

1 **New insights into the aquatic photo-induced toxicity of nine sulfonamide**
2 **antibiotics: Apparent photolysis, toxicity evolution and mechanisms**

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8
9 **ABSTRACT**

10 As newly recognized environmental contaminants, sulfonamide antibiotics (SAs) widely exist
11 in aqueous systems and photo-degrade significantly in sunlit surface waters, requiring new
12 insights into their aqueous apparent photolytic mechanisms and photo-modified toxicity. In this
13 study, reaction kinetics, self-sensitized photo-oxidation, transformation pathways, antibacterial
14 activity and photo-induced toxicity were investigated during the apparent photolysis of nine
15 SAs including sulfamethoxazole (SMX), sulfisoxazole, sulfathiazole, sulfamethizole,
16 sulfadimethoxine (SDM), sulfadiazine, sulfachloropyridazine, sulfamerazine and
17 sulfamethazine. Under simulated sunlight irradiation ($\lambda > 290$ nm), most five-membered
18 heterocyclic SAs underwent photodegradation at a faster rate than all six-membered
19 heterocyclic SAs, which was ascribed to their chemical structures including different
20 heterocyclic groups and associated substituents ($-\text{CH}_3$). The scavenging experiments indicated
21 that SMX underwent self-sensitized photo-degradation via $\text{O}_2^{\cdot-}$, $^1\text{O}_2$ and $\cdot\text{OH}$, and direct

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22 photo-degradation via $^3P^*$. The contribution of $O_2^{\cdot-}$ (40.71%–59.01%) and 1O_2 (29.13%)
23 mediated self-sensitized photo-oxidation was found to be the highest under acidic and alkaline
24 conditions, respectively. Photo-product identification was carried out with HPLC-MS², which
25 identified proposed transformation pathways of five-membered heterocyclic SMX which
26 involved photo-cleavage and photo-polymerization, while six-membered heterocyclic SDM
27 underwent S–N bond cleavage, hydroxylation, desulfonation, methylation and addition reaction
28 at sulfonyl-N. Based on multiple bioassays (*Escherichia coli* and *Vibrio fischeri*), the
29 antibacterial activity changes and photo-modified toxicity evolution appeared to depend on
30 dominant photo-degradation pathways and primary intermediates. Furthermore, the ECOSAR
31 model confirmed the comparable or higher toxicities of the intermediates compared to the
32 parent compounds. These findings provide new photochemical mechanistic insights into the
33 large class of SAs, which allows a better assessment of their photochemical persistence, fate
34 and risks in aquatic environments.

35 *Keywords:* sulfonamide antibiotics; self-sensitized photolysis; transformation products;
36 antibacterial activity; photo-modified toxicity

37

38 **1. Introduction**

39 Antibiotics have been considered as a critical environmental issue due to their widespread
40 occurrence, and potential harmful effects on microbial populations (Wang et al., 2024; Li et al.,
41 2023). When discharged into wastewater streams, most antibiotics are incompletely removed
42 by traditional treatment methods (Cao et al., 2024; Guo et al., 2025a), and as , they continuously
43 enter aquatic environments they can be considered to show pseudo-persistence (Wei et al.,
44 2024). Their persistence in aqueous systems may result in the propagation of antibiotic resistant
45 bacteria with multiple drug resistance, posing a serious risk to aquatic systems and human
46 health (Liu et al., 2023a; Hou et al., 2023). As a large important category of antibiotics,

47 sulfonamides (SAs) are extensively used in the aquaculture, animal husbandry and human
48 health. They have been detected in aquatic environments through a variety of monitoring studies
49 all over the world (Cao et al., 2024; Jia et al., 2023; Wang et al., 2023) (Table S1). Average
50 concentrations in surface waters of rivers and adjacent seas of China have been reported to
51 range from 0.1 to 150.83 ng L⁻¹ (Fig. S1). Therefore, given the ubiquity of the antibiotic
52 pollutants in aquatic systems, it is essential to explore their transformation in aqueous
53 environments, as well as to evaluate the subsequent toxicity and risks to aquatic organisms.

54 Previous studies have investigated the ecotoxicological effects of SAs on aquatic biota (Li et
55 al., 2025a; Huo et al., 2023; Zhu et al., 2021). By selectively inhibiting microorganisms in
56 ecological systems, SA residues can demonstrably influence natural microbial communities
57 (Välitalo et al., 2017), and result in ecotoxicological effects and resistance genes in
58 microorganisms (Zainab et al., 2020). Interestingly, the photo-transformation of SAs was
59 determined to have a important impact on their biological effects (Guo et al., 2021; Cheng et
60 al., 2021; Ge et al., 2019). In sunlight-irradiated surface waters, apparent photo-chemical
61 degradation is the main transformation pathway in determining the fate and risk of SAs (Tang
62 et al., 2024). Furthermore, photo-degradation irreversibly changes molecular structures, and
63 results in the formation of transformation products (TPs) including reactive species and organic
64 intermediates (Löffler et al., 2023; Kairigo et al., 2020). TPs may exhibit significantly different
65 physiochemical properties and higher toxicity to organisms, urging the consideration of the
66 photo-reaction types and photo-induced toxicity when assessing their environmental fate and
67 ecological risks.

