

# Accepted Manuscript

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PII: S0378-5955(16)30354-9

DOI: [10.1016/j.heares.2016.12.002](https://doi.org/10.1016/j.heares.2016.12.002)

Reference: HEARES 7284

To appear in: *Hearing Research*

Received Date: 13 August 2016

Revised Date: 6 December 2016

Accepted Date: 8 December 2016

Please cite this article as: Guest, H., Munro, K.J., Prendergast, G., Howe, S., Plack, C.J., Tinnitus with a normal audiogram: Relation to noise exposure but no evidence for cochlear synaptopathy, *Hearing Research* (2017), doi: 10.1016/j.heares.2016.12.002.

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# **Tinnitus with a normal audiogram: Relation to noise exposure but no evidence for cochlear synaptopathy**

Hannah Guest<sup>a,\*</sup>, Kevin J. Munro<sup>a,c</sup>, Garreth Prendergast<sup>a</sup>, Simon Howe<sup>b</sup>, Christopher J. Plack<sup>a,d</sup>

<sup>a</sup> Manchester Centre for Audiology and Deafness, University of Manchester, Manchester Academic Health Science Centre, UK

<sup>b</sup> Audiology Department, James Cook University Hospital, South Tees Hospitals NHS Foundation Trust, Middlesbrough, UK

<sup>c</sup> Central Manchester University Hospitals NHS Foundation Trust, Manchester, UK

<sup>d</sup> Department of Psychology, Lancaster University, Lancaster, UK

\* Corresponding author. Manchester Centre for Audiology and Deafness, HCDH Office, Ellen Wilkinson Building, University of Manchester, Oxford Road, Manchester M13 9PL, UK. Tel: 0161 275 8568. Email address: [hannah.guest@manchester.ac.uk](mailto:hannah.guest@manchester.ac.uk)

## 1 Abstract

2 In rodents, exposure to high-level noise can destroy synapses between inner hair cells and auditory  
3 nerve fibers, without causing hair cell loss or permanent threshold elevation. Such “cochlear  
4 synaptopathy” is associated with amplitude reductions in wave I of the auditory brainstem response  
5 (ABR) at moderate-to-high sound levels. Similar ABR results have been reported in humans with  
6 tinnitus and normal audiometric thresholds, leading to the suggestion that tinnitus in these cases  
7 might be a consequence of synaptopathy. However, the ABR is an indirect measure of  
8 synaptopathy and it is unclear whether the results in humans reflect the same mechanisms  
9 demonstrated in rodents. Measures of noise exposure were not obtained in the human studies, and  
10 high frequency audiometric loss may have impacted ABR amplitudes. To clarify the role of cochlear  
11 synaptopathy in tinnitus with a normal audiogram, we recorded ABRs, envelope following responses  
12 (EFRs), and noise exposure histories in young adults with tinnitus and matched controls. Tinnitus  
13 was associated with significantly greater lifetime noise exposure, despite close matching for age,  
14 sex, and audiometric thresholds up to 14 kHz. However, tinnitus was not associated with reduced  
15 ABR wave I amplitude, nor with significant effects on EFR measures of synaptopathy. These  
16 electrophysiological measures were also uncorrelated with lifetime noise exposure, providing no  
17 evidence of noise-induced synaptopathy in this cohort, despite a wide range of exposures. In young  
18 adults with normal audiograms, tinnitus may be related not to cochlear synaptopathy but to other  
19 effects of noise exposure.

20 **Keywords:** Tinnitus; Cochlear synaptopathy; Hidden hearing loss; Auditory brainstem response;  
21 Envelope following response; Noise-induced hearing loss

22 **Abbreviations:** ABR, auditory brainstem response; AN, auditory nerve; EFR, envelope  
23 following response; SR, spontaneous rate; TNA, tinnitus with a normal audiogram.

## 24 1. Introduction

25 Subjective tinnitus – the perception of sound without an acoustic source – is most often associated  
26 with hearing loss (Nicolas-Puel et al., 2002; Sanchez et al., 2005). It is widely agreed that these  
27 phenomena are related, with hearing loss usually regarded as a trigger for neuroplastic changes in  
28 the central auditory system, giving rise to the tinnitus percept. While these central changes differ in  
29 the various prevailing neural models of tinnitus, they are generally thought to be provoked by loss of  
30 input from the auditory nerve (AN) to central auditory structures (Henry et al., 2014; Schaette,  
31 2014).

32 Seemingly at odds with this widespread account of tinnitus generation, approximately 8% of tinnitus  
33 patients have pure tone audiometric thresholds within the normal range (Barnea et al., 1990;  
34 Sanchez et al., 2005). The prevalence of tinnitus with a normal audiogram (TNA) might be taken to  
35 indicate that cochlear damage is not a routine requirement of tinnitus generation. However, recent  
36 findings in a variety of rodent models have suggested otherwise, by demonstrating that substantial  
37 damage to the auditory periphery can occur without affecting cochlear thresholds. Seminal research  
38 in mice by Kujawa and Liberman (2009) revealed that carefully titrated noise exposure can lead to  
39 immediate and extensive loss of synapses between cochlear inner hair cells and AN fibers, yet  
40 leave inner and outer hair cells macroscopically intact. Termed “cochlear synaptopathy”, this  
41 primary deafferentation has also been observed in noise-exposed guinea pigs (Lin et al., 2011) and  
42 in aging mice without significant noise exposure (Sergeyenko et al., 2013). Crucially, the pathology  
43 does not compromise sensitivity to low-level sounds, seemingly due to preferential loss of AN fibers  
44 with low spontaneous firing rates (SRs) and high thresholds (Furman et al., 2013). Consistent with  
45 low-SR fiber loss, abnormal auditory processing is evident at higher sound levels. Synaptopathic  
46 ears exhibit permanent reductions in the amplitude of wave I of the auditory brainstem response  
47 (ABR) to tone bursts with moderate-to-high sound levels (Kujawa and Liberman, 2009).

48 Similar electrophysiological evidence of deafferentation has been reported in humans with TNA.  
49 Schaette and McAlpine (2011) recorded ABRs to clicks with high sound levels and demonstrated  
50 reductions in wave I amplitude in TNA subjects relative to audiogram-matched controls. The results  
51 were interpreted as evidence of deafferentation consistent with cochlear synaptopathy: a “hidden  
52 hearing loss” which might resolve the enigma of TNA. The absence of any tinnitus-related reduction

53 in ABR wave V was tentatively attributed to increased central gain in the auditory brainstem,  
54 suggested as a mechanism of tinnitus generation. Gu et al. (2012) reported similar findings in  
55 subjects with near-normal hearing.

56 However, the latter study demonstrated significant wave I amplitude reductions only for the highest  
57 stimulus level used, 120 dB peSPL, and not for lower levels more comparable with those of  
58 Schaette and McAlpine ( $\leq 100$  dB peSPL). Missing ABR data at this high stimulus level led to  
59 reduced participant groups with unmatched audiograms at high frequencies (tinnitus had  
60 systematically poorer mean thresholds above 8 kHz). This disparity may have accounted for the  
61 group difference in ABR amplitude, since wave I is dominated by the responses of high frequency  
62 AN fibers (Don and Eggermont, 1978). Schaette and McAlpine's tinnitus and control groups also  
63 differed in high frequency sensitivity. Mean 12 kHz threshold was elevated by  $\sim 3.5$  dB in the  
64 tinnitus group, and thresholds at even higher frequencies were not reported. Additionally, a recent  
65 study by Gilles et al. (2016) found no wave I amplitude reduction in young people with tinnitus,  
66 though statistical power was compromised by high measurement variability. Given the growing  
67 interest in cochlear synaptopathy in humans, the evidence for its role in tinnitus could benefit from  
68 careful confirmation.

69 Investigation of the condition in living humans is necessarily indirect and requires a sensitive, non-  
70 invasive measure. The transient-evoked ABR may offer limited sensitivity to synaptopathy in  
71 humans, despite clear correlations with the pathology in rodent models. ABR amplitudes are highly  
72 variable, influenced by factors such as head size, cochlear dispersion, and skull thickness  
73 (Michalewski, 1980; Trune et al., 1988; Don et al., 1994), which might obscure the effects of  
74 synaptopathy. Differential ABR measures may minimize the influence of these non-synaptopathic  
75 factors (Plack et al., 2016), but recent evidence suggests a more fundamental shortcoming of the  
76 ABR. Recordings in gerbils and guinea pigs after ototoxic exposure indicate that AN fibers with the  
77 lowest SRs do not contribute to the compound action potential, equivalent to ABR wave I (Bourien  
78 et al., 2014). The low-SR fibers affected in animal models of synaptopathy exhibit a somewhat wider  
79 range of firing rates than those described by Bourien and colleagues (Furman et al., 2013).  
80 Nevertheless, the former exhibit relatively weak onset responses (Taberner and Liberman, 2005),  
81 limiting their contribution to the ABR (Shaheen et al., 2015).

