, a correlation of 0.59 (p < 0.001) and 0.56 (p < 0.001), which was higher than the maximum correlations of any measured and/or calculated traits (NDVI (Z3.4), r = 0.48, p < 0.001; LUE (Z3.4), r = 0.47, p < 0.001; respectively) (Table 2). For 2016, Yield_p (r = 0.64, p < 0.001) presented higher correlation to grain yield than the measured and calculated traits, WI (Z7.4) (r = -0.59, p < 0.001) and LUE (Z3.7 and Z5.7) (r = 0.44, p < 0.001), respectively (Table 2). The use of the model improved the capacity of predicting yield if compared to single traits by 11% (0.59-0.48=0.11) and 5% (0.64-0.59=0.05), in 2015 and 2016, respectively.

The correlation between predicted biomass and yield increased along the crop cycle reaching its maximum value around flowering and grain development stages. For 2015, the maximum value was at anthesis (Z6.5; r = 0.54, p < 0.001), but was also high at booting (Z4.5; r = 0.53, p < 0.001). For 2016, the maximum correlation was at grain development (Z7.1, Z7.5 and Z7.9; r = 0.50, p < 0.001) but was close to that at anthesis (Z6.5; r = 0.48, p < 0.001) and ear emergence (Z5.7; r = 0.47, p < 0.001) (Table 2).

Sensitivity analysis did not reveal a main influencer for yield prediction

The sensitivity analysis of the Yield_p Model, through the evaluation of the Reduced Models, did not reveal a main influencer for yield prediction. In 2015, the NDVI_{cor} presented the biggest influence to the model, followed by LUE and HI. In 2016, HI had a bigger influence to yield prediction, followed by NDVI_{cor} and LUE. This lack of a consistent pattern for the influence of individual terms to the final model highlights the flexible response of the model to different seasons (Table 3). Importantly, removing NDVI_{cor} and, especially HI of the model had a large impact on the RMSE value (Table 3), demonstrating that both terms cause a large change in the amplitude of predicted values, which is not observed for LUE. For instance, a HI of 0.5 means that half of the above ground biomass was stored in the grain. A model without HI would generate predicted yield values of an amplitude of the double of the measured value, thus increasing RMSE.

Increased phenotyping frequency, especially at post-anthesis, improved model fitting

The comparison between Yield_p Model and the Short Model for 2016 allowed an analysis of the impact of the phenotyping frequency in the model fitting, as follow: the Yield_p Model, for 2016, presented a correlation of 0.64 (p<0.001) with a RMSE of 1.75; for the Short model, the correlation was reduced to 0.31 (p<0.001) and the RMSE to 1.54. The increased phenotyping frequency, mainly at post-anthesis, enhanced the fitting of the model for 2016. The improvement of the model fitting promoted by an intensification of phenotyping at post-anthesis cannot be pointed as a general conclusion, as it must be tested for different seasons. The fact that the modelling for 2015, using the same number of time points as the short model for 2016 presented a higher fitting, suggests the need of evaluating the impact of post-anthesis phenotyping for more years, in repeated experiments.

The Yield, Model highlights the limitations to yield formation throughout the crop cycle

For the 2016 season, the high frequency of measurements enabled an evaluation of the factors limiting yield formation (Fig. 3). Biomass accumulation was responsive to high NDVI and LUE, until ear emergence (Z5.7). Maintenance of NDVI and an increase in WS, as well as higher solar radiation, kept the biomass accumulation rate until Z7.5, despite a decreased LUE. From Z7.5, biomass accumulation rate decreased with the decrease of NDVI, WS and LUE (Fig. 3).

Multiple Linear Regressions were dependent on phenotyping capacity

The use of MLR to predict grain yield using the same traits presented in the Yield_p Model allowed a comparison between the methods, as presented in the sequence. For 2015, the MLR, composed of 7 terms presented $R^2_{adj} = 37.9\%$ (r = 0.62). For 2016, the Multiple Linear Regression composed of 9 terms presented $R^2_{adj} = 67.3\%$ (r = 0.82). Although the MLR presented higher correlations to measured grain yield in both seasons, a high variation in the model fitting between 2015 and 2016 was observed. The frequency of phenotyping impacted greatly the reliability of the regressions.