68 When irradiated by sunlight, aqueous organic pollutants can not only undergo direct photo-
69 degradation via singlet and triplet excited states, but also undergo reactive-oxygen-species
70 (ROS, e.g., •OH, ¹O₂ and O₂^{•-}) mediated self-sensitized photo-oxidation (Hsu et al., 2019;
71 Zhang et al., 2018). These two distinct reactions collectively constitute an apparent photolysis

72 process. Clarifying the photo-reaction types and mechanisms is of great importance to assess
73 their environmental persistence and understand the risks they may pose. Previous studies have
74 reported apparent photo-degradation of SAs, showing that the photolytic efficiency of SAs was
75 significantly affected by the solution pH (Oliveira et al., 2019; Vibha et al., 2025). However, it
76 is not clear whether SAs undergo self-sensitized photo-oxidation, and which ROS might be
77 involved in the reaction. Therefore, it is necessary to quantify the contribution of self-sensitized
78 photo-degradation via ROS on their apparent photolysis at a range of pHs, and to reveal the
79 reactivity of different dissociated SA forms towards these ROS.

80 As for photo-induced bioeffects of SAs, previous studies have reported photo-sensitized and
81 photo-modified toxicities (Andino-Enrquez et al., 2025; Liu et al., 2023b; Zeng et al., 2021).
82 When compared with the dark controls, the continuous exposure with simulated sunlight
83 enhanced the acute toxicity of aqueous SAs to *Daphnia magna* (*D. magna*), attributing to the
84 generation of ROS and the subsequent photo-sensitized toxicity of the parent compounds (Jung
85 et al., 2008). As for the photo-modified toxicity of SAs, Xu et al. (2014) and Trov et al. (2009)
86 observed that the photo-degradation intermediates of SAs exhibited higher toxicity to *Vibrio*
87 *fischeri* and greater antibacterial capacity to *D. magna* than the parent compounds, respectively.
88 Similarly, Voigt et al. investigated the antimicrobial efficacy of photo-degraded SAs by
89 determining their minimal inhibitory concentrations (MIC) to *Pseudomonas fluorescens* and
90 *Bacillus subtilis* during the photo-degradation process, revealing that the MIC values of
91 photolyzed SAs exhibited 2.5- to 5-fold increase compared to those of the parent compounds
92 (Voigt et al., 2017). In addition, the inhibitory or stimulatory effects of photolytic products
93 several SAs (sulfamethoxazole (SMX), sulfacetamide (SA), sulfadiazine (SD) and sulfathiazole
94 (ST)) on the growth of *Calluna vulgaris* were also discovered (Zessel et al., 2014). These
95 previous studies have demonstrated the photo-generated toxicity of SAs to non-target
96 organisms, and provide theoretical support for new insights into the photo-induced toxicity of

97 SAs, though toxicity evolution and related mechanisms were not fully considered. To better
98 assess the photo-degradation induced ecological risks of SAs, photochemical transformation
99 products and pathways need to be identified, and the mechanisms of photo-induced toxicity
100 should be clarified.

101 This study seeks new insights into self-sensitized photo-degradation, photo-degradation
102 pathways, toxicity changes and mechanisms of nine SAs. These representative SAs selected as
103 model compounds have been frequently detected in surface waters and known to undergo solar
104 photo-degradation (Fig. S1) (Cao et al., 2024; Mac Loughlin et al., 2024). In order illustrate the
105 photo-induced toxicity mechanisms of SAs, their aqueous photo-degradation kinetics and key
106 by-products were investigated, along with an evaluation of their toxicity and antibacterial
107 activity changes during the photo-transformation processes. The present study provides a
108 comprehensive understanding of photochemical behavior and photo-modified toxicity of SAs,
109 which are important to better assess the environmental fate and ecological risks of the emerging
110 contaminants.

111

112 **2. Materials and methods**

113 *2.1 Reagents and materials*

114 Nine SAs (purity > 98 %) including SMX, sulfisoxazole (SIX), ST, sulfamethizole (SMT),
115 SDM, SD, sulfachloropyridazine (SCP), sulfamerazine (SM) and sulfamethazine (SMZ) were
116 obtained from different suppliers. The information related to SAs is given in Table S2.
117 Isopropanol (IPA), acetonitrile (ACN) and methanol (MeOH) were of HPLC grade and
118 purchased from Alfa Aesar and TEDIA (Fairfield, USA). Furfuryl alcohol (FFA, purity 98%)
119 and benzoic acid (BA, 98%) were procured from J&K Technology Co., Ltd. Hydrogen peroxide
120 (H₂O₂, 30%) was provided by Acros Organics. Other reagents such as benzoquinone were of
121 analytically pure (at least 97% purity) and used as received. Beef extract, peptone and agar

122 powder were purchased from Beijing Aoboxing Biology Technology Co., Ltd. *Escherichia coli*
123 (*E. coli*) and Freeze-dried *Vibrio fischeri* were supplied by the Shanghai Bioresource Collection
124 Center and the Zhejiang Yangtze Delta Region Institute of Tsinghua Univ., respectively. To
125 prepare photolytic solutions, ultrapure water (18.2 MΩ cm) was obtained with a Millipore Milli-
126 Q® water purifier.

127 2.2 Photolytic experiments

128 The emission spectra of the light sources (Fig S2) and the absorption spectra of the model
129 compounds (Fig S3) were determined by the Acton SP-300 monochromator (Princeton
130 Instruments, Inc.) and TU1901 UV-Visible spectrophotometer (Beijing Purkinje General
131 Instrument Co., Ltd.), respectively. To carry out the photochemical experiments, all solutions
132 of individual SAs ($C_0 = 10 \mu\text{M}$) were pipetted into quartz tubes (vol. 50 mL, dia. 20 mm), and
133 placed in photochemical reactor (Fig S4), of which the temperature was maintained at 25 °C.
134 The solutions were irradiated with a high-pressure mercury lamp (500 W, $\lambda > 290 \text{ nm}$), which
135 was cooled and filtered by a Pyrex-well (Ge et al., 2023; Luo et al., 2019). Using a UV
136 radiometer (Beijing Normal University photoelectric instrumental factory), the light intensities
137 in the center of the solutions were measured to be 7.52 Mw cm^{-2} at 420 nm and 5.32 Mw cm^{-2}
138 at 365 nm, respectively.