82 In contrast, low-SR fibers surpass high-SR fibers in their synchronization to amplitude-modulated  
83 stimuli (Joris et al., 2004). Hence they make robust contributions to the subcortical envelope  
84 following response (EFR): a sustained response representing neural synchrony to the envelope of  
85 an amplitude-modulated stimulus. Relatively high modulation frequencies are necessary to elicit the  
86 subcortical EFR. At lower frequencies, below 80 Hz, responses are dominated by cortical  
87 generators (Kuwada et al., 2002). Using EFR stimuli optimized to enhance the contribution from the  
88 AN, Shaheen et al. (2015) demonstrated that EFR amplitude afforded greater sensitivity to noise-  
89 induced cochlear synaptopathy in mice than ABR amplitude. An additional strategy to enhance the  
90 sensitivity of the EFR was devised by Bharadwaj et al. (2015), who reasoned that stimuli with high  
91 sound levels and shallow modulations should be weakly encoded in synaptopathic ears, due to  
92 saturation of high-SR fibers and consequent reliance on low-SR units. To reduce variability from  
93 non-synaptopathic sources that might affect raw EFR amplitude, the researchers computed the  
94 slope of the function relating EFR amplitude to stimulus modulation depth. This measure was shown  
95 to correlate with behavioral measures of temporal coding and auditory selective attention in  
96 audiometrically normal humans, with synaptopathy proposed as a potential underlying cause.  
97 Hence carefully designed EFR measures may be of value in the identification of cochlear  
98 synaptopathy in humans.

99 Finally, previous studies associating TNA with evidence of cochlear synaptopathy have not obtained  
100 measures of lifetime noise exposure. Indeed, to the authors' knowledge, no previous study has  
101 reported that TNA is associated with elevated noise exposure compared to audiogram-matched  
102 controls. It is therefore unclear whether the reported electrophysiological effects in TNA are caused  
103 by the same mechanisms demonstrated in rodent models of noise-induced synaptopathy.

104 The fourfold aims of the present study were: (a) To determine whether TNA is associated with  
105 greater lifetime noise exposure; (b) To provide a further test of the hypothesis that TNA is  
106 associated with ABR effects consistent with cochlear synaptopathy, controlling for high frequency  
107 sensitivity; (c) To determine whether TNA is associated with temporal coding deficits consistent with  
108 synaptopathy; (d) To examine the relations between electrophysiological measures of synaptopathy  
109 and lifetime noise exposure.

## 110 **2. Material and methods**

### 111 **2.1 Participants**

112 Control participants were recruited from the University of Manchester staff and student population  
113 (via poster and on-line advertising) and from the general Manchester population (via on-line  
114 advertising). Tinnitus participants were recruited from the same sources, with the addition of  
115 patients identified by local audiology services. All participants were required to exhibit bilaterally  
116 normal pure tone audiometric thresholds ( $\leq 20$  dB HL at 0.25 to 8 kHz) and middle ear function  
117 (compliance 0.3 to 1.6 ml; middle ear pressure -50 to +50 daPa). All were without history of head  
118 trauma, middle ear surgery, neurological disorder, and ototoxic exposure.

119 Tinnitus participants ( $n = 20$ , female = 10) were aged  $25.7 \pm 1.3$  years (mean  $\pm$  standard error of the  
120 mean). All reported prolonged spontaneous tinnitus that was stable ( $> 4$  months) and non-pulsatile.  
121 Tinnitus characteristics are summarized in Table 1. The mean Tinnitus Functional Index (TFI) score  
122 was  $33 (\pm 7)$ , which corresponds to “moderate” problems with tinnitus on average (Henry et al.,  
123 2016).

124 Control participants ( $n = 20$ , female = 10, mean age =  $25.5 \pm 1.3$  years) were individually matched  
125 with tinnitus participants on the basis of age (to within 18 months) and sex. Mean audiometric  
126 thresholds were matched between groups to within 2.3 dB at all test frequencies from 0.25 to 14  
127 kHz, after averaging the left and right ear thresholds. At the extended high frequencies (10 and 14  
128 kHz), the group means differed by  $< 1$  dB (Fig. 1).

129 Sample size was selected to provide 80% power ( $\alpha = 0.05$ , one-tailed) to detect the ABR effect size  
130 demonstrated by Schaette and McAlpine (2011) for a 100 dB peSPL stimulus. It should be noted  
131 that the previous study recruited only female participants, whereas the present study recruited a  
132 mixed sex sample, potentially inflating ABR amplitude variability. However, variability from other  
133 sources was expected to be reduced (e.g. by use of active electrodes) and this expectation was  
134 fulfilled (see 3.2 and 4.2 for post-hoc power analysis).

### 135 **2.2 Noise exposure history**

### 136 2.2.1 *General procedure*

137 Each participant provided a detailed history of lifetime noise exposure via structured interview,  
138 based on the procedure described by Lutman et al. (2008). For all exposures estimated to exceed  
139 80 dBA (see 2.2.3), data were gathered on estimated sound level, total duration of exposure, and  
140 use of personal hearing protection. The participant provided information first on occupational noise  
141 exposure, followed by social noise exposure. The duration of the structured interview ranged from 5  
142 to 45 minutes. Example noise exposure data for a single participant are given in Table 3 of  
143 Supplementary Material.

### 144 2.2.2 *Determination of activities incurring noise exposure*

145 The participant was asked to recall activities that routinely involved exposure to sound levels  $\geq 80$   
146 dBA (see 2.2.2). A list of the most common social activities involving noise was provided (given in  
147 Lutman et al., 2008). Each activity identified by the participant was marked as an entry in their noise  
148 record, and associated information sought on duration and sound level. An activity was treated as a  
149 single entry only if it entailed approximately consistent sound levels throughout all exposures. If the  
150 sound level varied, then the exposures were broken down into two or more activities (e.g. “loud  
151 bars” and “quieter bars” or “metal gigs” and “rock gigs”).

### 152 2.2.3 *Estimation of sound level*

153 For free-field exposures, sound levels were estimated based on vocal effort required to hold a  
154 conversation at a distance of 1.2 m. Reported vocal effort was converted to dBA level using a  
155 speech communication table (Lutman et al., 2008; see Table 2 of Supplementary Material). For  
156 example, if the participant recalled that it was necessary merely to “raise one’s voice” to hold a  
157 conversation (rather than “talk very loudly” or “shout”), an estimated level of 87 dBA was selected.  
158 Information was also provided on use of personal hearing protection: type, attenuation (if known),  
159 and proportion of time worn during each activity. When attenuation was unknown, it was estimated  
160 from type of protector (see Lutman et al., 2008).

161 For exposures incurred through use of personal music players, the participant reported the typical  
162 setting of the volume control on their device, expressed as a percentage of the maximum setting.



163 This value was converted to a free-field equivalent output level, based on the output levels  
 164 measured by Portnuff et al. (2011) across a variety of devices coupled to stock earphones (see  
 165 Table 2 of Supplementary Material).

### 166 2.2.3 Estimation of exposure duration

167 For a given activity, the participant identified a time period (usually a number of years) during which  
 168 they had engaged in the activity with approximately uniform regularity. The participant then  
 169 estimated the number of hours per day, days per week, and weeks per year of exposure during that  
 170 period, allowing calculation of total hours of exposure. Often, the participant would report having  
 171 engaged in an activity more frequently during one period than another. Hours of exposure would be  
 172 calculated for each period separately, then summed. Additionally, where hearing protection had  
 173 been worn only part of the time, it was necessary to calculate the protected and unprotected  
 174 exposure durations.

### 175 2.2.4 Calculation of units of noise exposure

176 For each activity in the noise record, duration, level, and protector attenuation were combined to  
 177 generate units of noise exposure based on the equal energy principle:

$$178 \quad U = 10^{(L-A-90)/10} \times T / 2080$$

179 where:  $U$  = units of noise exposure

180  $L$  = level (dBA)

181  $A$  = attenuation of ear protection (dBA)

182  $T$  = total exposure time (hours)

183 The units from all exposures, regardless of whether they occurred in social or occupational settings,  
 184 were summed to yield the total units of lifetime noise exposure. The resulting measure is linearly  
 185 related to the total energy of exposure above 80 dBA.

