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Formation of Two-Dimensional Micelles on Graphene: A Multi-Scale Theoretical and Experimental Study

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ABSTRACT: Graphene and related two-dimensional (2D) materials possess outstanding electronic and mechanical properties, chemical stability and high surface area. However, to realize graphene's potential for a range of applications in materials science and nanotechnology there is a need to understand and control the interaction of graphene with tailored high-performance surfactants designed to facilitate the preparation, manipulation and functionalization of new graphene systems. Here we report a combined experimental and theoretical study of the surface structure and dynamics on graphene of pyrene-oligoethylene glycol (OEG) -based surfactants, which have previously been shown to disperse carbon nanotubes in water. Molecular self-assembly of the surfactants on graphitic surfaces is experimentally monitored and optimized using a graphene coated quartz crystal microbalance in ambient and vacuum environments. Real-space nanoscale resolution nanomechanical and topographical mapping of sub-monolayer surfactant coverage, using ultrasonic and atomic force microscopies both in ambient and ultra-high vacuum, reveals complex, multi-length-scale self-assembled structures. Molecular dynamics simulations show that at the nanoscale these structures, on atomically-flat graphitic surfaces, are dependent upon the surfactant OEG chain length and are predicted to display a previously unseen class of 2D self-arranged 'starfish' micelles (2DSMs). Whilst three-dimensional micelles are well known for their widespread uses ranging from microreactors to drug-delivery vehicles, these 2DSMs possess the highly desirable and tunable characteristics of high surface affinity coupled with unimpeded mobility, opening up strategies for processing and functionalizing 2D materials.

Keywords: 2D micelles, surfactants, graphene, scanning probe microscopy, molecular dynamics

Surfactants are essential¹⁻⁴ for the efficient separation, dispersion and functionalisation of graphitic materials⁵ in aqueous solution and provide enabling steps in the production of graphene devices.⁶ Surfactants play an essential role in promoting, *via* ultrasonic or chemically-driven exfoliation techniques,² the creation of large graphene flakes with better defined geometries and reduced defect densities⁷ that are vital for the realization of device architectures in fields such as energy storage and generation,⁸ flexible displays⁹ and sensors.¹⁰ Whilst the behavior of surfactants in aqueous environments, where they self-assemble into supramolecular assemblies such as three-dimensional spherical micelles with hydrophobic moieties shielded by a hydrophilic shell,¹¹⁻¹³ is well understood, their behavior as (sub) monolayers on surfaces, has received limited attention.¹⁴⁻¹⁶ 3D micelles may be transferred to solid-liquid or solid-air interfaces resulting in hemispherical or, it has been proposed, disk-like structures,¹³ depending on the interplay between cohesive and adhesive energies, with height in both cases of the same order as the lateral size. However, for sensing and energy storage applications, separation of graphene sheets by surface-stable surfactant structures with a smaller thickness is highly desirable.^{17,18} We now report a combined experimental and theoretical study of the molecular assembly of a tailored class of surfactants on graphitic surfaces. The resulting nano-microscale structuring is shown to derive from the formation of flat 2D starfish micelles (2DSMs), whose height is much smaller than their lateral surface dimensions. These 2DSMs are not formed by transferring pre-formed 3D micelles onto surfaces, but instead arise from the post-deposition aggregation of surfactant molecules on the surface.

RESULTS AND DISCUSSION: The recently-synthesized family of surfactants¹⁹ (see Figure 1) contain pyrene groups for stable planar anchoring to the graphene surface, connected by oligoethylene glycol (OEG) chains to hydrophilic head groups derived from Newkome dendrons.²⁰ Pyrene is known to bind to graphene with a binding energy of -1.09 eV, and to experience a low energy barrier of ca. 0.01 eV to sliding and rotation parallel to the sheet.²¹⁻²⁴ This results in a desirable combination of i) strong adsorption of the surfactants onto graphitic surfaces, and ii) free lateral movement over them. Surfactants **1-4** were synthesized as described previously.¹⁹