139 To determine if the SAs underwent self-sensitized photo-oxidation via $\bullet\text{OH}$, $^1\text{O}_2$ and $\text{O}_2^{\bullet-}$ as
140 well as direct photolysis via $^3\text{P}^*$, SMX was selected as a representative compound, and included
141 RS scavenging experiments with IPA, triethylene diamine (DABCO), *p*-benzoquinone (*p*-BQ)
142 and 2,4,6-trimethylphenol (TMP) were undertaken. With reference to a previous study (Cui et
143 al., 2025), the oxidation reactivity of SA towards these RS was investigated using competing
144 kinetic experiments. The methods for calculating the self-sensitized photo-oxidation
145 contribution ratios (R_{ROS}) and bimolecular reaction rate constants are described in S1 and S2,
146 respectively. These photolytic experiments and dark controls were carried out at least in

147 triplicate. In order to analyze the SA concentrations and identify the key transformation
148 products, an Agilent 1260 HPLC and an Agilent 1260-6470B HPLC-MS/MS were employed,
149 respectively. The analysis methods were detailed in the [Text S3](#).

150 2.3 Antibacterial bioassay

151 The antibacterial activities of the SA photo-degradation solutions against *E. coli* were
152 determined to clarify the changes of antibacterial activity as a result of their photo-degradation.
153 The bioassay was performed in accordance with China standard method QB/T 2738-2012 ([Ge
154 et al., 2015](#)), with *E. coli* as the indicator of vitro bacteriostatic experiment. The diameters of
155 inhibition zone were determined as the index for antibacterial activity ([Tao et al., 2010](#);
156 [Klančnik et al., 2010](#)). The significant differences in antibacterial activity between the
157 photolyzed and raw solutions of SAs were evaluated using one-way analysis of variance
158 (ANOVA).

159 2.4 Biototoxicity experiment

160 The *Vibrio fischeri* bioassay was performed to assess the photo-induced toxicity evolution of
161 SAs during the photo-degradation process. 15 min acute ecological toxicity was determined
162 using an international standard method (ISO11348-3-2007), with test conditions optimized.
163 Luminous intensities were measured using HACH Eclox water-quality detector (USA). The
164 luminescence inhibition rates ($I\%$) were derived according to Eq. (1):

$$165 \quad I\% = \frac{(I_0 - I_{15})}{I_0} \times 100\% \quad (1)$$

166 where I_0 and I_{15} stand for the luminescence intensities of the photo-degradation samples at 0
167 min and 15 min, respectively. Statistical significance was accessed at a level of $p = 0.05$ with
168 SPSS version 27.0. In order to predict the photoinduced risk to multi-level organisms, the acute
169 toxicity of individual SAs and photolytic products to fish, daphnia, and algae was evaluated
170 using the Ecological Structure Activity Relationship model (ECOSAR, version 2.2).

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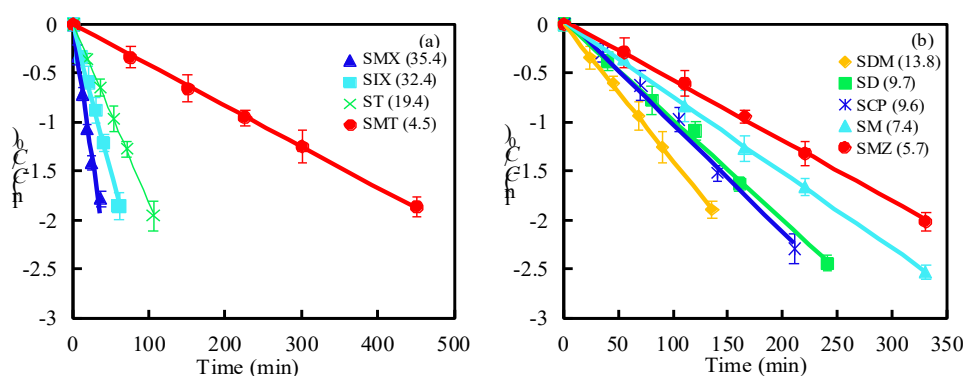
172 **3. Results and discussion**

173 *3.1 Apparent photolysis kinetics*

174 No significant loss of SAs was found in dark controls, suggesting that the antibiotic
175 compounds selected for study were unaffected by other degradation reactions (e.g., thermal or
176 hydrolytic degradation) during the photolytic experiments. However, the nine SAs absorbed the
177 actinic portion of illumination lights and suffered from rapid photo-degradation under simulated
178 sunlight ($\lambda > 290$ nm). The parent SA concentrations were confirmed to undergo logarithmic
179 decay during the attenuation by photo-degradation, indicating that the reaction followed
180 pseudo-first-order kinetics with a good linear regression ($r^2 > 0.95$) between $\ln(C/C_0)$ and time
181 (t). Their kinetic rate constants (k) and half-lives ($t_{1/2}$) are provided in [Table S3](#). The
182 corresponding logarithmic decay of the nine SAs has been plotted in [Fig. 1](#). When comparing
183 their photo-degradation kinetics of the study chemicals, SMX photo-degradation was the fastest
184 ($k = (35.4 \pm 3.1) \times 10^{-3} \text{ min}^{-1}$) among SAs with five-membered heterocycle groups, followed
185 by SIX, ST and SMT. Whereas, among SAs with six-membered heterocyclic groups, SDM
186 photo-degradation was the fastest ($k = (13.8 \pm 0.7) \times 10^{-3} \text{ min}^{-1}$), followed by SD, SCP, SM
187 and SMZ. The $t_{1/2}$ values of nine SAs increased by one order of magnitude from 12.2 min for
188 SMX to 153.3 min for SMT.