## 186 2.3 Behavioral testing

187 Participants were seated in a double-walled sound-attenuating booth, providing responses using a  
188 button (pure tone audiometry) or mouse and computer monitor (high frequency audiometry). Air  
189 conduction pure tone audiometric thresholds were obtained in accordance with British Society of  
190 Audiology recommended procedures (British Society of Audiology, 2011) at 0.25, 0.5, 1, 2, 3, 4, 6,  
191 and 8 kHz, using a GSI Arrow audiometer, TDH-39 supra-aural headphones, and MX-41 ear  
192 cushions. High frequency thresholds were obtained using a three-interval, three-alternative, forced-  
193 choice paradigm, with stimuli delivered through Sennheiser HDA 200 circum-aural headphones  
194 driven by an E-MU 0202 external audio interface. In order to minimize the influence of threshold  
195 microstructure and ear canal resonance, stimuli were 1/3-octave bands of noise centered at 10 and  
196 14 kHz. Steady state duration was 180 ms, with the addition of 10 ms raised-cosine onset and offset  
197 ramps. Stimulus level was varied adaptively using a two-down, one-up rule. Threshold was attained  
198 using three initial turnpoints (6 dB step size) and eight subsequent turnpoints (2 dB step size). The  
199 stimulus level at the final eight turnpoints was averaged to obtain threshold. Thresholds were  
200 obtained for each ear separately and then averaged across ears. Prior to testing, each participant  
201 performed a practice run containing at least three turnpoints.

## 202 **2.4 Auditory evoked potentials**

### 203 *2.4.1 General procedure*

204 Participants reclined comfortably with eyes closed in a double-walled sound-attenuating booth.  
205 Auditory stimuli were delivered through EARtone 3A insert earphones with mu-metal and aluminum  
206 shielding, driven by an Avid FastTrack C400 external audio interface (48 kHz output). Evoked  
207 responses were recorded using the BioSemi ActiveTwo measurement system, with active  
208 electrodes at Cz, C7, and both mastoids. Common Mode Sense and Driven Right Leg electrodes  
209 were located on the low forehead and electrode offsets were maintained within  $\pm 40$  mV throughout  
210 each recording. Bioelectric activity from each electrode was digitized at a sampling rate of 16384 Hz  
211 and processed off-line in MATLAB (The Mathworks, 2013). EEG data files incorporated stimulus  
212 timing information by means of a custom trigger box connecting the external audio interface to the  
213 BioSemi USB interface.

### 214 *2.4.2 Auditory brainstem response*

215 Digital stimuli were single-polarity high-pass filtered clicks (first-order butterworth, 2.4 kHz cutoff).  
216 Due to the low-pass response of the ER3A inserts, the stimuli in the ear canal had a 10 dB  
217 bandwidth extending from about 1.2 to 4.7 kHz (measured in a Gras IEC60711 occluded ear  
218 simulator coupled to ER3A insert earphones). In order to minimize recording time, presentation  
219 alternated between ears, at a rate of 7.05 per second in each ear, so that a click in one ear was  
220 followed after approximately 71 ms by a click in the other ear. This gave an overall presentation rate  
221 of 14.1 per second and a total of 7040 presentations per ear. The inter-stimulus interval was jittered  
222 by a maximum of 10%, so as to prevent accumulation of stationary interference. In order to  
223 stimulate low-SR fibers, a presentation level of 102 dB peSPL (peak-to-peak) was selected, 2 dB  
224 higher than the maximum level used by Schaette and McAlpine (2011).

225 Activity between Cz and ipsilateral mastoid was filtered (30-1500 Hz; fourth-order butterworth) and  
226 divided into epochs extending from 10 ms pre-stimulus to 13 ms post-stimulus, after correcting for  
227 the 0.8 ms acoustic delay introduced by the sound tube. Post-hoc artifact rejection eliminated  
228 epochs whose RMS amplitude exceeded the mean by more than two standard deviations. The  
229 remaining epochs were averaged and corrected for any linear drift by subtracting a linear fit to the  
230 pre-stimulus baseline.

231 Waves I and V of the ABR were identified and quantified automatically in MATLAB (The Mathworks,  
232 2013), based on waveform characteristics within specified time windows. The window for wave I  
233 extended from 1.55 to 2.05 ms after stimulus peak and the window for wave V from 5.1 to 6.5 ms.  
234 The trough of wave I was required to occur 0.3 to 1.0 ms after its peak. The peak and trough of  
235 wave I were defined as local maxima and minima. Wave V required more subtle denotation, in order  
236 to appropriately interpret waveforms featuring a prominent wave IV or blended wave IV/wave V  
237 complex. Hence the peak of wave V was defined as either a local maximum or a downward  
238 inflection point on a falling portion of the waveform (a maximum in the first derivative where the first  
239 derivative  $< 0$ ). Wave I amplitude was measured from peak to following trough. Wave V was  
240 measured from peak to baseline, in order to capture the gradual rise in amplitude from pre-stimulus  
241 baseline to wave V peak observed in all waveforms (presented in Supplementary Material). Post-  
242 hoc subjective review verified that all waveforms had been appropriately interpreted by the peak-  
243 picking algorithm. The resulting amplitudes and latencies were averaged across left and right ears  
244 for each participant.

### 245 2.4.3 Envelope following response

246 Subcortical EFRs were recorded using the variable-modulation-depth paradigm described by  
247 Bharadwaj et al. (2015). Stimuli were 75 dB SPL transposed tones (Bernstein and Trahiotis, 2002)  
248 with a 4000 Hz carrier and 100 Hz modulator (Fig. 2). The steady-state duration was 400 ms with  
249 the addition of 15 ms onset and offset ramps. Off-frequency contributions were attenuated by  
250 notched-noise maskers (10-20000 Hz overall bandwidth, with a notch width of 800 Hz centered on  
251 4000 Hz) applied at a signal-to-noise ratio (SNR) of 20 dB (broadband RMS). The noise was  
252 realized separately for each of the 180 trials in a block, rather than being frozen between trials.  
253 Stimuli were of two modulation depths (0 dB and -6 dB re: 100% modulation) and each was  
254 presented in two polarities. The resulting four stimuli were presented in the sequence: 0 dB;  
255 inverted 0 dB; -6 dB; and inverted -6 dB. The average inter-stimulus interval was 400 ms, jittered by  
256 up to 10%. This sequence was presented 630 times.

257 Activity in the vertical channel from Cz to C7 was divided into epochs extending from 4 to 404 ms  
258 after the onset of the steady-state portion of the stimulus. Post-hoc artifact rejection eliminated  
259 epochs whose RMS level exceeded the 99<sup>th</sup> percentile for the recording. The remaining epochs  
260 were averaged and the opposing-polarity averages added to give the response to the temporal  
261 envelope. Response spectra were analyzed to yield EFR amplitude at the 100 Hz modulation  
262 frequency, as well a measure equal to the difference in EFR amplitude (in dB) at the two stimulus  
263 modulation depths (Fig. 2). The EFR difference measure is closely related to that of Bharadwaj et al.  
264 (2015) - the slope of the function relating EFR amplitude to modulation depth - though slope was  
265 defined by a three-point function in the previous study. Unlike the other electrophysiological  
266 measures, the EFR difference measure was expected to *increase* due to synaptopathy, since ears  
267 with depleted low-SR fibers should exhibit particularly weak encoding of shallow modulations. In  
268 order to compute the difference measure for a given participant, significant 100 Hz EFR peaks were  
269 required in response to both modulation depths (defined as > 3 dB SNR, with noise being estimated  
270 from the mean amplitude in 10 adjacent frequency bins).

## 271 2.4 Statistical analysis

272 Statistical analysis was performed using R (R Core Team, 2015). All significance tests were  
273 conducted two-tailed. Data were checked for normality and homogeneity of variance prior to testing,  
274 and non-parametric tests applied where necessary. No data points were missing for any variable,  
275 therefore analyses were based on a total sample size  $N = 40$ , divided evenly between tinnitus and  
276 control groups. For supplemental sex-separated analyses, the four subgroups (tinnitus male,  
277 tinnitus female, control male, and control female) were each sized  $n = 10$ .

### 278 **3. Results**

#### 279 **3.1 Noise exposure history**

280 Participants with TNA reported greater lifetime exposure than controls to sound levels over 80 dBA,  
281 Wilcoxon-Mann-Whitney  $U = 283$ ,  $p = 0.02$ . However, as can be seen from Fig. 3, the spread of  
282 exposure values was greater for the TNA group, with some tinnitus participants presenting exposure  
283 scores in the same range as those of controls.

#### 284 **3.2 Auditory brainstem response**

285 All participants produced unambiguous ABRs bilaterally, with waves I and V clearly evident at  
286 appropriate latencies. (Automatically interpreted waveforms are presented in Supplementary  
287 Material. Grand average waveforms are displayed in Fig. 4A.) Resulting amplitude and latency data  
288 are given in Table 4 (Supplementary Material).

289 As can be seen from Fig. 4B, the amplitude of ABR wave I was not significantly reduced in  
290 participants with tinnitus relative to controls,  $t(37.0) = -0.11$ ,  $p = 0.91$ , Student's t-test. Note that had  
291 a one-tailed test been applied to these data, the result would have remained non-significant,  $p =$   
292  $0.46$ . Measurement variability was low (coefficient of variation 0.26 in controls, 0.30 in tinnitus),  
293 giving statistical power of 90% ( $\alpha = 0.05$ , one-tailed) to detect the 26% reduction in wave I amplitude  
294 for tinnitus versus controls reported by Schaette and McAlpine (2011) for a 100 dB peSPL click.