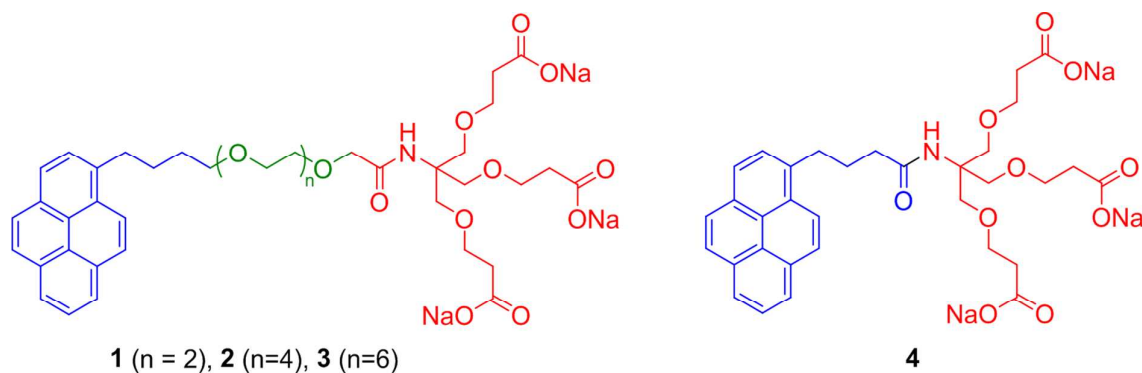


Figure 1. Left: The structures of the homologous series of surfactants 1-3, which vary only in the length n of the oligoethylene glycol (OEG) linker chain. Right: The structure of surfactant 4, an analogue without an OEG linker. The anchoring unit derived from pyrenebutanol (in 1-3) or pyrenebutyric acid (in 4) is indicated in blue, the OEG linker unit (in 1-3) is shown in green, and the hydrophilic head group based on a first generation Newkome dendron is shown in red.

Experimentally, molecular assembly of **2** was monitored by adsorbed molecular layer formation on a quartz crystal microbalance (QCM) sensor surface, on both Au and graphene,²⁵ from aqueous solutions in the concentration range of 0.001 – 0.1 mg/ml. A Sauerbrey analysis²⁶ of the frequency change during assembly was used to determine the area per molecule at frequency plateaus corresponding to stable sub-monolayer, mono- and multi-layer structures. As pyrene is expected to dominate binding between the surfactants and both Au²⁷ and graphene,²⁸ we used the pre-characterized Au coated crystals to benchmark assembly prior to graphene deposition. Molecular concentration and exposure times were modified over several orders of magnitude allowing fine control over the assembly process. Even for the lowest studied concentration of 0.001 mg/ml and a brief 10 s deposition, the QCM measurements reveal the presence of added mass in the molecular system due to both surfactant and substrate bound water. Whilst deposition on Au consistently followed a clear two-stage mechanism (see Figure 2a for example), equivalent conditions resulted in a more stochastic process on graphene (see Figure 2b). This disorder is attributed to the higher surface mobility of the surfactant on graphene²⁴ compared to Au.²⁷

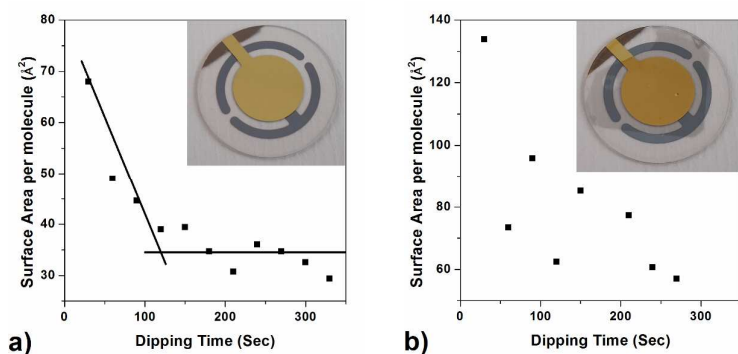


Figure 2. QCM monitored molecular area as a function of the total dipping time of **2 on a 5 MHz QCM with either a (a) gold or (b) graphene surface under ambient condition (inserts - typical Au and graphene coated QCMs).**

We were able to differentiate between water bound directly to QCM surface and bound surfactant-structural water, (*i.e.* that associated with the surfactant layer on the surfaces) by analyzing QCM response at the elevated temperatures and vacuum. The QCM response was first calibrated to eliminate temperature dependence of the crystal frequency itself, detailed in SI 1.3. The application of vacuum led to an increase in frequency (indicating decrease in surface mass) of 7.7 ± 2.2 Hz and 6.5 ± 1.5 Hz for the Au and graphene systems, respectively. This change in frequency is opposite and smaller than the ca. 30 Hz frequency decrease (showing mass increase) due to assembly of **2** on the surfaces and is independent of the number of vacuum cycles applied. Ab-initio calculations^{24,29} suggest that the binding strength of **2** through the pyrene moiety to both Au and graphene is sufficient to ensure that surfactant molecules remain anchored throughout the vacuum and heating cycles. We therefore ascribe the initial vacuum induced mass decrease to removing of loosely bound physisorbed water on the highly hydrophilic surfactant surface.³⁰ Frequency changes arising from 20→60→20 °C heating and cooling cycles under vacuum show a 7 Hz difference for Au and a 4 Hz difference for graphene and, as would be expected without the system being returned to ambient, are again independent of number of heating cycles and indicates all water is removed on the first heat cycle and that no surfactant is removed on subsequent cycles. These changes are assumed to correspond to 'structural' water. Based on mass loss calculations we find the removal of 14 water molecules per **2** molecule for Au surfaces and 17 water molecules per **2** molecule for graphene surfaces. If the extra physisorbed water is taken into account, a total of 28 water molecules per **2** molecule for the gold surface and 40 molecules per **2** molecule for the graphene surface are removed. This corresponds with the number of water molecules calculated within our MD

simulations, discussed in detail below, suggesting excellent agreement between simulation and experiment for these systems.