189 The observations in [Fig. 1](#) demonstrated the apparent photolytic difference across the nine
190 SAs, which can be distinguished based on their chemical structures ([Table S2](#)). SAs contain an
191 identical backbone structure (sulfanilamido group) but differ solely in their heterocyclic
192 substituents. Thus, these heterocyclic groups and associated substituents are key determinants
193 of the distinct photochemical behavior of various SAs in water. Generally, the photolytic rates
194 of most SAs (i.e. ST, SIX, SMX) with five-membered heterocycle groups were faster than all
195 SAs (i.e., SMZ, SM, SCP, SD, SDM) with six-membered heterocycle groups. Furthermore, the
196 SA degradation kinetics were found to be also dependent on the number of methyl groups

197 ($-\text{CH}_3$) associated in the similar heterocyclic groups. For instance, the SAs with similar
 198 heterocycle groups (five-membered heterocyclic SMX and SIX; six-membered heterocyclic SD,
 199 SM and SMZ), the rate constant k decreased relative to the increase of the $-\text{CH}_3$ numbers. In
 200 previous studies, faster photo-degradation rates for the five-membered heterocyclic SAs were
 201 also observed across several SAs (Voigt et al., 2017; Boreen et al., 2005; 2004). However, the
 202 influence of associated $-\text{CH}_3$ numbers on the photolysis rates was under-represented in the
 203 literature. The kinetic dependence on chemical structure suggested that SAs might photo-
 204 degrade at rates based on heterocyclic groups and associated substituents (Boreen et al., 2004;
 205 Zhou et al., 2019). Whereas, the proposed photolytic site needs to be verified by the product
 206 identification of SAs.

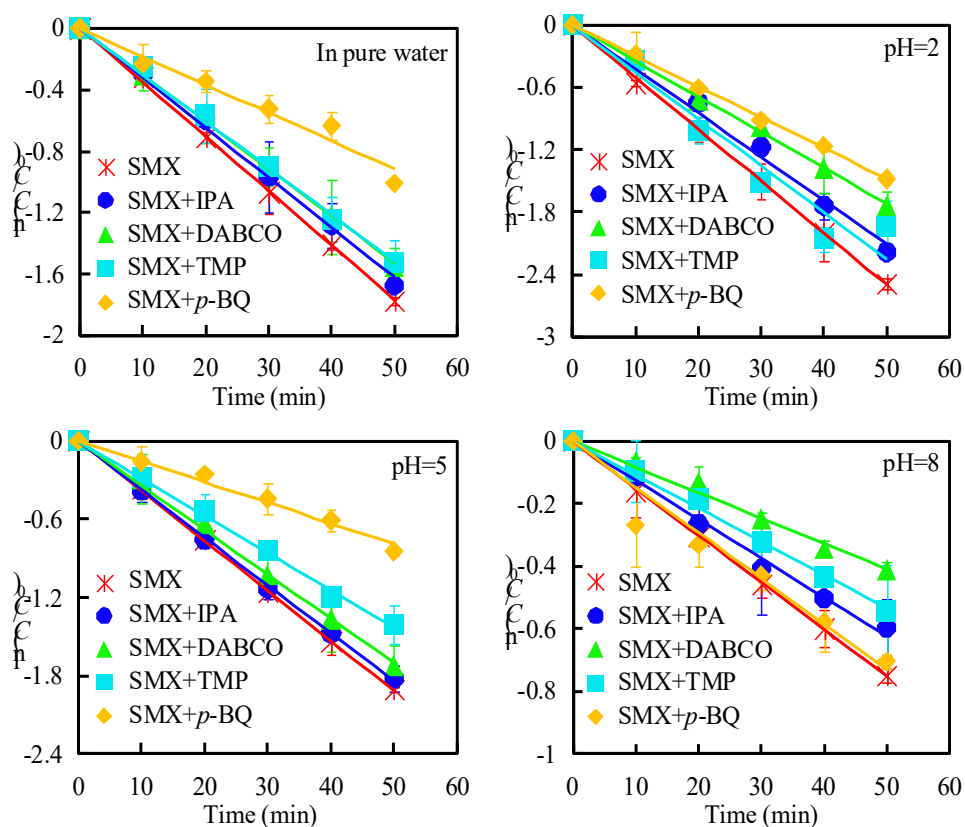


207
 208 **Fig. 1.** Apparent photo-degradation kinetics of nine SAs (rate constants $k \times 10^3 \text{ min}^{-1}$) under simulated
 209 sunlight irradiation: (a) SAs with five-membered heterocyclic groups; (b) SAs with six-membered
 210 heterocyclic groups.