For 2015, the terms presented in the MLR were: AGA, SG, NDVI (estimated at Z2.4), PRI (Z6.5), WI (Z6.5) and HI. For 2016, the terms were: SG, NDVI (estimated at Z2.4, Z9.9), PRI (Z6.5, Z7.4, Z7.8, Z9.9), WI (estimated at Z2.4, Z7.4, Z7.8, Z9.4, Z9.7) and HI. Although some terms contribute to the predictive regressions presented above, they did not present biological meaning, as for instance, canopy WI at the very late stages in the season (Z9.4 and Z9.7), close to harvest. Four terms were presented in the regressions for both years: SG, NDVI (estimated at Z3.4), PRI (Z6.5) and HI. When the linear regression analysis was performed with only these four terms, the final fitting was of $R^2_{adj} = 30.3\%$ (r = 0.55) and $R^2_{adj} = 41.5\%$ (r = 0.64) for 2015 and 2016, respectively. These results were very similar to the general fitting of the Yield_p Model proposed in this article (r = 0.59 in 2015 and r = 0.64 in 2016) (Table 2).

DISCUSSION

The use of a model to calculate wheat grain yield had the main objective of enabling robust predictions of final yield. An additional aim was to better understand the factors contributing to yield formation and the influence of particular combinations of traits measured during the crop development cycle to the final productivity. There are many factors limiting biomass accumulation and yield formation during the crop cycle, but they will essentially, constrain the efficiencies presented in the yield formation equation. The use of a simplistic model could help enhance the understanding of the limitations to yield along the crop cycle.

The Yield_p Model, based on the yield formation equation, was developed to predict wheat grain yield using canopy reflectance indices. The model was validated using a wheat population grown and phenotyped under UK field conditions, for two consecutive seasons. The Yield_p Model output presented higher correlation to measured grain yield than any of the single traits measured in the study. Significant correlations between wheat yield and NDVI (Raun et al. 2001; Gizaw et al. 2016; Pennacchi et al. 2018a), PRI (Peñuelas et al. 2011; Gizaw et al. 2018a) or WI (Pradhan et al. 2015; Pennacchi et al. 2018a) have

been previously reported. Here, the mathematical combination of these parameters and canopy cover indices in a simple model has shown an improvement in yield prediction, with correlation coefficients of r = 0.59 ($R^2 = 0.35$, p < 0.001) and 0.64 ($R^2 = 0.41$, p < 0.001) for 2015 and 2016, respectively.

The correlation between predicted and measured yield was intermediate, although not particularly high. This may be related to the fact that the models were applied to single environment conditions with the genotypic composition of the lines as the only source of variation. The genetic variability in a double-haploid population is not expected to be as high as in diverse germplasm panels (Gaynor et al. 2017).

The inclusion of canopy cover indices correction to NDVI improved the Yield_p Model fitting, mainly for the 2015 season, when the number of measurements was lower compared to 2016. This can be useful to improve yield prediction models in field situations where reflectance measurements cannot be frequently performed and highlights the importance of access to simple and easy to deploy methods of phenotyping (Casadesus et al. 2007) for farmers with limited access to phenotyping equipment throughout the growing season. The inclusion of a water index (WI) also improved fitting. The 2% increase in *r* when WI was included might seem trivial, but may acquire greater proportions under conditions of increasing water deficit, which was not the particular cases in the two seasons evaluated herein. Moreover than a canopy water status index, WI may be important to inform about stomata control to the diffusive process and help to better predict changes in ε (Pietragalla et al. 2012). In combination with NDVI and PRI, WI may also be an indicator of plant vigour and help to infer about lines with increased resilience to drought and heat stresses or even though to help of the decision of irrigation practices.

The use of WI as part of the model during the whole crop development may be subject to further investigation as, at the later stages of the cycle, a rapid decrease in canopy water status may be favourable to grain maturation and prevention of diseases (AHDB, 2016).