MOLECULAR IMAGING: Direct nanoscale imaging was performed after assembly of surfactant layers on graphene-like freshly-cleaved highly ordered pyrolytic graphite (HOPG) surfaces. Intermittent contact (tapping mode) atomic force microscopy (AFM) was used in both ambient and UHV conditions and ultrasonic force microscopy (UFM)^{31,32} was used in ambient conditions. Freshly-cleaved HOPG was exposed to 0.001 mg/ml solutions of **1-4** for 10 seconds, rinsed with deionized (DI) water, dried with dry nitrogen and imaged in ambient conditions (Figure 3).

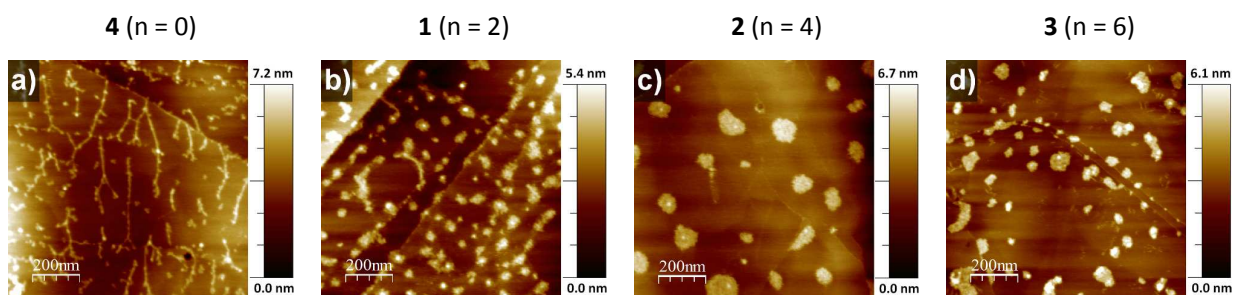


Figure 3. Tapping mode AFM topography images of assembled molecular-structures of surfactants (1-4) on HOPG in ambient conditions where a) the shortest chain surfactant (4) forms elongated chain-like structures. b) For 1 a combination of short chain-like and “island” structures are formed. c) For 2 large, low surface density, islands of the surfactant are formed. d) For 3 similar island features to 2 are observed. In all cases, at these large scan sizes, section analysis shows that the maximum island height is <4 nm which is an order of magnitude less than their typical diameter (see table 1).

The typical ‘island’ structure of these surface features remains unchanged when the coated HOPG is washed with copious amounts of water and rescanned, confirming the high affinity of the surfactants to the graphene-like surfaces. The features did not appear in control images of HOPG exposed to DI water alone. Although the structures formed vary significantly between surfactants, there are several consistent features. Firstly, the height profiles of all ‘islands’ are typically <4 nm, which is small relative to the lateral dimension of the features, and, secondly, there are indications of internal structure within these 2D features. Finally, these island features are of very different length scales to the 3D micelles observed in bulk solution as shown in Table 1 below (see also SI-3).

Surfactant	Typical 2D dimension from AFM (nm)	Typical 3D dimension from DLS (nm)
1	44.65 ± 11.60	384 ± 4
2	54.44 ± 25.50	435 ± 4
3	46.43 ± 18.09	498 ± 3

Table I. Typical diameters of circular features formation by surfactants **1-3** as observed *via* tapping AFM on the HOPG surface and *via* DLS in bulk.

This fine structure within the islands was subsequently investigated using UFM and small area tapping mode topography, shown in Figure 4 d and e, respectively. UFM eliminates the friction due to vertical ultrasonic oscillation of the sample,³³ while preserving the nanoscale nanomechanical contrast to both stiff and soft objects.^{32,34}