295 In an attempt to manage non-synaptopathic sources of variability in ABR amplitude, we computed  
296 the ratio of wave I to wave V amplitude, thought to provide a measure of central gain in the auditory  
297 brainstem (Schaette and McAlpine, 2011). This self-normalized difference measure did not differ

298 significantly between groups, Wilcoxon-Mann-Whitney  $U = 192$ ,  $p = 0.84$ . Nor did the amplitude of  
299 wave V,  $t(34.7) = 0.60$ ,  $p = 0.55$ , Student's t-test. Supplemental sex-separated analyses revealed no  
300 significant effects of tinnitus on wave I amplitude (female  $p = 0.56$ , male  $p = 0.54$ , Student's t-tests)  
301 nor on wave I/V amplitude ratio (female  $p = 0.52$ , unequal variance t-test; male  $p = 0.44$ , Wilcoxon-  
302 Mann-Whitney test).

### 303 **3.3 Envelope following response**

304 EFRs to stimuli of both modulation depths exceeded the noise floor for all participants, allowing  
305 analysis of both EFR amplitude (Fig. 5A) and the EFR difference measure (dB difference in  
306 response amplitude at the two modulation depths, Fig. 5B). The transposed tone with shallow  
307 modulations invariably elicited a lower EFR amplitude than the fully modulated stimulus, yielding  
308 consistently positive values of the EFR difference measure (see Table 5 of Supplementary  
309 Material). EFR amplitudes were entered into a two-way ANOVA, with tinnitus group as a between-  
310 subjects factor and stimulus modulation depth as a within-subject factor. There was a non-  
311 significant main effect of group,  $F(1,38) = 2.83$ ,  $p = 0.10$ , with tinnitus subjects producing lower  
312 response amplitudes than controls. The absence of a significant interaction effect indicates that  
313 tinnitus is not significantly associated with differences in the EFR difference measure,  $F(1,38) =$   
314  $0.324$ ,  $p = 0.57$ . When the same analysis was performed on each sex separately, the results  
315 revealed no significant effects of tinnitus on EFR amplitude (male  $p = 0.29$ ; female  $p = 0.23$ ), nor  
316 significant interactions between group and depth (male  $p = 0.31$ ; female  $p = 0.81$ ).

### 317 **3.4 Correlations between noise exposure and electrophysiological measures**

318 Pearson's product-moment correlation coefficients were computed to test the linear relations  
319 between log-transformed units of lifetime noise exposure and the various measures of neural  
320 function (Fig. 6). No association was evident between noise exposure and the amplitude of ABR  
321 wave I,  $r = 0.15$ ,  $p = 0.36$ , nor between noise exposure and the ratio of wave I to wave V amplitude,  
322  $r = 0.15$ ,  $p = 0.35$ . Nor did noise exposure relate to EFR amplitude at a shallow modulation depth,  $r$   
323  $= 0.01$ ,  $p = 0.94$ , or to the EFR difference measure,  $r = -0.16$ ,  $p = 0.31$ . Note that in the latter case, it  
324 is predicted that the measure should increase with increasing noise exposure.

## 325 **4. Discussion**

### 326 **4.1 A role for noise exposure in tinnitus with a normal audiogram**

327 Reported lifetime noise exposure of tinnitus subjects exceeded that of controls, despite close  
328 matching on the basis of sex, age, and audiometric thresholds. To the authors' knowledge, these  
329 data represent the first published evidence implicating noise exposure in tinnitus without threshold  
330 elevation. Previous research has associated excessive noise exposure and tinnitus in normally  
331 hearing young people (Davis et al., 1998; Meyer-Bisch, 1996) but not through comparison with  
332 audiometrically matched controls. Hence noise exposure in previous reports may have been related  
333 to tinnitus through sub-clinical threshold changes.

334 In contrast, our tinnitus group exhibited no significant reduction in hearing sensitivity at any of 10  
335 measurement frequencies between 0.25 and 14 kHz. Though we cannot rule out the existence of  
336 narrow audiometric "notches" in our tinnitus subjects, undetected by standard audiometry (Zhao et  
337 al., 2014), these findings nonetheless cast new light on the hazards of noise to the auditory system.  
338 It seems that excessive noise exposure can induce changes in auditory function that spare the  
339 audiogram, even at high frequencies, and yet may lead to disturbing perceptual consequences.

### 340 **4.2 No ABR evidence for tinnitus-related or noise-induced synaptopathy**

341 The nature of these noise-induced changes is very much less clear, since our measures revealed  
342 no evidence for cochlear synaptopathy in TNA. In particular, the expected reduction of ABR wave I  
343 amplitude was not observed. This finding stands in contrast with those of Schaette and McAlpine  
344 (2011), whose TNA subjects exhibited reduced wave I amplitudes relative to matched controls:  
345 reductions of 25% and 26% at 90 and 100 dB peSPL, respectively. (Fig. 7 compares Schaette and  
346 McAlpine's 100 dB data with the data obtained in the present study.)

347 Type II error is unlikely to account for these divergent findings, since post-hoc power analysis for the  
348 present study indicates 90% power to detect a 26% reduction in wave I amplitude (see Section 3.2).  
349 This is despite inclusion of participants of both sexes, which might reasonably be expected to  
350 increase ABR amplitude variability. The present study's wave I amplitude data are less variable than

351 those of Schaette and McAlpine, perhaps due to the use of research-grade recording equipment.  
352 Therefore, other possible explanations for our null result must be considered.

353 It is plausible that differences in participant age between the two studies are responsible, an  
354 explanation which would have important implications for our understanding of both cochlear  
355 synaptopathy and tinnitus heterogeneity. Participants in the present study were considerably  
356 younger (mean tinnitus age 25.7 years, control 25.5 years) than those of Schaette and McAlpine  
357 (mean tinnitus age 36.3 years, control 33.2 years). It may be that cochlear synaptopathy is a  
358 significant etiology of tinnitus with normal audiogram in older humans, but not among the very  
359 young, in whom other etiologies dominate.

360 It is therefore notable that evidence of human cochlear synaptopathy in relation to noise exposure is  
361 considerably less concrete than the evidence in relation to aging. Age-related loss of spiral ganglion  
362 cells was observed by Makary et al. (2011) in a large study of human temporal bones without  
363 significant hair cell loss. Parallel findings in mice (Sergeyenko et al., 2013) and preliminary synaptic  
364 counts in humans (Viana et al., 2015) strongly suggest that this decline is the delayed sequel to  
365 age-related cochlear synaptopathy progressing throughout the lifespan. In contrast, research  
366 relating human AN function to noise exposure has relied on electrophysiological measures, with  
367 mixed results. The results of the present study show no relation of lifetime noise exposure to ABR  
368 wave I amplitude, nor to ABR wave I/V amplitude ratio. Previously, Stamper and Johnson (2015a)  
369 reported a negative relation between noise exposure (estimated over the previous 12 months) and  
370 ABR wave I amplitude, but results were confounded by sex. Subsequent sex-separated analysis  
371 revealed that the correlation was present only in females in response to a 120 dB peSPL stimulus.  
372 Using electrocochleography in college students, Liberman et al. (2016) found no significant  
373 association between reported noise exposure and the amplitude of the compound action potential  
374 (equivalent to ABR wave I), although a noise-related enhancement of the summing potential was  
375 observed. In a large study of 126 normally hearing young listeners, Prendergast et al. (2016)  
376 demonstrated no relation between lifetime noise exposure and wave I amplitude or EFR  
377 synchronization strength.

378 One explanation for this pattern of results is that audiometrically normal humans do not exhibit  
379 substantial synaptopathy solely as a result of noise exposure. Other possible explanations exist,



380 such as insensitivity of electrophysiological measures (discussed later in Section 4.2) and diverse  
381 genetic susceptibility to synaptopathy in humans, who might have “tough” and “tender” ears  
382 (Henderson et al., 1993). However, it remains plausible that synaptopathy arises in humans due  
383 primarily to aging, or to an interaction between aging and noise exposure (as demonstrated in mice  
384 by Fernandez et al., 2015). This manifestation would represent a divergence from mouse models,  
385 but increasing evidence suggests that such inter-species differences are to be expected. Noise-  
386 induced synaptopathy in guinea pigs requires higher sound levels than in mice and long-term  
387 degeneration of spiral ganglion cells is less pronounced (Lin et al., 2011). In stark contrast with  
388 mouse data, guinea pig synapses damaged by noise appear largely repairable (Liu et al., 2012; Shi  
389 et al., 2013), leading to only transient changes in the distribution of spontaneous rates among AN  
390 fibers (Song et al., 2016). Early indications from a macaque model suggest that primates may  
391 exhibit even greater resistance to noise-induced synaptopathy (Burton et al., 2016).