One of the strengths of the proposed yield prediction model is the capacity to predict grain yields as early as at anthesis. In both growing seasons studied, the Yield_p Model predicted biomass accumulated at anthesis (Z6.5) presented strong correlation to measured grain yield. The detection of early predictors of grain yield is a major challenge to plant breeding; the proposed model will decrease the plant selection time, reducing costs and speeding up the release of high yielding cultivars (Becker and Schmidhalter 2017). A third important

contribution of the present study is the capacity to inform the main yield limitation in specific crop stages, which could improve to the understanding of climatic constrains to grain yield. For instance, in 2016, LUE appeared to be the main limitation after ear emergence (Fig. 3), which could be minimized by the maintenance of photosynthetic response from booting to anthesis, as reported by Carmo-Silva et al. (2017).

The comparison of the Yield_p Model with MLR methods allowed an analysis of its advantages and disadvantages, by comparing their specific fittings. Although the MLR methods presented higher correlation to measured yield, they presented different conformations for each of the analysed seasons. The advantage of the Yield_p Model in relation to MLRis the fact that it responded particularly well for both seasons with the same conformation, showing its robustness and capacity of application on-farm. The Yield_p Model also allowed the estimation of yield before the end of the season being more efficient as a predictive model. The capacity of predicting final yield at early stages in the crop cycle will be helpful to inform farmers about crop practices such as late fertilization or crop protection. Contrastingly, MLR methods may be more recommended for a post-harvest analysis of the main yield drivers in a specific season.

Some factors have to be considered when phenotyping with the aim of yield prediction. NDVI, PRI and WI are influenced by the canopy albedo, which can have diurnal and seasonal variations in wheat (Zhang et al. 2013). SRIs, in general, are affected by external factors as light conditions (overcast), sun position, and also wind. The change in canopy structure as the emergence of the ears with awns, or not, as well as the anthers may affected the signal measured. An index like PRI can also present rapid variation according to the light intensity, as well as the air temperature (Gammon et al. 1997; Dobrowski et al. 2005), mainly considering the normal spatial-temporal variation of photosynthesis (Neto et al. 2021). Pennacchi et al. (2018a) also reported high variability for PRI values in the 2015 experiment with the same wheat population. Starting reflectance measurements at early developmental stages, taking measurements at similar weather conditions and time of day, and reducing the time to cover the whole experimental field could potentially improve model fitting by reducing experimental error.

High-throughput methods including phenotyping platforms (Virlet et al. 2017) and unmanned aerial vehicles (UAVs) (Yang et al. 2017) with multi-spectral cameras could improve the data acquisition, the understanding of space-time plant adaption (Galviz et al. 2022) and

consequently the model fitting. An increased phenotyping frequency could reduce the influence of each single time point on the final prediction of the model, mainly considering the uncertainty on predicting canopy characteristics in the interval between measurements. The use of SRI calculated from satellite images may be a great advance to yield modelling at field level (Hoefsloot et al. 2012), mainly due to the availability of free-source data as the Landsat database (Wulder et al. 2012). The adaption of the Yield_p Model to satellite imaging data may allow a further step ahead in yield prediction.

Further improvements to the Yield_p Model could be achieved by the insertion of components that could help explain the main efficiencies involved in the yield formation equation (Long et al. 2015) and the impact of non-ideal growth conditions at the farm level. Moreover, the use of genetic data, related to the traits in the model, could be informative of the genotype-phenotype-environment multiple interactions (Yin and Struik 2010) and improve yield modelling under drought and heat conditions (Parent and Tardieu 2014).

CONCLUSIONS

The Yield_p Model was responsive to crop growth and development and capable of predicting biomass and grain yield early in the season with an intermediate fit. Advantages of the proposed model include simplicity, ease of use and low cost of the phenotyping and modelling techniques. Finally, the model presents biological meaning as it is based on the yield formation equation. The application of the model to multiple datasets in different locations and contrasting climatic conditions warrants further study and is likely to reveal new potential for improvement and expanding its practical applications.