211 3.2 Contribution of self-sensitized photo-oxidation

212 As shown in Fig. S5, it is hypothesized that SAs undergo direct photo-degradation via excited
 213 singlet ($^1\text{SAs}^*$) and triplet states ($^3\text{P}^*$), as well as self-sensitized photo-degradation initiated by
 214 reactive oxygen species (ROS, e.g., $\bullet\text{OH}$, $^1\text{O}_2$ and $\text{O}_2^{\bullet-}$) during their apparent photolysis. Herein,
 215 these ROS are generated through energy transfer from $^3\text{P}^*$ to H_2O or dissolved O_2 . To verify the
 216 hypothesis, the ROS and $^3\text{P}^*$ scavenging experiments were carried out using SMX as a
 217 representative SA. As shown in Fig. 2, the addition of IPA ($\bullet\text{OH}$ scavenger), DABCO ($\bullet\text{OH}/^1\text{O}_2$

218 scavenger), *p*-BQ ($O_2^{\cdot-}$ scavenger) and TMP ($^3P^*$ scavenger) induced pronounced inhibitions
 219 of SMX photo-degradation of ($p < 0.05$). It was demonstrated that SMX not only suffered from
 220 the self-sensitized photo-oxidation via $\cdot OH$, 1O_2 and $O_2^{\cdot-}$, but also underwent direct photo-
 221 degradation via $^3P^*$. The scavenging reaction kinetic parameters (k and $t_{1/2}$), as shown in Table
 222 S4, were found to be dependent on solution pH (2, 5 and 8).



223

224

225 **Fig. 2.** The effects of adding different scavengers on apparent photolysis of SMX in pure water and at
 226 different pHs.

227 Furthermore, the data demonstrated that the contribution from different micro-reactions ($R_{\cdot OH}$,
 228 $R_{^1O_2}$, $R_{O_2^{\cdot-}}$ and $R_{^3P^*}$) to SMX apparent photolysis under different conditions were quantified
 229 (Table 1). The contribution of $O_2^{\cdot-}$ induced self-sensitized photo-oxidation (40.71%–59.01%)
 230 was verified to be higher than that of all other reactive species in pure water and under acidic
 231 conditions (pHs 2 and 5), implying that $O_2^{\cdot-}$ played a major role in the self-sensitized photo-
 232 degradation of SMX. This can be attributed to the acidic solutions facilitating the generation of
 233 $O_2^{\cdot-}$ from $^3P^*$, and subsequently enhancing oxidation of SMX towards $O_2^{\cdot-}$. However, the

234 contribution of $^1\text{O}_2$ mediated self-sensitized photo-oxidation (29.13%) was the highest at pH 8.
 235 This suggested the increasing formation of $^1\text{O}_2$ from $^3\text{P}^*$ in the alkaline conditions. This
 236 phenomenon can be ascribed to the pH effect on the dissociated forms of the model compound
 237 SMX, which in turn altered the contribution of ROS mediated self-sensitized photo-oxidation
 238 (Ge et al., 2025a).

239 **Table 1**

240 The contribution rates (R_{RS}) of self-sensitized photo-degradation via ROS and direct photo-degradation via
 241 $^3\text{P}^*$ to apparent photolysis of SMX in pure water and under different pH conditions.

Reaction conditions	$R_{\bullet\text{OH}}$	$R_{^1\text{O}_2}$	$R_{\text{O}_2^{\bullet-}}$	$R_{^3\text{P}^*}$
Pure water	8.19%	5.64%	48.30%	14.41%
pH = 2	15.56%	15.76%	40.71%	9.78%
pH = 5	3.65%	7.31%	59.01%	25.32%
pH = 8	17.21%	29.13%	3.31%	28.47%

242 Although $\bullet\text{OH}$ was not the dominant ROS for SMX degradation under the range of pH values
 243 (Table 1), it was still anticipated to efficiently react with SMX in sunlit surface waters due to
 244 its high reactivity and non-selectivity (Kolasinski, 2025). At various pHs, the bimolecular
 245 reaction rates constants ($k_{\text{RS,SMX}}$) were determined to quantify the reactivity of SMX towards
 246 $\bullet\text{OH}$, $^1\text{O}_2$ and $^3\text{P}^*$ (Tables S5 and S6). It was observed that the $k_{\text{RS,SMX}}$ values exhibited a
 247 significant pH dependence ($p < 0.05$), with the highest oxidation reactivity of SMX towards
 248 $\bullet\text{OH}$, $^1\text{O}_2$ and $^3\text{P}^*$ at pH 8. This was attributed to the different molecular morphologies and
 249 deprotonation degrees at various pHs (Ge et al., 2019; Ge et al., 2024). These apparent
 250 photolytic reactions might act on the photochemical risk and fate of SAs, urging further research
 251 into their transformation pathways and photo-modified toxicity.