392 Alternatively, it is conceivable that synaptopathy exists in audiometrically normal young humans, but  
393 is limited to extremely basal cochlear regions. This possibility is suggested by differences in ABR  
394 stimulus bandwidth between the present study and that of Schaette and McAlpine (2011). In order  
395 to limit the unwanted influence of very high frequency audiometric loss, we selected stimuli with a  
396 10 dB bandwidth extending from 1.2 to 4.7 kHz. By comparison, our measurements indicate that the  
397 10 dB bandwidth of Schaette and McAlpine’s 100 dB clicks extends to 7.1 kHz (recorded in a Bruel  
398 and Kjaer 4153 artificial ear coupled to TDH-49 headphones). The high presentation level of our  
399 stimuli ought to elicit the “half-octave basalward shift” in the travelling wave, leading to strong  
400 excitation of characteristic frequencies up to approximately 7 kHz. With the addition of upward  
401 spread of excitation, the stimulated region should encompass the 3 to 6 kHz characteristic  
402 frequency region where early noise damage is usually manifest (Coles et al., 2000). Nevertheless, it  
403 remains possible that synaptopathy existed in our tinnitus cohort, but was restricted to even higher  
404 frequencies. Participants generally reported tinnitus with a high frequency percept (ringing or  
405 hissing) and tinnitus pitch was not measured.

406 A crucial and related issue is that of high frequency audiometric loss and its influence on ABR wave  
407 I. It is possible that the ABR findings of Schaette and McAlpine (2011) and Gu et al. (2012) reflect  
408 basal loss of sensitivity in tinnitus participants, rather than an audiometrically “hidden” hearing loss.  
409 Failure to replicate these findings might indicate robustness of our methods against the unwanted

410 influence of audiometric loss, given the audiometric and stimulus differences between the present  
411 study and the previous reports. Wave I of the ABR is dominated by contributions from high  
412 frequency portions of the cochlear partition, where reduced dispersion enhances the synchrony of  
413 neuronal firing (Don and Eggermont, 1978). At high stimulus levels, upward spread of excitation  
414 involves increasingly basal generators (Eggermont and Don, 1980). Hence the unambiguous  
415 interpretation of wave I amplitude may require careful control of audiometric thresholds at  
416 frequencies well beyond the bandwidth of the ABR stimuli. The present study used not only a  
417 narrower stimulus bandwidth than the previous studies, but closer audiometric matching (group  
418 means differed by  $< 1$  dB at 10 and 14 kHz). Schaette and McAlpine's groups differed in  
419 audiometric sensitivity at 12 kHz, where mean threshold for the tinnitus group was  $\sim 3.5$  dB higher  
420 than for controls. Missing data (from five tinnitus subjects and three control subjects) prevented  
421 comparison at higher frequencies. Similarly, Gu et al. (2012) reported a significant reduction in wave  
422 I amplitude only for their 120 dB peSPL stimulus, for which missing ABR data led to systematic  
423 differences between groups in high frequency hearing sensitivity (tinnitus group had  $\sim 10$  dB higher  
424 thresholds at 14 kHz). The band-limited ABR stimuli used in these studies fall within the low-  
425 frequency tails of high-frequency AN fiber tuning curves, and hence the response of these fibers  
426 should be relatively unaffected by outer hair cell dysfunction at least (Liberman and Dodds, 1984).  
427 However, it remains possible that tinnitus-related ABR differences in previous reports were at least  
428 partially driven by basal loss of sensitivity.

429 Finally, it is worth considering that absence of ABR evidence for tinnitus-related synaptopathy might  
430 reflect insensitivity of the ABR rather than absence of synaptopathy. In addition to the variability of  
431 ABR amplitude, which has many sources and might obscure neuropathic effects, the findings of  
432 Bourien et al. (2014) cast doubt on the fundamental contribution of low-SR fibers to ABR wave I  
433 (see Section 1). Ongoing attempts to develop more sensitive electrophysiological measures of  
434 cochlear neuropathy are clearly warranted.

### 435 **4.3 No EFR evidence for tinnitus-related or noise-induced synaptopathy**

436 Several alternatives to the ABR have been proposed as viable measures of synaptopathy in  
437 humans, including the amplitude ratio of the compound action potential to the summing potential  
438 (Liberman et al., 2016) and round window neural noise (Batrel et al., 2016). Among them, the EFR

439 has shown promise in both animals and humans and has the advantage of being recordable non-  
440 invasively, without the use of ear canal or transtympanic electrodes. However, the relation of the  
441 EFR to AN function is difficult to interpret, since contributions from different auditory centers are not  
442 separated in time as they are for the ABR, and the resulting response is dependent on neural  
443 function central to the AN. Additionally, and in common with the ABR, EFR amplitude reflects many  
444 non-synaptopathic sources of variability. Hence researchers have sought to innovative EFR  
445 measures with enhanced sensitivity to synaptopathy. The difference measure devised by Bharadwaj  
446 et al. (2015) - the slope of the function relating EFR amplitude to stimulus modulation depth - was  
447 intended as a sensitive, self-normalized measure of low-SR fiber loss. EFR slope was shown to  
448 correlate with behavioral measures of temporal coding and auditory selective attention, with  
449 individual differences tentatively attributed to synaptopathy (Bharadwaj et al, 2015).

450 The present study utilized an EFR difference measure very closely related to that of Bharadwaj and  
451 colleagues: the difference in EFR amplitude (in dB) at two stimulus modulation depths. Many  
452 stimulus characteristics were also shared with the previous study: level, duration, carrier frequency,  
453 modulation frequency, and off-frequency masking characteristics. Yet this measure was not  
454 associated with tinnitus status, nor with lifetime noise exposure. These results might be taken to  
455 indicate lack of noise-induced or tinnitus-related cochlear synaptopathy in our cohort. However, it is  
456 also possible that this pathology is not, after all, a major source of individual differences in EFR  
457 slope. The hypothesized sensitivity of the measure to synaptopathy relies upon several  
458 assumptions, including preferential damage to low-SR fibers in humans and saturation of high-SR  
459 units by stimuli with shallow modulations. There is some evidence, for example, that the high-SR  
460 fiber dynamic range for modulated stimuli considerably exceeds that for steady-state stimuli (Smith  
461 and Brachman, 1980). Interpretation of the present results would be aided by validation of the EFR  
462 slope measure in an animal model of synaptopathy.

463 Methodological differences between the present study and that of Bharadwaj et al. (2015) are also  
464 to be considered, though they appear unlikely to compromise sensitivity. The earlier study  
465 computed slopes using a minimum modulation depth of -8 dB, employing multichannel recording  
466 and principal component analysis to enhance response SNR. The present study used a single  
467 channel and selected a -6 dB minimum modulation depth to ensure that all responses exceeded the  
468 noise floor. However, Bharadwaj and colleagues reported that temporal perceptual performance

469 correlated not only with EFR slope but also with raw EFR amplitude for a -4 dB depth, implying that  
470 extremely shallow modulations were not an essential stimulus feature.

471 In addition to the EFR difference measure, the present study also analyzed straightforward EFR  
472 amplitude. EFR amplitude was not associated with lifetime noise exposure and did not differ  
473 significantly between tinnitus and control groups. Data from a mouse model indicate that EFR  
474 amplitude can be a robust measure of cochlear synaptopathy, but suggest that some features of our  
475 stimuli (and those of Bharadwaj et al., 2015) were suboptimal (Shaheen et al., 2015). The  
476 researchers used fully modulated EFR stimuli, optimized to enhance the contribution of the AN, and  
477 found that synaptopathy led to greater changes in EFR amplitude than in EFR phase locking value  
478 or ABR amplitude. Optimum sensitivity was achieved with high modulation frequencies (~ 1 kHz),  
479 which limited the influence of more central nuclei. In contrast, the present study used a much lower  
480 modulation frequency and likely elicited the responses of higher centers, where the effects of  
481 deafferentation might be mitigated by enhanced central gain (Brotherton et al., 2015; Chambers et  
482 al., 2016). Hence the present EFR amplitude data must be interpreted with caution. The observed  
483 trend for lower amplitudes in TNA was not significant, but it is possible that stimuli with higher  
484 modulation rates might have been more effective in revealing AN temporal coding deficits. Future  
485 investigation of cochlear synaptopathy in humans might be well served by optimized EFR measures  
486 paralleling those applied successfully in rodent models.

#### 487 **4.4 Conclusions**

488 The ABR and EFR results of the present study provide no evidence for cochlear synaptopathy in  
489 young humans with tinnitus and normal audiometric thresholds. Nor do these electrophysiological  
490 measures relate to lifetime noise exposure, providing no evidence for noise-induced synaptopathy  
491 in this cohort. It is importance to emphasize, however, that our results do not imply that  
492 synaptopathy is not prevalent in humans. It is possible, for example, that synaptopathy would have  
493 been measurable in an older population, through assessment of characteristic frequencies above 7  
494 kHz, or through use of a more sensitive measure.