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REFERENCES

Adams M.P., Collier C.J., Uthicke, S., Ow Y-X., Langlois L. & O'Brien K.R. (2017) Model fit versus biological relevance: Evaluating photosynthesis-temperature models for three tropical seagrass species. *Scientific Reports*, **7**, 1-12

AHDB (2016) Wheat disease management guide. Warwickshire, UK: Agriculture and Horticulture Development Board.

Alexandratos N., Bruinsma J. (2012) World agriculture towards 2030/2050: the 2012 Revision. ESA Working Paper No. 12–03. Agricultural Development Economics Division, Food and Agriculture Organization of the United Nations. Rome

Araus J.L., Slafer G.A., Royo C. & Serret M.D. (2008) Breeding for yield potential and stress adaptation in cereals. *Critical Reviews in Plant Science*, **27**, 377-412

Avery B.W. & Catt J.A. (1995) The Soil at Rothamsted, Map prepared by E M Thompson and the Soil Survey and Land Research Centre, Cranfield University, Lawes Agricultural Trust, Harpenden, UK

Babar M.A., Reynolds M.P., van Ginkel M., Klatt A. R., Raun W.R. & Stone M.L. (2006) Spectral Reflectance Indices as a Potential Indirect Selection Criteria for Wheat Yield under Irrigation. *Crop Sciences*, **46**, 578-588

Balaghi R., Tychon B., Eerens H. & Jlibene M. (2008) Empirical regression models using NDVI, rainfall and temperature data for the early prediction of wheat grain yields in Morocco. *International Journal of Applied Earth Observation and Geoinformation*, **10**, 438-452

Becker E. & Schmidhalter U. (2017) Evaluation of Yield and Drought Using Active and Passive Spectral Sensing Systems at the Reproductive Stage in Wheat. *Frontiers in Plant Sciences*, **8**, 1-15

Carmo-Silva, E., Andralojc, P.J., Scales J.C., Driever S.M., Mead A., Lawson T., Raines C.A. & Parry M.A.J. (2017) Phenotyping of field-grown wheat in the UK highlights contribution of light response of photosynthesis and flag leaf longevity to grain yield. *Journal of Experimental Botany*, **68**, 3472-3486

Casadesus J., Kaya Y., Bort J., Nachit M.M., Arau, J.L., Amor S., Ferrazzano G., Maalouf F., Maccaferri M., Martos V., Ouabbou H. & Villegas D. (2007). Using vegetation indices derived from conventional digital cameras as selection criteria for wheat breeding in water-limited environments. *Annals of Applied Biology*, **150**, 227-236

Davies, W.J. (2014) Exploiting plant drought stress biology to increase resource use efficiency and yield of crops under water scarcity. *Theor. Exp. Plant Physiol*, **26**, 1–3

Dobrowski S.Z., Pushnik J.C., Zarco-Tejada P.J. & Ustin S.L. (2005) Simple reflectance indices track heat and water stress-induced changes in steady-state chlorophyll fluorescence at the canopy scale. *Remote Sensing of Environment*, **97**, 403-414

Dodd I.C., Whalley W.R., Ober E.S. & Parry M.A.J. (2011) Genetic and management approaches to boost UK wheat yields by ameliorating water deficits. *Journal of Experimental Botany*, **62**, 5241-5248

El-Hendawy S.E., Hassan W.M., Al-Suhaibani N.A. & Schmidhalter U. (2017) Spectral assessment of drought tolerance indices and grain yield in advanced spring wheat lines grown under full and limited water irrigation. *Agricultural Water Management*, **182**, 1-12

Evans L.T. & Fischer R.A. (1999) Yield potential: its definition, measurement and significance. *Crop Science*, **39**, 1544-1551

FAO (2002) The state of food insecurity in the world. ESA Working Paper. Rome

Fischer R.A., Byerlee D. & Edmeades G.O. (2014) Crop yields and global food security: will yield increase continue to feed the world? ACIAR Monograph No. 158. Canberra

Gaynor R.C., Gorjanc G., Bentley A.R., Ober E.S., Howell P., Jackson R., Mackay I.A. & Hickey J.M. (2017) A Two-Part Strategy for Using Genomic Selection to Develop Inbred Lines. *Crop Science*, **57**, 2372-2386