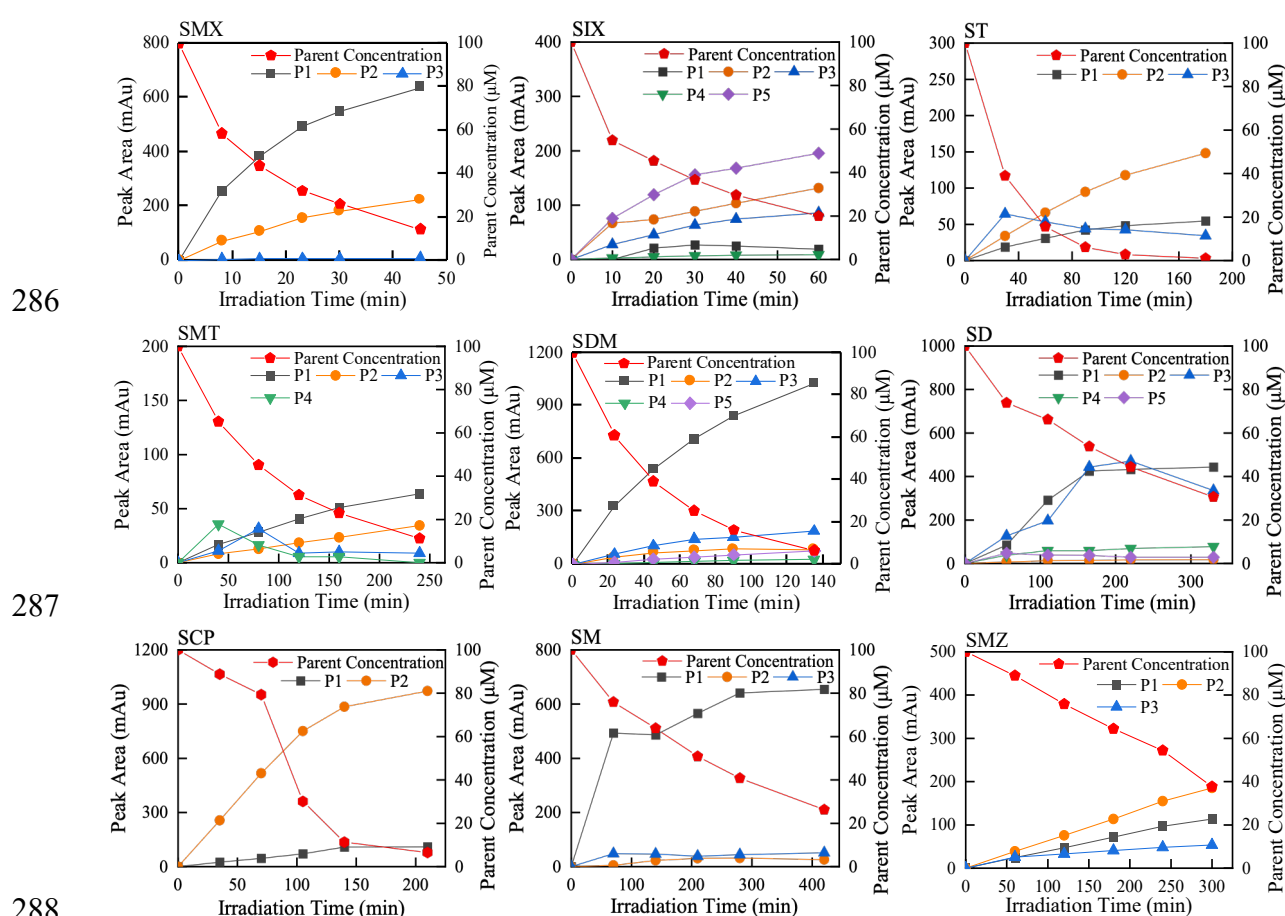
252 3.3 Photo-transformation products and pathways

253 Based on the chromatographic separation and mass spectrometry analysis, the evolution of

254 thirty-three intermediates with time shown is in Fig. 3. The chromatographic peak areas of ST-
255 P3, SMT-P3 and SMT-P4 showed a parabolic tendency, characterized by an initial rise and a
256 subsequent decline in their abundance. The peak areas of SD-P2, SD-P5, SM-P2 and SM-P3
257 showed no significant increase with extended illumination time. These trends indicated that
258 these intermediates were photo-reactive and underwent further photo-degradation. As for SMX-
259 P1, SDM-P1, SCP-P2 and SM-P1, their peak abundance increased with irradiation time,
260 suggesting lower reactivity of these products towards light exposure. Furthermore, given the
261 higher degradation rates (apparent photolytic kinetics shown in Fig. 1), SMX and SDM were
262 selected as model compounds to investigate their photo-degradation intermediates and
263 transformation pathways under simulated solar irradiation. Their total ion currents (TIC) are
264 shown in Fig. S6, from which eight significant intermediates were identified from SMX and
265 SDM. The molecular weights (M_w), retention times (t_R) and MS fragment m/z are summarized
266 in Table S7 and Fig. S7. As illustrated in Fig. 4, their corresponding chemical structures and
267 photo-transformation pathways are proposed.

268 The photo-transformation of SMX with five-membered heterocyclic groups mainly involved
269 two pathways: photo-cleavage and photo-polymerization (Fig. 4). In the photo-cleavage
270 pathway (Path 1), apparent photolysis occurs via S–N bond cleavage, yielding sulfanilic acid
271 (SMX-P2) as a common byproduct (Boreen et al., 2004). In Path 2, SMX-P3 was formed
272 through addition reaction at the sulfonyl-N site. As for SDM containing six-membered
273 heterocyclic substituents, four photo-degradation pathways were identified. In Path 1, the
274 transformation pathways mainly involved cleavage of S–N bond (SDM-P1), hydroxylation and
275 desulfonation (SDM-P4). Hydroxylation is due to the presence of the electron-rich aniline
276 moiety, which in turn enables further $\bullet\text{OH}$ addition (Shah and Hao, 2017; Bilea et al., 2024). In
277 Path 2, SDM was primarily transformed into a desulfonation product (SDM-P2) with the loss
278 of SO_2 . The mechanism for SO_2 extrusion involved the aromatic nucleophilic substitution of an

279 aniline radical cation, followed by a Smiles rearrangement (Ji et al., 2017). Furthermore, this
 280 pathway also involved the amino-N removal. In Path 3, SDM first suffered methylation, and
 281 then underwent addition reaction at sulfonyl-N, yielding SDM-P3. In Path 4, SDM was
 282 primarily transformed into SDM-P5 via the introduction of a $-OCH_3$ to the six-membered
 283 heterocyclic group. Combined with the abundant detection of photo-products (Fig. 3), the
 284 photo-cleavage process with cleavage of S-N bond was confirmed as the primary
 285 transformation pathway for SMX and SDM.