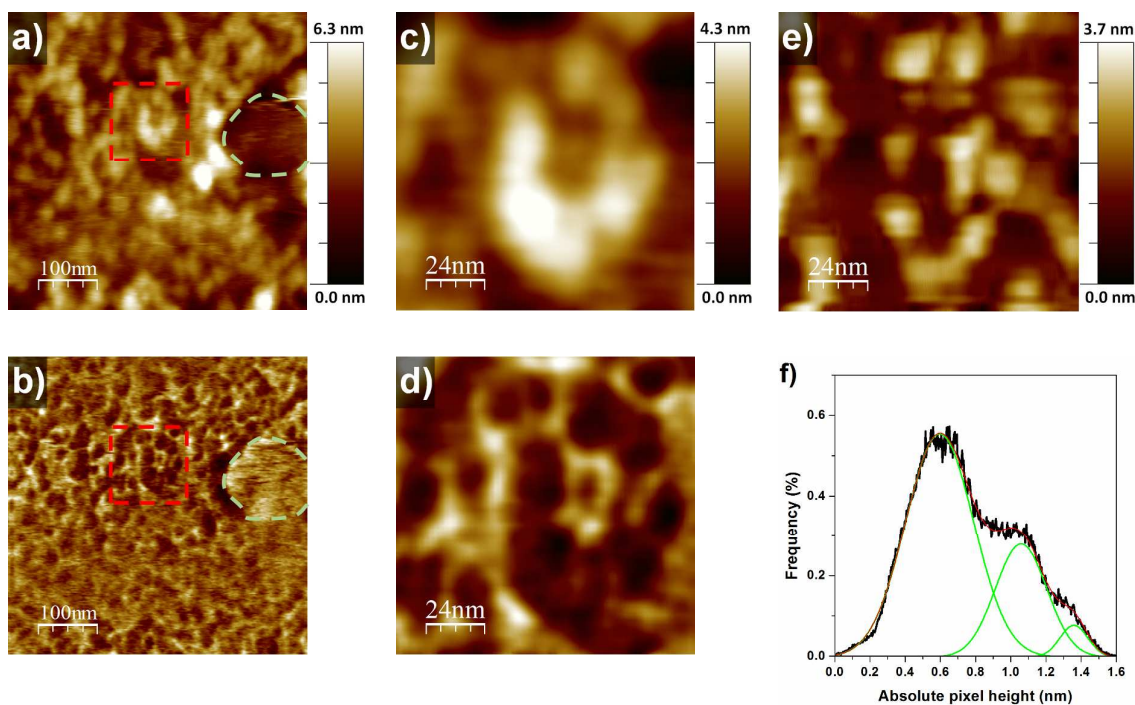
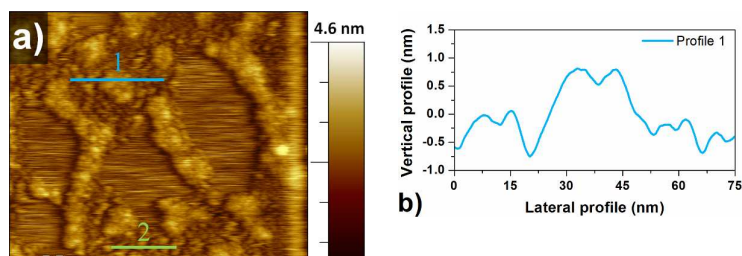


Figure 4. High resolution SPM imaging of surfactant nano-structure in ambient conditions showing a) contact mode AFM topography and in b) the simultaneously acquired UFM nanomechanical response (where darker areas correspond to lower stiffness) of a densely packed film of **2** on HOPG. Topography image c) and corresponding UFM image in d) show zoomed-in details of molecular clusters shown within red dashed

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3 boxes in a) and b) respectively. e) The small scan size tapping mode topography of **2** on HOPG and f) the
4 corresponding image height distribution histogram. Exposed areas of HOPG, indicated with a green dashed
5 line, are clearly visible in both the topography and UFM nanomechanical images.
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10 The UFM scan in Figure 4b shows the compliant (darker contrast) areas corresponding to higher
11 protrusions observed in contact mode topography (Figure 4a) – typical for UFM nanomechanical
12 contrast of small organic systems.³⁵ The area shows a surface region of high molecular coverage –
13 the HOPG substrate can be seen on the right hand edge of the image, indicated by circular green
14 dashes. The small scale (120 x 120 nm) tapping mode scan of Figure 4e shows distinct populations
15 within the surfactant structures; with height distribution analysis (Figure 4f) indicating typical
16 heights of these features at 0.60 ± 0.21 nm (with typical diameters obtained by section analysis of
17 12.79 ± 2.93 nm) and lower frequency higher features in the range 1.1 – 1.4 nm (16.77 ± 1.99 nm
18 diameter), matching well with the UFM nanomechanical maps. Blurring and step-like shifts in the
19 fast scan (horizontal) axis in Figure 4e are due to tip-induced disruption of the film, despite the
20 extremely small lateral forces inherent to tapping mode AFM. This is consistent with the high lateral
21 mobility of the surfactants on the substrate which also precluded clear imaging with scanning
22 tunneling microscopy (STM) even in UHV at 77 K. Some elongation in the slow (vertical) scan axis is
23 believed to be due to thermal drift arising from the slow scan speeds necessary for tapping mode
24 imaging of soft objects. Comparable imaging of **2** was performed in the absence of surface water
25 (Figure 5) by imaging in UHV at 77 K. Sample preparation for UHV measurements followed exactly
26 the same protocol as the ambient samples, with only the addition of a further annealing step under
27 vacuum (50 °C for ~11 hours) before being introduced into the main scan chamber. As with ambient
28 scanning conditions, multi-length scale structures were observed with smallest feature sizes
29 corresponding to lateral dimensions of ~10 nm (Figure 5, line profile 2) and a height of 1-2 nm. In the
30 UHV measurements a second layer structure was also observed confirming a layer height of ~ 1 nm
31 between the first and second molecular layers (Figure 5, line profile 1). This removes any possibility
32 that the molecular layer appears with a lower height due to possible compression by the AFM tip
33 compared to the practically incompressible HOPG surface.
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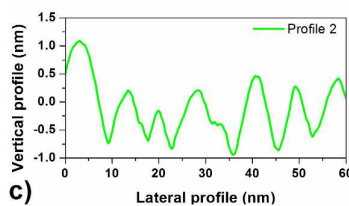


Figure 5. High resolution frequency modulation FM-AFM imaging under UHV conditions with a qPlus sensor. (a) Constant Δf feedback image showing mono and bilayer molecular structures. Scan size: 250 x 250 nm, Δf set point = +10Hz, $\alpha_0 = 225$ pm, $f_0 = 23.7$ kHz. Line profiles show (b) a molecular height profile varying from ~ 1 -2 nm and (c) smallest lateral feature sizes of ~ 10 nm.