Galviz Y., Souza G.M. & Lüttge U. (2022) The biological concept of stress revisited: relations of stress and memory of plants as a matter of space–time. *Theor. Exp. Plant Physiol.*, **34**, 239–264

Gizaw S.A., Garland-Campbell K. & Carter A.H. (2016) Evaluation of agronomic traits and spectral reflectance in Pacific Northwest winter wheat under rain-fed and irrigated conditions. *Field Crops Research*, **196**, 168-179

Godfray H.C.J., Beddington J.R., Crute I.R., Haddad L., Lawrence D., Muir J.F., Pretty J., Robinson S., Thomas S.M. & Toulmin C. (2010) Food Security: The challenge of feeding 9 billion people. *Science*, **327**, 812-818

Hengsdijk H. & Langeveld J.W.A. (2009) Yield trends and yield gap analysis of major crops in the world. Wettelijke Onderzoekstaken Natuur & Milieu, Wageningen.

Hernandez J., Lobos G.A, Matus I., del Pozo A., Silva P. & Galleguillos M. (2015) Using Ridge Regression Models to Estimate Grain Yield from Field Spectral Data in Bread Wheat (*Triticum aestivum* L.) Grown under Three Water Regimes. *Remote Sensing*, **7**, 2109-2126

Hoefsloot P., Ines A., van Dam J., Duveiller G., Kayitakire F., Hansen J. (2012) Combining Crop Models and Remote Sensing for Yield-prediction: Concepts, Applications and Challenges for Heterogeneous Smallholder Environments. Publications Office of the European Union, 52 pp. Luxembourg

Jin X., Kumar L., Li Z., Xu X., Yang G., & Wang J. (2016) Estimation of Winter Wheat Biomass and Yield by Combining the AquaCrop Model and Field Hyperspectral Data. *Remote Sensing*, **8**, 972-984

Lobell D.B., Cassman K.G. & Field C.B. (2009) Crop yield gaps: their importance, magnitudes, and causes. *Annual Review of Environment and Resources*, **34**, 179-204

Long S.P., Marshall-Colon A. & Zhu X.G. (2015) Meeting the global food demand of the future by engineering crop photosynthesis and yield potential. *Cell*, **161**, 56-66

Marti J., Bort J., Slafer G.A. & Araus J.L. (2007) Can wheat yield be assessed by early measurements of Normalized Difference Vegetation Index? *Annals of Applied Biology*, **150**, 253-257

Monteith J.L. & Moss C.J. (1977) Climate and the efficiency of crop production in Britain. *Philosophical Transactions of the Royal Society of London B*, **281**, 277-294

Montesinos-López O.A., Montesinos-López A., Crossa J., de los Campos G., Alvarado G., Suchismita M., Rutkoski J., González-Pérez L. & Burgueño J. (2017) Predicting grain yield using canopy hyperspectral reflectance in wheat breeding data. *Plant Methods*, **13**, 4-27

Neto M.C.L., Carvalho F.E.L., Souza G.M. & Silveira J.A.G. (2021) Understanding photosynthesis in a spatial–temporal multiscale: The need for a systemic view. *Theor. Exp. Plant Physiol*, **33**, 113–124

Parent B. & Tardieu F. (2014) Can current crop models be used in the phenotyping era for predicting the genetic variability of yield of plants subjected to drought or high temperature? *Journal of Experimental Botany*, **65**, 6179-6189

Pask A. & Pietragalla J. (2012) Leaf area, green crop area and senescence. *In: Physiological Breeding II: A Field Guide to Wheat Phenotyping*. CIMMYT. Mexico, D.F. pp. 58-62

Pennacchi J.P., Carmo-Silva E., Andralojc P.J., Feuerhelm D., Powers SJ. & Parry M.A.J. (2018a) Dissecting wheat grain yield drivers in a mapping population in the UK. *Agronomy*, **8**, 94-108

Pennacchi J.P., Carmo-Silva E., Andralojc P.J., Lawson T., Allen A.M., Raines C.A. & Parry M.A.J. (2018b) Stability of wheat grain yields over three field seasons in the UK. *Food and Energy Security*, 8:e00147.