495 Tinnitus participants are, as a group, more noise exposed than controls, though also more  
496 heterogeneous in this regard. Uncertainty about mechanisms notwithstanding, the findings relating

497 noise exposure and TNA have important implications. Even in tinnitus sufferers whose audiometric  
498 thresholds are indistinguishable from those of controls, symptoms may arise from sub-clinical  
499 damage due to excessive noise exposure.

## 500 **Acknowledgments**

501 The authors are grateful to two anonymous reviewers for constructive comments on an earlier  
502 version of the manuscript. The authors are also grateful to Dr Hari Bharadwaj for providing EFR  
503 recording software, and to Keith Wilbraham, Dr Michael Stone, and Dr Richard Baker for essential  
504 technical advice. The research was supported by an Action on Hearing Loss studentship, funded by  
505 the Marston Family Foundation, and by the Medical Research Council UK (MR/L003589/1).

## 506 **References**

- 507 Barnea, G., Attias, J., Gold, S., Shahar, A., 1990. Tinnitus with Normal Hearing Sensitivity:  
508 Extended High-Frequency Audiometry and Auditory-Nerve Brain-Stem-Evoked Responses. *Audiol.*  
509 29, 36-45.
- 510 Batrel, C., Huet, A., Desmadryl, G., Puel, J.L., Bourien, J., 2016. Peri-stimulus time response of the  
511 auditory nerve recorded at the round window. *Assoc. Res. Otolaryngol. Abs.*, 199.
- 512 Bernstein, L.R., Trahiotis, C., 2002. Enhancing sensitivity to interaural delays at high frequencies by  
513 using "transposed stimuli". *J. Acoust. Soc. Am.* 112, 1026-1036.
- 514 Bharadwaj, H.M., Masud, S., Mehraei, G., Verhulst, S., Shinn-Cunningham, B.G., 2015. Individual  
515 differences reveal correlates of hidden hearing deficits. *J. Neurosci.* 35, 2161-2172.
- 516 Bourien, J., Tang, Y., Batrel, C., Huet, A., Lenoir, M., Ladrech, S., Desmadryl, G., Nouvian, R., Puel,  
517 J.L., Wang, J., 2014. Contribution of auditory nerve fibers to compound action potential of the  
518 auditory nerve. *J. Neurophysiol.* 112, 1025-1039.129
- 519 British Society of Audiology, 2011. Pure-tone air-conduction and bone-conduction threshold  
520 audiometry with and without masking. Reading, UK: British Society of Audiology.
- 521 Brotherton, H., Plack, C.J., Maslin, M., Schaette, R., Munro, K.J., 2015. Pump Up the Volume:  
522 Could Excessive Neural Gain Explain Tinnitus and Hyperacusis? *Audiol. Neurotol.* 20, 273-282.
- 523 Burton, J., Hauser, S., Watson, J., Valero, M., Hackett, T., Ramachandran, R., 2016. Toward a  
524 Nonhuman Primate Model of Noise Induced Hearing Loss. *Assoc. Res. Otolaryngol. Abs.*, 533.

- 525 Chambers, A.R., Resnik, J., Yuan, Y., Whitton, J.P., Edge, A.S., Liberman, M.C., Polley, D.B., 2016.  
526 Central Gain Restores Auditory Processing following Near-Complete Cochlear Denervation. *Neuron*  
527 89, 867-79.
- 528 Coles, R.R.A., Lutman, M.E., Buffin, J.T., 2000. Guidelines on the diagnosis of noise-induced  
529 hearing loss for medico-legal purposes. *Clin. Otolaryngol.* 25, 264-273.
- 530 Davis A.C., Lovell E.A., Smith P.A., Ferguson M.A., 1998. The contribution of social noise to tinnitus  
531 in young people - A preliminary report. *Noise Health* 1, 40-46.
- 532 Don, M., Eggermont, J.J., 1978. Analysis of the click-evoked brainstem potentials in man using  
533 high-pass noise masking. *J. Acoust. Soc. Am.* 63, 1084-92.
- 534 Don, M., Ponton, C.W., Eggermont, J.J., Masuda, A., 1994. Auditory brainstem response (ABR)  
535 peak amplitude variability reflects individual differences in cochlear response times. *J. Acoust. Soc.*  
536 *Am.* 96, 3476-91.
- 537 Eggermont, J.J., Don, M., 1980. Analysis of the click-evoked brainstem potentials in humans using  
538 high-pass noise masking. II. Effect of click intensity. *J. Acoust. Soc. Am.* 68, 1671-5.
- 539 Furman, A.C., Kujawa, S.G., Liberman, M.C., 2013. Noise-induced cochlear neuropathy is selective  
540 for fibers with low spontaneous rates. *J. Neurophysiol.* 110, 577-586.
- 541 Gilles, A., Schlee, W., Rabau, S., Wouters, K., Franssen, E., Van de Heyning, P., 2016. Decreased  
542 Speech-In-Noise Understanding in Young Adults with Tinnitus. *Front. Neurosci.* 10, 288.
- 543 Gu, J.W., Herrmann, B.S., Levine, R.A., Melcher, J.R., 2012. Brainstem Auditory Evoked Potentials  
544 Suggest a Role for the Ventral Cochlear Nucleus in Tinnitus. *J. Assoc. Res. Otolaryngol.* 13, 819-  
545 833.
- 546 Henderson, D., Subramaniam, M., Boettcher, F.A., 1993. Individual susceptibility to noise-induced  
547 hearing loss: an old topic revisited. *Ear. Hear.* 14, 152-168.
- 548 Henry, J.A., Roberts, L.E., Caspary, D.M., Theodoroff, S.M., Salvi, R.J., 2014. Underlying  
549 Mechanisms of Tinnitus: Review and Clinical Implications. *J. Am. Acad. Audiol.* 25, 5-22.
- 550 Henry, J.A., Griest, S., Thielman, E., McMillan, G., Kaelin, C., Carlson, K.F., 2016. Tinnitus  
551 Functional Index: Development, validation, outcomes research, and clinical Application. *Hear.*  
552 *Res.* 334, 58-64.
- 553 Joris, P.X., Schreiner, C. E. & Rees, A., 2004. Neural Processing of Amplitude-Modulated Sounds.  
554 *Physiol. Rev.* 84, 541-577.
- 555 Kujawa, S.G., Liberman, M.C., 2009. Adding insult to injury: Cochlear nerve degeneration after  
556 "temporary" noise-induced hearing loss. *J. Neurosci.* 29, 14077-14085.
- 557 Kuwada, S., Anderson, J.S., Batra, R., Fitzpatrick, D.C., Teissier, N., D'Angelo, W.R., 2002.  
558 Sources of the scalp-recorded amplitude-modulation following response. *J. Am. Acad. Audiol.* 13,  
559 188-204.

- 560 Liberman, M.C., Dodds, L.W., 1984. Single-neuron labeling and chronic cochlear pathology. III.  
561 Stereocilia damage and alterations of threshold tuning curves. *Hear. Res.* 16, 55-74.
- 562 Liberman, M.C., Epstein, M.J., Cleveland, S.S., Wang, H., Maison, S.F., 2016. Toward a Differential  
563 Diagnosis of Hidden Hearing Loss in Humans. *PLoS ONE* 11, e0162726.
- 564 Lin, H.W., Furman, A.C., Kujawa, S.G., Liberman, M.C., 2011. Primary neural degeneration in the  
565 Guinea pig cochlea after reversible noise-induced threshold shift. *J. Assoc. Res. Otolaryngol.* 12,  
566 605-616.
- 567 Liu, L., Wang, H., Shi, L., Almklass, A., He, T., Aiken, S., Bance, M., Yin, S., Wang, J., 2012. Silent  
568 Damage of Noise on Cochlear Afferent Innervation in Guinea Pigs and the Impact on Temporal  
569 Processing. *PLoS ONE* 7, e49550.
- 570 Lutman, M.E., Davis, A.C., Ferguson, M.A., 2008. Epidemiological Evidence for the Effectiveness of  
571 the Noise at Work Regulations, RR669. Sudbury, UK: Health and Safety Executive.
- 572 Makary, C., Shin, J., Kujawa, S., Liberman, M.C., Merchant, S., 2011. Age-Related Primary  
573 Cochlear Neuronal Degeneration in Human Temporal Bones. *J. Assoc. Res. Otolaryngol.* 12, 711-  
574 717.
- 575 Meyer-Bisch, C., 1996. Epidemiological evaluation of hearing damage related to strongly amplified  
576 music (personal cassette players, discotheques, rock concerts)--high-definition audiometric survey  
577 on 1364 subjects. *Audiol.* 35, 121-42.
- 578 Michalewski, H.J., Thompson, L.W., Patterson, J.V., Bowman, T.E., Litzelman, D., 1980. Sex  
579 differences in the amplitudes and latencies of the human auditory brain stem potential.  
580 *Electroencephalogr. Clin. Neurophysiol.* 48, 351-6.
- 581 Nicolas-Puel, C., Faulconbridge, R.L., Guitton, M., Puel, J., Mondain, M., Uziel, A., 2002.  
582 Characteristics of Tinnitus and Etiology of Associated Hearing Loss: A Study of 123 Patients. *Int.*  
583 *Tinnitus. J.* 8, 37-44
- 584 Peake, W.T., Kiang, N.Y.S., 1962. Cochlear Responses to Condensation and Rarefaction Clicks.  
585 *Biophys. J.* 2, 23-34.
- 586 Plack, C.J., Léger, A., Prendergast, G., Kluk, K., Guest, H., Munro, K.J., in press. Toward a  
587 Diagnostic Test for Hidden Hearing Loss. *Trends in Hearing*.
- 588 Portnuff, C.D., Fligor, B.J., Arehart, K.H., 2011. Teenage use of portable listening devices: a hazard  
589 to hearing? *J. Am. Acad. Audiol.* 22, 663-77.
- 590 Prendergast, G., Guest, H., Munro, K., Kluk, K., Léger, A., Hall, D., Heinz, M., Plack, C., 2016.  
591 Effects of noise exposure on young adults with normal audiograms I: Electrophysiology. *Hear. Res.*  
592 Advance online publication. doi: 10.1016/j.heares.2016.10.028
- 593 R Core Team, 2015. R: A language and environment for statistical computing. Vienna, Austria: R  
594 Foundation for Statistical Computing. URL <http://www.R-project.org/>.