2DSM SIMULATIONS: To understand the nature of the observed structures, in both ambient and UHV conditions, we performed molecular dynamic (MD) simulations (see S4 1-3: for details), initially investigating the assembly of the surfactants on a disordered graphene sheet both in the absence and presence of water. In both cases the non-bonded Na^+ counterions of the hydrophilic head group were free to escape and re-coordinate with the three carboxylate moieties during the simulation; however, in the absence of water, none of were observed to do so. Upon introduction of water the Na^+ counterions were able to escape from the vicinity of the head group (see SI) Figure 6 shows the resultant structures for surfactants **1**, **2** and **3**. The first six snapshots (Figure 6 a-c) show that for all three surfactants, in the absence of water, structures form in which the hydrophilic charge-neutral Newkome Dendron head groups aggregate *via* van der Waals forces in the interior with the hydrophobic pyrene anchors on the exterior. However, upon the introduction of water to the simulation, as shown in panels g-i of Figure 6, the hydrophobic pyrene anchors flip from the exterior to the interior of the structures, where they aggregate by van der Waals forces and are surrounded by the OEG linkers and head groups. In this case, the Newkome dendrons can extend into the water layers above the graphene. This is a 2D analogue of the well-known spherical micelle. These 2D micelles possess a 'starfish-like' structure (2DSM) with thicknesses two orders of magnitude smaller than their typical lateral diameter corresponding with the dimensions of the features observed by SPM imaging. In the MD simulations at low coverage of surfactant **1** in water (see S4: Cluster Sizes) the surfactant has a lateral diameter of 43.8 Å with an extension from the graphene surface of 8 Å. The lateral diameter increases with the surfactant concentration and levels out at 56.4 Å at the maximum surface coverage predicted to occur when $m=15$. At higher concentrations the surfactants cluster to form conical structures. See S4:4 Cluster Sizes for details. Starfish 2DSMs are distinct from