Peñuelas J., Garbulsky M.F., Filella I. (2011) Photochemical reflectance index (PRI) and remote sensing of plant CO₂ uptake. *New Phytologist*,**191**, 596-599

Pietragalla J., Mullan D. & Mendoza R.S. (2012) Spectral reflectance. *In: Physiological Breeding II: A Field Guide to Wheat Phenotyping*. CIMMYT. Mexico, D.F. pp. 32-36

Pingali P.L. (2006) Westernization of Asian diets and the transformation of food systems: Implications for research and policy. *Food Policy*, **32**, 281-298

Pradhan S., Bandyopadhyay K.K., Sahoo R.N., Sehgal V.K., Singh R., Gupta V.K. & Joshi D.K. (2014) Predicting Wheat Grain and Biomass Yield Using Canopy Reflectance of Booting Stage. *Journal of the Indian Society of Remote Sensing*, **42**, 711-718.

Prasad B., Carver B.F., Stone M.L., Babar M.A., Raun W.R. & Klatt A.R. (2007) Genetic Analysis of Indirect Selection for Winter Wheat Grain Yield Using Spectral Reflectance Indices. *Crop Sciences*, **47**, 1416-1425

Raun W.R., Johnson G.V., Stone M.L., Solie J.B., Lukina E.V., Thomason W.E. & Schepers, J. S. (2001) In-season prediction of potential grain yield in winter wheat using canopy reflectance. *Agronomy Journal*, **93**, 131-138

Rebetzke G.J. & Richards R.A. (1999) Genetic improvement of early vigour in wheat. *Australian Journal of Agricultural Research*, **50**, 291-301

Reynolds M. & Langridge P. (2016) Physiological breeding. *Current Opinion in Plant Biology*, **31**, 162-171

Singh M., Niwas R., Khichar M.L. & Yadav M.K. (2006) Spectral Models for Estimation of Chlorophyll Content, Growth and Yield of Wheat Crop. *Journal of the Indian Society of Remote Sensing*, **34**, 1-5

Thomas H. & Smart, C.M. (1993) Crops that stay green. *Annals of Applied Biology*, **123**, 193-201

Tilman D. & Clark M. (2015) Food, agriculture & the environment: Can we feed the world & save the earth? *American Academy of Arts & Sciences*, **144**, 1-23

Virlet N., Sabermanesh K., Sadeghi-Tehran P. & Hawkesford MJ (2017). Field Scanalyzer: An automated robotic field phenotyping platform for detailed crop monitoring. *Functional Plant Biology*, **44**, 143–153

Wu C., Huang W., Yang Q. & Xie Q (2015) Improved estimation of light use efficiency by removal of canopy structural effect from the photochemical reflectance index (PRI). *Agriculture, Ecosystems and Environment*, **199**, 333-338

Wulder M.A., Masek J.D., Cohen W.B., Loveland T.R. & Woodcock C.E (2012) Opening the archive: How free data has enabled the science and monitoring promise of Landsat. *Remote Sensing of Environment*, **122**, 2-10

Yang G., Liu J., Zhao C., Li Z., Huang W., Yu H., Xu B., Yang X., Zhu D., Zhang X., Zhang R., Feng H., Zhao X., Li Z., Li H. & Yang H. (2017) Sensing for Field-Based Crop Phenotyping: Current Status and Perspectives. *Frontiers in Plant Sciences*, **8**, 1-26.