- 595 Sachs, M.B., Abbas, P.J., 1974. Rate versus level functions for auditory nerve fibers in cats: tone-  
596 burst stimuli. *J. Acoust. Soc. Am.* 56, 1835–1847.
- 597 Sanchez, T.G., de Medeiros, Í.R.T., Levy, C.P.D., Ramalho, J.R.O., Bento, R. F., 2005. Tinnitus in  
598 normally hearing patients: clinical aspects and repercussions. *Rev. Bras. Otorrinolaringol.* 71, 427-  
599 31.
- 600 Schaette, R., 2014. Tinnitus in men, mice (as well as other rodents), and machines. *Hear. Res.* 311,  
601 63-71.
- 602 Schaette, R., McAlpine, D., 2011. Tinnitus with a Normal Audiogram: Physiological Evidence for  
603 Hidden Hearing Loss and Computational Model. *J. Neurosci. Off. J. Soc. Neurosci.* 31, 13452-  
604 13457.
- 605 Sergeyenko, Y., Lall, K., Liberman, M.C., Kujawa, S.G., 2013. Age-related cochlear synaptopathy:  
606 an early-onset contributor to auditory functional decline. *J. Neurosci. Off. J. Soc. Neurosci.* 33,  
607 13686-13694.
- 608 Shaheen, L.A., Valero, M.D., Liberman, M.C., 2015. Towards a Diagnosis of Cochlear Neuropathy  
609 with Envelope Following Responses. *J. Assoc. Res. Otolaryngol.* 16, 727-745.
- 610 Shi, L., Liu, L., He, T., Guo, X., Yu, Z., Yin, S., Wang, J., 2013. Ribbon Synapse Plasticity in the  
611 Cochleae of Guinea Pigs after Noise-Induced Silent Damage. *PLoS ONE* 8, e81566.
- 612 Smith, R.L., Brachman, M.L., 1980. Response modulation of auditory-nerve fibers by AM stimuli:  
613 Effects of average intensity. *Hear. Res.* 2, 123-133.
- 614 Song, Q., Shen, P., Li, X., Shi, L., Liu, L., Wang, J., Yu, Z., Stephen, K., Aiken, S., Yin, S., Wang, J.,  
615 2016. Coding deficits in hidden hearing loss induced by noise: the nature and impacts. *Sci. Rep.* 6,  
616 25200.
- 617 Stamper, G.C., Johnson, T.A., 2015a. Auditory function in normal-hearing, noise-exposed human  
618 ears. *Ear. Hear.* 36, 172-84.
- 619 Stamper, G.C., Johnson, T.A., 2015b. Letter to the Editor: Examination of Potential Sex Influences  
620 in Stamper, G.C., Johnson, T.A., 2015a. Auditory Function in Normal-Hearing, Noise-Exposed  
621 Human Ears, *Ear. Hear.* 36, 172–184. *Ear. Hear.* 36, 738-740.
- 622 Taberner, A.M., Liberman, M.C., 2005. Response properties of single auditory nerve fibers in the  
623 mouse. *J. Neurophysiol.* 93, 557-69.
- 624 The Mathworks, Inc., 2013. MATLAB Release 2013a. Natick, Massachusetts: The MathWorks, Inc.
- 625 Trune, D.R., Mitchell, C., Phillips, D.S., 1988. The relative importance of head size, gender and age  
626 on the auditory brainstem response. *Hear. Res.* 32, 165-74.
- 627 Viana, L.M., O'Malley, J.T., Burgess, B.J., Jones, D.D., Oliveira, C.A.C.P., Santos, F., Merchant, S.  
628 N., Liberman, L.D., Liberman, M.C., 2015. Cochlear neuropathy in human presbycusis: Confocal  
629 analysis of hidden hearing loss in post-mortem tissue. *Hear. Res.* 327, 78-88.



- 630 Zhao, F., Stephens, S.D., Ishak, W.S., Meyer-Bisch, C., 2014. The characteristics of Audioscan and  
631 DPOAE measures in tinnitus patients with normal hearing thresholds. *Int. J. Audiol.* 53, 309-17.

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632 **Figure Captions**

633 Table 1. Tinnitus characteristics.

634 Fig. 1. Audiometric thresholds for tinnitus and control groups, presented as group mean  $\pm$  standard  
635 error of the mean. A: Pure tone audiometric thresholds. Groups means differ by  $<2.25$  dB at all  
636 frequencies. B: High frequency thresholds for 1/3-octave narrowband noise using a three-interval,  
637 three-alternative, forced-choice paradigm and a two-down, one-up rule. Group means differ by  $<1$   
638 dB.

639 Fig. 2. A schematic illustration of the EFR paradigm, including responses and response spectra  
640 from a single participant. Analyzed measures were the raw response amplitude at the frequency of  
641 interest, 100 Hz, and an EFR difference measure comparing response amplitudes at two stimulus  
642 modulation depths. It was predicted that loss of low-SR fibres should primarily impair responses at  
643 the shallow modulation depth, leading to higher values of the difference measure in synaptopathic  
644 ears.

645 Fig. 3. Units of lifetime noise exposure for participants in tinnitus and control groups. Points  
646 correspond to individual participants, upper and lower hinges to first and third quartiles, upper  
647 whiskers to the highest value within  $1.5 * \text{IQR}$  of the upper hinge (where IQR is the interquartile  
648 range), and lower whiskers to the lowest value within  $1.5 * \text{IQR}$  of the lower hinge.

649 Fig. 4. ABRs in response to 102 dB peSPL clicks for tinnitus and control groups. A: Grand average  
650 waveforms. Shaded areas correspond to the standard error of the mean. B: Wave I and wave V  
651 amplitudes, presented as mean  $\pm$  standard error of the mean.

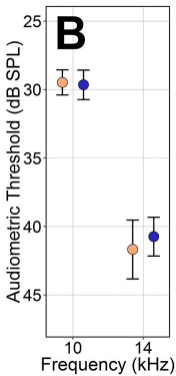
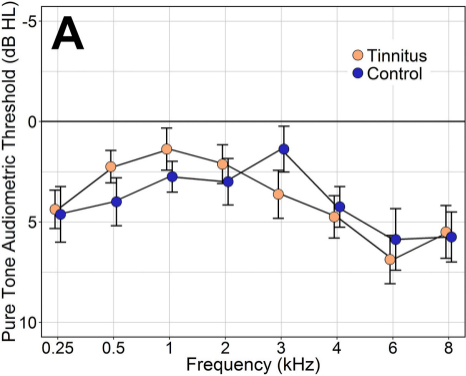
652 Fig. 5. EFR measures for tinnitus and control groups, presented as group mean  $\pm$  standard error of  
653 the mean. A: EFRs to transposed tones with a shallow ( $-6$  dB) and full ( $0$  dB) modulation depth. The  
654 tinnitus-related reduction in response amplitude is non-significant. The lines connecting the  
655 responses illustrate the “EFR slope” measure devised by Bharadwaj et al. (2015), though defined by

656 a two-point function. B: The difference in EFR amplitude (in dB) at the two modulation depths. The  
657 hypothesized enhancement in the tinnitus group is not evident.