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3 conventional micelles or lipid membranes³⁶ as surface interactions and topological steric hindrance,
4 determined by the length of the OEG bridges separating the hydrophobic and hydrophilic moieties,
5 play a major role in determining their dynamics and shapes, as do their local environment. One such
6 environmental effect can be seen in the snapshot shown in Figure 6 panel g for surfactant **1** (chain
7 length $n=2$ and density $m=12$ molecules), which shows examples of Na^+ ions (colored blue) both
8 coordinated to the carboxylate moieties and dissociated into the water. As a cross-check to the
9 robustness of the prediction of 2-D micelle formation the graphene sheet was increased fourfold so
10 that the simulation area is increased to 170.32 nm^2 . This avoids periodic effects as the lower
11 concentrations now do not interact between unit cells. There was no change to the quantitative
12 observation of 2-D micelle formation (see SI S4.2).
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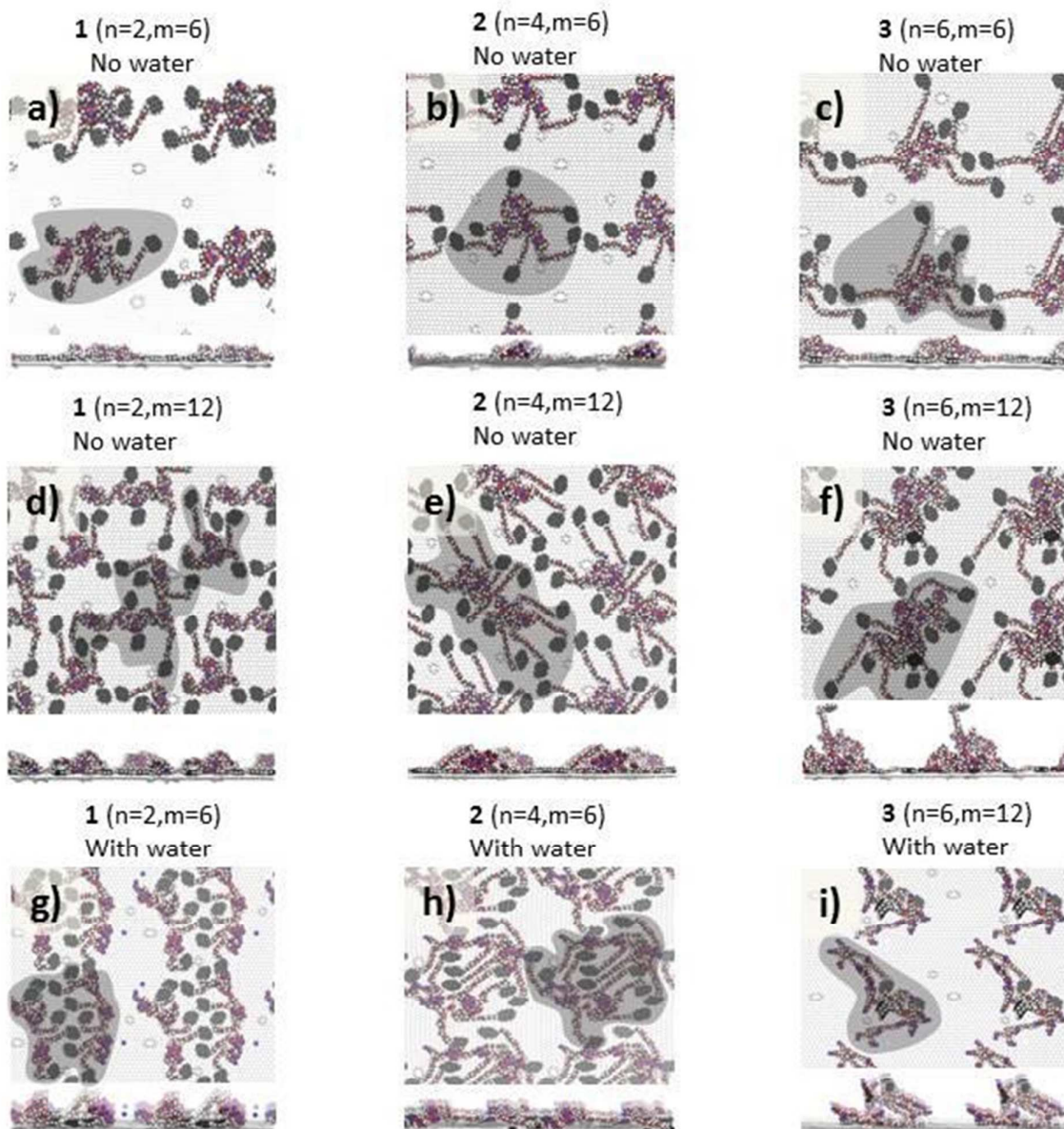
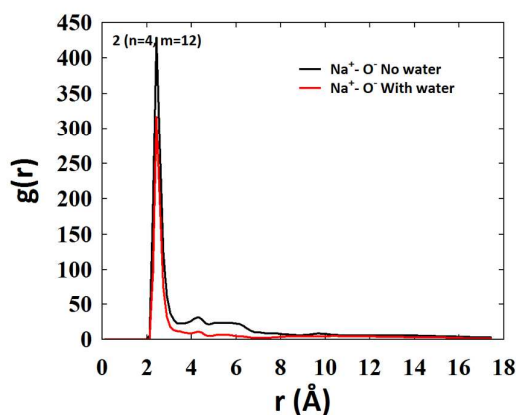


Figure 6. The MD simulations of 2DSM assembly formed by surfactants 1, 2, and 3 (with chain lengths $n = 2$, 4 and 6 respectively and containing $m = 6$ or $m = 12$ surfactant molecules) on a 42.58 nm^2 defective graphene surface at 300 K and 1 atmosphere. Panels a-f show snapshots of the 2DSM assembly in the absence of water and panels g-i show snapshots in the presence of water. The snapshots show four periodic duplicate cells of the graphene sheet with the water not shown for clarity. The integer m denotes the number of surfactant molecules / 42.58 nm^2 sheet. The sheet carbon atoms are shown in light grey with other carbons in dark grey/black, oxygen in red, hydrogen in white, nitrogen in purple and sodium in blue.

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3 The details of the distribution of the Na^+ ions are revealed by the pair correlation function for the
4 separation of Na^+ and the O^- ions in the carboxylate (COO^-) moieties. Figure 7 for surfactant **2**
5 compares the pair correlation function in the absence and presence of water. Clearly in the absence
6 of water (shown by the black curve) the Na^+ ions are coordinated to the O^- ion in the (COO^-) moiety
7 at a separation of 2.5 Å and also associated with the neighboring carboxylate moieties at a
8 separation of ~ 5.0 Å. In the presence of water (shown by the red curve) the Na^+ ions spend much
9 less time coordinated with an O^- ion and are more randomly distributed over the simulation cell. This
10 result is also demonstrated in Figure S10 for surfactant **1**. The anchor units remain strongly adsorbed
11 onto the graphene surface; however, in the presence of water, as the Na^+ ions leave the initially
12 charge-neutral head groups, the anionic dendrons repel each other and are attracted into the lower
13 layers of water by van der Waals forces. This effect can be seen by comparing Figure S9 panel 3
14 where at a high concentration of surfactant **2** in the absence of water the molecules spread out over
15 the surface but in the presence of water in panel 6 conical structures form.