Yin X., Struik P.C (2010) Modelling the crop: from system dynamics to systems biology. *Journal of Experimental Botany*, **61**, 2171-2183

Zadoks I.C., Chang T.T. & Konzak C.F. (1974) A decimal code for the growth stages of cereals. *Weed Research*, **14**, 415-421

TABLES

Models	Model description	2015		2016	
Widdels	Model description	r	RMSE	r 0.57	RMSE
Model 1	Yield _{M1} = RAD.NDVI.LUE.HI	0.50	1.66	0.57	0.42
Model 2	Yield _{M2} = RAD.NDVI.LUE.WS.HI	0.52	0.97	0.62	1.75
Model 3	Yield _{M3} = RAD.NDVI _{cor} .LUE.HI	0.57	1.66	0.60	0.43
Yield Model (Model 4)	Yield _p = RAD.NDVIcor.LUE.WS.HI	0.59	0.99	0.64	1.75

Table 1. Evaluation indices for yield prediction models (Model 1 to 4) in relation to measured grain yield

Significance levels for correlations are given by an F-test on 1 and 120 degrees of freedom: p < 0.001. r, Pearson correlation coefficient between predicted and measured yield for each proposed model; RMSE, Root Mean Square Error for each proposed model. Yield_p#, yield predicted by the proposed Model #; HI, harvest index; NDVI_{cor}, corrected Normalized Difference Vegetation Index; LUE, Light Use Efficiency; WS, Water Status index; RAD, sunlight radiation.

2015		2016		20	15	2016	
Yield _p	0.59***	Yield _p	0.64***	EV	0.26**	EV	0.30***
н	0.07 ^{NS}	HI	0.36***	AGA	0.42***	AGA	0.23**
Biomass	0.56***	Biomass	0.48***	SG	0.25**	SG	0.25**
Biomass (<i>Z3.4</i>)	0.45***	Biomass (<i>Z3.7</i>)	0.44***	LUE (<i>Z3.4</i>)	0.47***	LUE (<i>Z3.7</i>)	0.44***
Biomass (<i>Z4.5</i>)	0.53***	-		LUE (<i>Z4.5</i>)	0.42***		-
-		Biomass (<i>25.7</i>)	0.47***		-	LUE (<i>Z5.7</i>)	0.44***
Biomass (<i>Z6.5</i>)	0.54***	Biomass (<i>Z6.5</i>)	0.48***	LUE (<i>Z6.5</i>)	0.33***	LUE (<i>Z6.5</i>)	0.37***
-		Biomass (27.1)	0.50***		-	LUE (<i>Z7.1</i>)	0.37***
-		Biomass (<i>27.5</i>)	0.50***		-	LUE (<i>27.5</i>)	0.22*
-		Biomass (<i>27.8</i>)	0.50***		-	LUE (<i>Z7.8</i>)	0.17 ^{NS}
-		Biomass (<i>27.9</i>)	0.49***		-	LUE (<i>Z7.9</i>)	0.19*
-		Biomass (<i>Z9.4</i>)	0.48***		-	LUE (<i>Z9.4</i>)	0.01 ^{NS}
-		Biomass (<i>Z9.7</i>)	0.48***		-	LUE (<i>Z9.7</i>)	-0.01 ^{NS}
NDVI (<i>Z3.4</i>)	0.48***	NDVI (<i>Z3.7</i>)	0.35***	WI (<i>Z3.4</i>)	-0.38***	WI (<i>Z3.7</i>)	-0.31***
NDVI (<i>Z4.5</i>)	0.28**	-		WI (Z4.5)	-0.22*		-
-		NDVI (<i>25.7</i>)	0.37***		-	WI (<i>25.7</i>)	-0.22*
NDVI (<i>Z6.5</i>)	0.18*	NDVI (<i>Z6.5</i>)	0.33***	WI (<i>Z6.5</i>)	-0.42***	WI (<i>Z6.5</i>)	-0.38***
-		NDVI (<i>27.1</i>)	0.40***		-	WI (<i>27.1</i>)	-0.49***
-		NDVI (<i>27.5</i>)	0.48***		-	WI (<i>27.5</i>)	-0.55***
-		NDVI (<i>27.8</i>)	0.41***		-	WI (<i>27.8</i>)	-0.55***
-		NDVI (<i>27.9</i>)	0.41***		-	WI (<i>27.9</i>)	-0.59***
-		NDVI (<i>29.4</i>)	0.19*		-	WI (<i>Z9.4</i>)	-0.32***
-		NDVI (<i>29.7</i>)	-0.08 ^{NS}		-	WI (<i>Z9.7</i>)	-0.17 ^{NS}