658 Fig. 6. Relations between lifetime noise exposure and electrophysiological measures of cochlear  
659 synaptopathy, including both raw amplitude measures and self-normalized difference measures.  
660 Shaded areas represent 95% confidence limits of linear regression lines for all subjects. Marginal  
661 density plots represent tinnitus and control group distributions. No significant correlation is evident  
662 between noise exposure and any electrophysiological measure. A: ABR wave I amplitude. B: ABR  
663 wave I/V amplitude ratio. C: EFR amplitude at a shallow (-6 dB) modulation depth. D: Difference in  
664 EFR amplitude (in dB) at two stimulus modulation depths. Note that D was hypothesized to exhibit a  
665 positive relation, whereas negative relations were expected in A to C.

666 Fig. 7. ABR data from the present study, elicited using 102 dB peSPL clicks, presented alongside  
667 those of Schaette and McAlpine (2011), elicited using 100 dB peSPL clicks. Points and error bars  
668 represent the mean  $\pm$  standard error of the mean. A: The raw amplitude of ABR wave I. B: The ratio  
669 of wave I amplitude to wave V amplitude.

Participant	Tinnitus location	Sound quality	Time since onset	Constant in quiet?	TFI score	Conscious awareness of tinnitus (% of waking hours)
1	Both ears	Ringling	9 years	Yes	26.8	30
6	Right ear	Ringling	2 years	Yes	28	30
7	Both ears	High pitched whine	10 years	Yes	22.8	30
8	Both ears (right louder than left)	Between whining and ringling	> 6 years	Yes	8.4	50
9	Both ears (right louder than left)	Ringling	14 years	Yes	29.6	60
10	Both ears	Shooshing	> 12 years	Yes	6.4	40
12	Both ears	Ringling	14 years	Yes	20.8	30
19	Both ears (right may be louder than left)	Buzzing	1 year	Yes	51.6	30
20	Both ears	High pitched tone	10 years	Yes	78	70
23	Both ears	Ringling	8 years	Yes	18	10
28	Both ears (left louder than right)	Ringling	2 years	Yes	32	20
29	Both ears (right louder than left)	Ringling	3 years	Yes	45.6	80
30	Central percept	Ringling or whining	1 year	Yes	48.4	50
32	Both ears	Ringling	8 years	Yes	23.6	40
34	Both ears	Ringling	As long as can remember	Yes	62	60
35	Both ears	Ringling	> 10 years	Yes	5.2	20
36	Both ears (left louder than right)	High frequency tone	7 years	Yes	24.4	30
37	Both ears	High pitched fridge noise	5 years	Yes	48	60
38	Can affect either ear	High pitched ringling	10 years	No (tinnitus lasts minutes to hours)	71.6	60
57	Both ears (left louder than right)	Ringling	4 months	Probably constant, but not certain	6.4	10



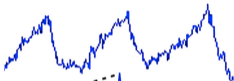
Shallow modulation (-6 dB)

Full modulation (0 dB)

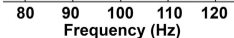
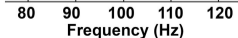
Stimulus



EFR  
in time domain



EFR  
magnitude  
spectrum



EFR difference measure  
( $\text{dB}_{\text{full}} - \text{dB}_{\text{shallow}}$ )

Units of Lifetime Noise Exposure

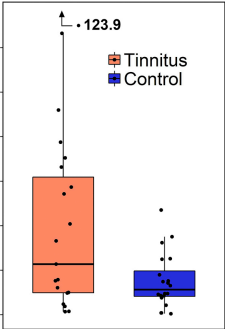
60  
50  
40  
30  
20  
10  
0

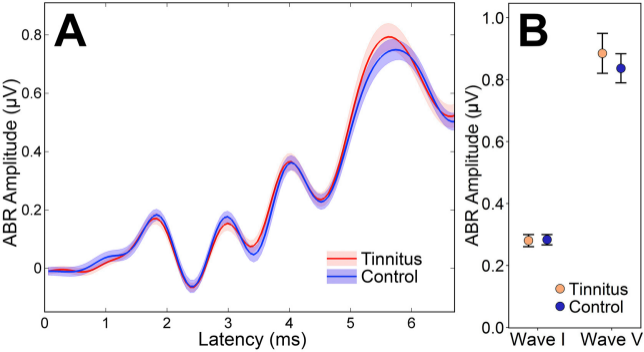
Tinnitus  
Control

Tinnitus

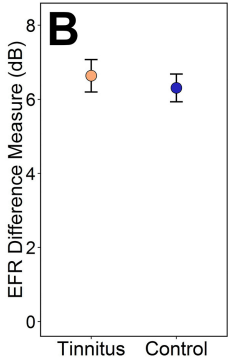
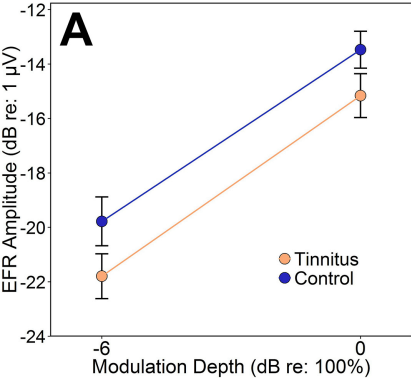
Control

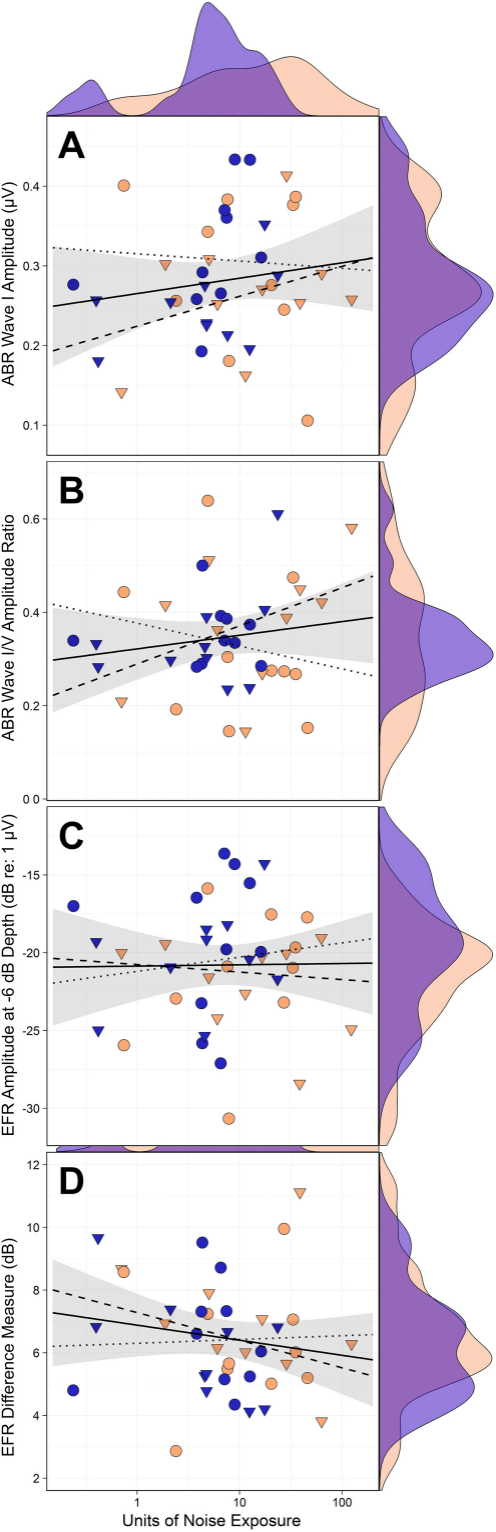
• 123.9

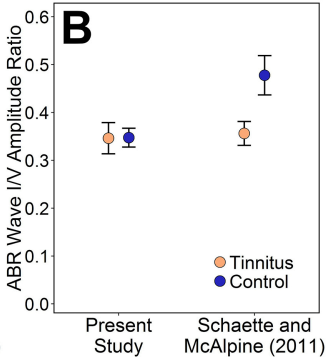
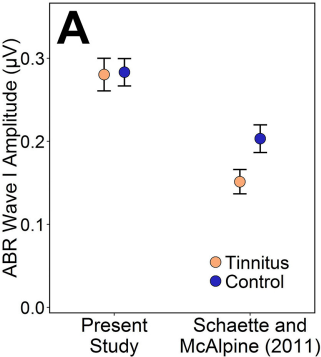












## Highlights

- Tinnitus participants matched with controls for age, sex, & audiogram up to 14 kHz
- Tinnitus participants more noise exposed, despite close audiometric matching
- No ABR or EFR evidence for cochlear synaptopathy in tinnitus participants
- No association between ABR or EFR measures and lifetime noise exposure

ACCEPTED MANUSCRIPT