41 In Figure 7 the pair correlation function over the production run of the simulation for the surfactant **2** is
42 shown. In the absence of water (shown by the black curve) the Na^+ ions are coordinated to the O^- ion in the
43 (COO^-) moiety at a separation of 2.5 Å and also associated with the neighboring carboxylate moieties at a
44 separation of ~ 5.0 Å. In the presence of water (shown by the red curve) the Na^+ ions spend much less time
45 coordinated with an O^- ion and are more randomly distributed over the simulation cell.

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49 The structures of the 2DSMs depend not only upon the length n of the OEG linker, but also the
50 coverage, which we quantify by the number m of molecules on the 42.58 nm^2 sheet. For example as
51 m increases from 6 to 12 surfactant **1** tends to form linear structures, whereas surfactant **2** forms a
52 12 legged 2DSM and surfactant **3** starts to develop a conical structure as some of the head groups
53 and the pyrene anchor units extend away from the graphene surface. Increasing m to 24 for
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3 surfactant **2** (shown in Figure S9 panels 1-3) results in 12 of the pyrene anchors forming a base which
4 further molecules stack on top of. Thus the graphene surface remains only partially covered and
5 complete surface coverage is not observed. The trends observed for the surfactants in the absence
6 of water in Figure 6 are very similar to those observed in the presence of water. Surfactant **1** tends
7 to form 2D linear micelles but in this case with the pyrene units sandwiched between the hydrophilic
8 units. Surfactant **2** forms a more circular 12 legged 2DSM as the hydrophilic groups surround the
9 pyrene units. Surfactant **3** again starts to develop a conical structure as some of the head groups and
10 pyrene anchor units extend away from the graphene surface. Figure S9 panels 4-6 for surfactant **2**
11 with $m=24$ in the presence of water also shows that 12 of the pyrene anchors form a base with
12 further molecules stacking on top. This again results in incomplete surface coverage and as the cone
13 extends into the water layers we can estimate the number of water molecules associated with the
14 surfactant outer layers as approximately 40 per molecule of **2**. This is in excellent agreement with
15 the number measured by QCM, above.
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25 **CONCLUSIONS:** The assembly and behavior of a series of tailored surfactant molecules on graphitic
26 surfaces has been investigated in both ambient and high vacuum conditions. In both cases multi-
27 length scale surfactant features are observed with the smallest observable feature possessing a near
28 2D profile. Assembly of such 2D structures is supported by MD simulations; furthermore MD
29 calculated the same ratio of surfactant to water molecules on the graphene surface as direct QCM
30 quantitative molecular adsorption experiments. MD confirms that these 2D structures behave as
31 planar micelles on the surface, with a structure dependent on the presence or absence of water. MD
32 also shows that the strongly surface-attached pyrene anchor units are located at the periphery of
33 the structures in the absence of water, but at the core in the presence of water. The existence and
34 manipulation of 2DSMs on graphene will be important for the control of surface functionalisation
35 and a range of associated technologies. The realization of 2DSM opens routes to developing 2D
36 microreactors which could lead to important advances, including control of the size and shape of 2D
37 nano-islands grown on surfaces, with applications to nano-scale circuitry and biosensing. The
38 chemical tunability of surfactants could enable the engineering of tailored structural molecular
39 features in multi-functional graphene-based materials. Furthermore the ability to use scanning
40 probe technologies to image 2DSMs in solution is likely to have ramifications for our understanding
41 of the fundamental science of micelle formation and control. A mechanism for non-covalent
42 molecular capture can be postulated where a molecule in solution is captured non-covalently by van
43 der Waals forces within the Newkome dendrons and as the surfactant is dried so the molecule is
44 held as the micelle reverses. Unwanted ions in solution can also exchange with the Na^+ ions
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3 associated with the carboxylate moieties and can be removed after drying. If the surfactants can be
4 repeatedly dried and rehydrated the captured molecules or ions could then be repeatedly captured
5 and released. The tendency for surfactant **3** to extend into the water environment can be exploited
6 as a device design mechanism. A layer of the surfactant on graphene forms conical structures and
7 depositing another graphene layer over the cones would result in the two graphene layers being
8 separated by discrete pillars. This could have applications for thermoelectrics as the soft micelle
9 pillars would inhibit phonon transport across the graphene layers.
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14 15 16 17 **METHODS:**