Table 2. Pearson coefficients (r) for correlation with grain yield for measured and predicted traits (based on the Yield_P Model) in a wheat population grown in the UK in 2015 and 2016

Significance levels for correlations are given by an F-test on 1 and 120 degrees of freedom: * p < 0.05; ** p < 0.01, *** p < 0.001, NS not significant. Yield_p, yield predicted by the proposed Yield_p Model; HI, harvest index; Biomass, accumulated biomass at the end of the cycle predicted by the Yield_p Model. EV, early vigour; AGA, accumulated green area; SG, stay green. Biomass (Z), accumulated biomass at the Z stage predicted by the proposed Yield_p Model. NDVI (Z), Normalized Difference Vegetation Index at the Z stage; LUE (Z), Light Use Efficiency at the Z stage; WI (Z), Water Index at the Z stage. Z is the Zadoks scale value for the plant development stage. Traits in **red** were predicted by the model, in **blue** calculated and in **black**, measured.

Table 3. Evaluation indices for Reduced Models from Yield_p Model

Reduced Models	Model description	2015		2016	
Reduced Models	Model description	r	RMSE	r	RMSE
Yield _p Model w/o NDVI _{cor}	Yield _{pR1} = RAD.LUE.WS.HI	0.49	2.49	0.57	4.89
Yield _p Model w/o LUE	Yield _{pR2} = RAD.NDVI _{cor} .WS.HI	0.50	0.50	0.62	1.77
Yield _p Model w/o HI	Yield _{pR3} = RAD.NDVI _{cor} .LUE.WS	0.55	7.62	0.48	10.97
Yield _p Model	Yield _p = RAD.NDVI _{cor} .LUE.WS.HI	0.59	0.99	0.64	1.75

Significance levels for correlations are given by an F-test on 1 and 120 degrees of freedom: p < 0.001. r, Pearson correlation coefficient between predicted and measured yield for each model; RMSE, Root Mean Square Error for each proposed model. Yield_{pR#}, yield predicted by the proposed Reduced Model (from 1 to 3); HI, harvest index; NDVI_{cor}, corrected Normalized Difference Vegetation Index; LUE, Light Use Efficiency; WS, Water Status index; RAD, sunlight radiation.



Figure 1. Graphic representation of leaf cover indices calculated from field level pictures. Leaf cover is given in m² of leaf area per m² of soil. Early vigour, as accumulated leaf area from Z2.4 to the date when 90% of plot area cover by leaves was reached; Accumulated green area, as accumulated leaf area from Z2.4 to Z6.5; Stay green, as accumulated leaf area from Z6.5 to S10. Z, Zadoks scale for crop development (Zadoks *et al.*, 1974); 90C, 90% of plot area cover by leaves; S, senescence scale (Pask and Pietragalla, 2012). Adapted from Pennacchi *et al.*, 2018a.



Figure 2. Yield prediction model for a wheat population grown in the UK in 2015 and 2016. Measured Grain Yield represents grain mass per area determined at full maturity and at 85% dry matter; Predicted Grain Yield represents the output of the proposed Yield_p Model (Equation 20) based on the yield formation equation. Pearson Product Moment correlation coefficients (*r*) were calculated to evaluate the correlation between measured and predicted yield at a significance level of 5% (F-tests). Dashed lines represent the 1:1 identity line.



Figure 3. Graphical representation of the wheat yield prediction model components and their progression during the growing season. Vertical dashed lines represent the measurement time points at specific Zadoks (Z) stages. Coloured dashed lines represent the trendline between estimated and measured points (i.e., before Z3.4/3.7 for both seasons and beyond Z6.5 for 2015). Solid lines represent the connection between measurement points for: LUE (Light Use Efficiency in g C MJ⁻¹; **brown**), WS (Water Status index, **blue**), NDVI_{cor} (in %; **light green**), RAD (Radiation in MJ m⁻²; **yellow**) and Predicted biomass (in t C m⁻²; **dark green**).