289 **Fig. 3.** The abundance and evolvement of main photo-products (peak area) and during the photo-degradation
 290 of nine SAs (parent concentration)

291 In environmental surface waters, SAs have been shown to be ubiquitous and would undergo
 292 many photochemical reactions including the apparent photolysis (Hernández-Tenorio, 2024;
 293 Loureiro Dos Louros et al., 2020) and ROS-induced sensitized photo-degradation (Andino-
 294 Enríquez et al., 2025; Liu et al., 2023b; Qin et al., 2024). However, results from this present

295 study elucidated the multifarious photolytic intermediates and pathways of SAs, providing
 296 fundamental insights into the phototransformation fate of this class of organic contaminants. It
 297 is worth noting that the photochemical reactions did not initially affect the main backbone
 298 structure of the SAs, suggesting that their antibacterial capacity might persist in those
 299 intermediates (Niu et al., 2017). Therefore, it is necessary to further examine the antimicrobial
 300 activities and biology toxicity of these transformation products.

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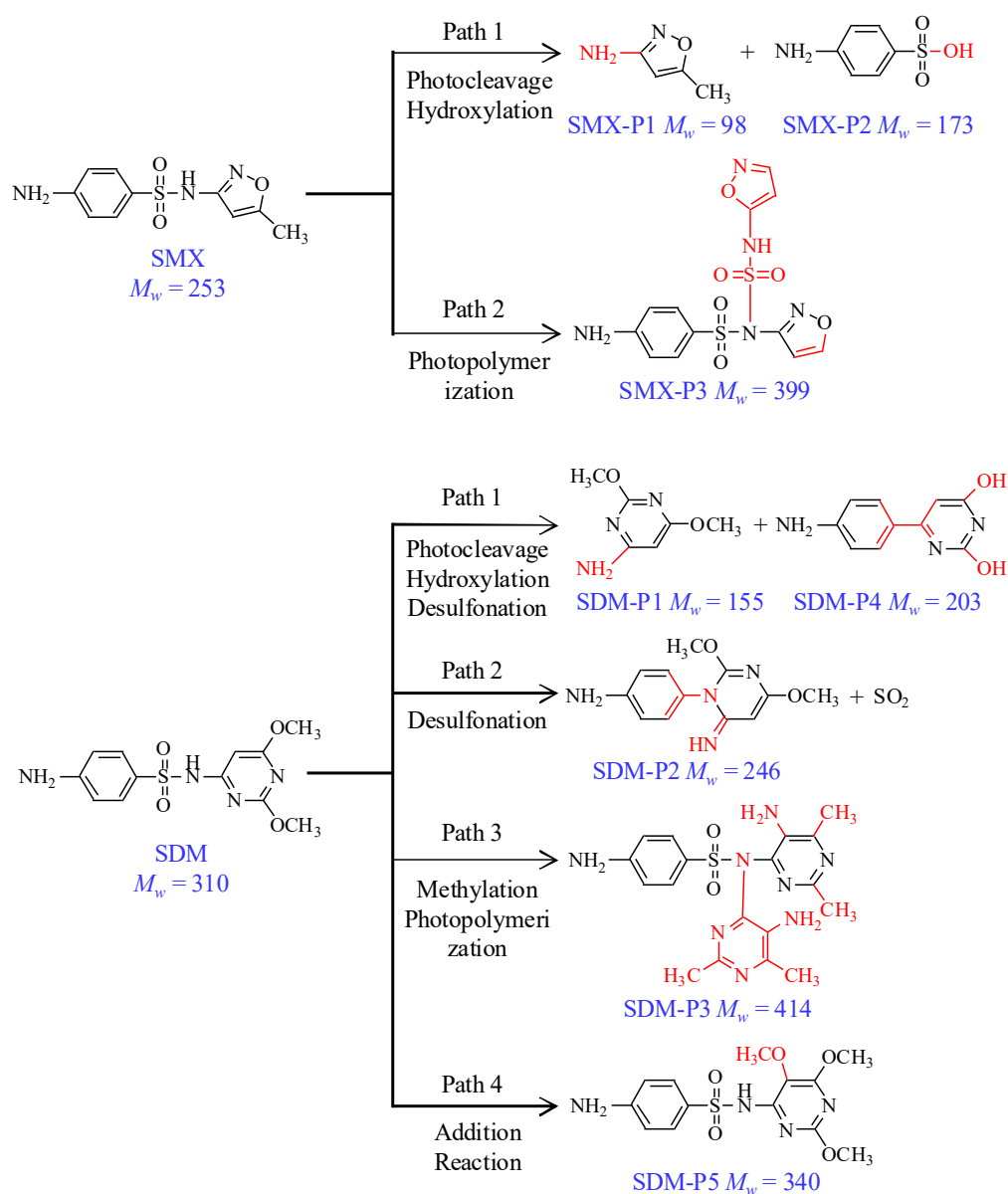