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19 **Quartz crystal microbalance:** Segregation grown graphene³⁷ was transferred onto the top Au
20 electrode of the QCM liquid handling head resulting in films of 1-2 graphene layers thickness, with
21 continuous coverage over the entire QCM electrode area resulting in maximal sensitivity (more
22 details in SI-1).³⁸ QCMs were dip-coated in surfactant solution, in the range 0.001 – 0.1 mg/ml for 10
23 s, prior to washing with DI water and nitrogen drying. The QCM frequency was measured and then
24 the process repeated. No change in frequency was observed for the subsequent DI water
25 washing/dry nitrogen drying stages suggesting these steps did not remove the surfactant. A custom
26 vacuum chamber comprising a high thermal conductivity aluminum base in a direct thermal contact
27 with a large area Peltier heater/cooler was used to ensure rapid thermal equilibrium in the system,
28 further details are given in SI-1.3. A relevant temperature range of 20 – 60 °C was used with a
29 pressure varied alternated between ambient and 10⁻¹ torr. Continuous vacuum pumping ensures
30 that water removed by heating was efficiently evacuated from the chamber. Both Au and graphene
31 QCMs were calibrated for temperature induced frequency changes; the frequency changes solely
32 due to water evaporation as a function of vacuum and temperature on graphene are given in Figure
33 S3.
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43 **Ambient SPM imaging:** Contact mode, tapping mode atomic force microscopy and ultrasonic force
44 microscopy were all performed using a Multimode Nanoscope IV AFM (Bruker AXS). For tapping
45 mode high stiffness $k = 40 \text{ Nm}^{-1}$ and 300 kHz resonance (Tap300-G, Budget Sensors) probes were
46 used. Topography and phase (shown in SI) were captured simultaneously. As UFM is essentially a
47 modification of contact mode imaging albeit with eliminated friction, probes with $k = 0.2 \text{ Nm}^{-1}$
48 (Contact-G, Budget Sensors) were used. A thin plate piezoceramic actuator (4 MHz thickness mode
49 resonance, PI) was bonded to the sample substrate and used to generate ultrasonic vibrations,
50 resulting in efficient ultrasonic excitation at frequencies up to 10 MHz.³² The piezoceramic actuator
51 was excited by a function generator (LXI Keithley) at 4.2 MHz ultrasonic frequency modulated at 2.3
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3 kHz; the cantilever deflection at this modulation frequency corresponds to the nonlinear UFM
4 response, which is highly dependent on the local nanomechanical properties of the sample, and was
5 detected *via* a lock-in amplifier (Stanford Research Systems, SRS-830) as described in detail
6 elsewhere.^{39,40}
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11 **High vacuum SPM imaging:** Frequency modulation atomic force microscopy (FM-AFM)
12 measurements collected under ultra-high vacuum (UHV) were obtained using an Omicron
13 Nanotechnology GmbH LT STM-AFM system operating at liquid nitrogen temperature (77 K) and an
14 Omicron Nanotechnology GmbH VT STM-AFM operating at room temperature. Both systems were
15 kept under ultrahigh vacuum conditions with a base pressure of 5×10^{-11} mbar or better. FM-AFM
16 measurements at LT were collected using a commercial qPlus sensor⁴¹ supplied by Omicron GmbH
17 with a resonant frequency of ~ 23.7 kHz and a stiffness of ~ 2000 Nm⁻¹. For all FM-AFM
18 measurements images were acquired under feedback by maintaining a constant Δf set point. Prior to
19 all measurements on the micelle samples, images were collected on clean Ag(111) surfaces where
20 the tip was conditioned (*via* standard voltage pulsing and tip-indentation methods) until atomically
21 sharp steps were observed, thus confirming the presence of an atomically sharp probe. We have
22 therefore neglected any contribution of tip dilation from quoted dimensions for these
23 measurements. Unusually, imaging in the conventional attractive mode (*i.e.* maintaining a negative
24 value of Δf) was found to be exceptionally challenging and snap-to-contact was often observed. We
25 therefore instead performed the majority of measurements using positive Δf set points where
26 images were found to be more stable.
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39 **Molecular modelling:** Molecular dynamic (MD) simulations were performed to investigate the
40 assembly of the surfactants on the surface of a disordered graphene sheet in water. The simulations
41 were computed by using the MD package DLPOLY_4.⁴² The system was modelled as a periodic
42 cuboid of water (parameterized to the TIP3P water model at 300 K, 1 atmosphere and a density of
43 1000kg/m^3) of side $60 \times 60 \times 75$ Å which contained a number, m , of surfactant units placed over a
44 periodic, disordered sheet of graphene of side 60×75 Å. Each calculation was relaxed for 8 ns prior
45 to obtaining the snapshots shown in Figure 6 of the main text using the Nosé-Hoover (npt)
46 thermostat with barostat ensemble. An adapted force field was constructed using the Dreiding force
47 field scheme with the parameters found in Mayo *et al.*⁴³ and charges allocated to each atomic
48 species calculated by the DFT package SIESTA⁴⁴ using the van der Waals density functional⁴⁵⁻⁴⁷ where
49 extended double zeta polarized basis sets of the pseudo atomic orbitals were used to optimize the
50 geometries by relaxing the atomic forces to less than 20 meV/Å. Importantly, the force field charges
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3 were carefully tuned to allow the Na⁺ ions coordinated to the three carboxylate moieties in the head
4 group shown in Figure 1, to disperse into the water environment and recombine during the
5 simulation. The overall system charge was neutral.
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22 **Supporting Information Available:** Details of quartz crystal microbalance measurements, additional
23 SPM imaging, details of dynamic light scattering studies of 3D micelle structures and extensive
24 details of molecular modelling. This material is available free of charge *via* the Internet at
25 <http://pubs.acs.org>.
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