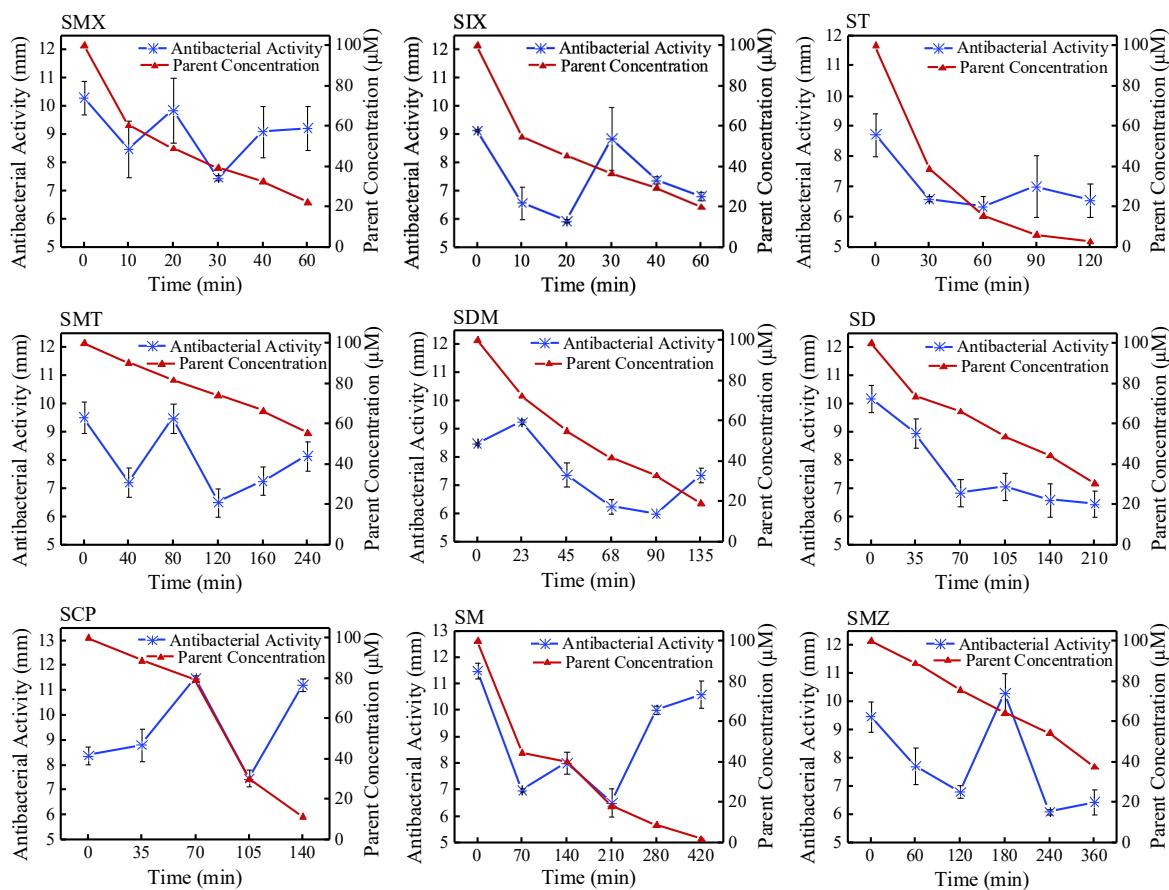
Fig. 4. Transformation products and pathways for apparent photo-degradation of five-membered heterocyclic SMX and six-membered heterocyclic SDM under simulated sunlight irradiation.

3.4 Changes in antibacterial activities against *E. coli*

306 The antibacterial activities of nine SAs were evaluated using *E. coli* as a bioindicator. As
307 depicted in Fig. S8, the initial antibacterial activities of raw SMX, SD and SM (10.31 mm,
308 10.20 mm and 11.50 mm respectively) were higher ($p < 0.05$) than those of the other 6 SAs
309 which range from 8.49 mm to 9.52 mm. This higher activity can be attributed to their lower
310 molecular weights, which reduces steric hindrance and facilitates their entry into *E. coli* cells,
311 thereby producing more significant antibacterial effects (Baek et al., 2019).

312 During apparent photolysis of SAs (Fig. 5), the antibacterial activities evolved in distinct
313 patterns as the parent compounds decayed and intermediates were formed. With increasing
314 illumination time, the antibacterial activities of ST and SD against *E. coli* showed a general
315 declining trend and lower than their initial values ($p < 0.05$). However, the antibacterial
316 activities of other SAs exhibited a fluctuating pattern. SDM and SCP first showed an increase
317 followed by a decrease in antibacterial activities ($p < 0.05$) at the initial illumination period.
318 This suggested that the two SAs generated photo-modified products with notable antibacterial
319 activities against *E. coli*. In comparison, the other five SAs displayed a significant decrease
320 followed by an increase ($p < 0.05$), indicating that short-time illumination reduced their
321 antibacterial activities. However, extending illumination time induced the formation of
322 antibacterial photo-products, which might be generated from photo-transformation pathways
323 (Fig. 4). Thus, the antibacterial activity evolution during the photo-degradation of SAs are
324 dependent on primary products and pathways. As for other classes of antibiotics, many photo-
325 transformation products of fluoroquinolone (FQs) and tetracycline (TCs) also exhibited
326 antibacterial activity against *E. coli*. (Huang et al., 2025; Guo et al., 2025b).

327 Apart from antibacterial activity, toxicity assessment is a critical component of the
328 environmental risk assessment for aquatic contaminants (Ge et al., 2025b; Ge et al., 2025c).
329 Therefore, it is essential to further evaluate the changes in toxicity of SAs to *Vibrio fischeri*, to
330 predict the toxicity of individual photo-products to multiple-level aquatic organisms.



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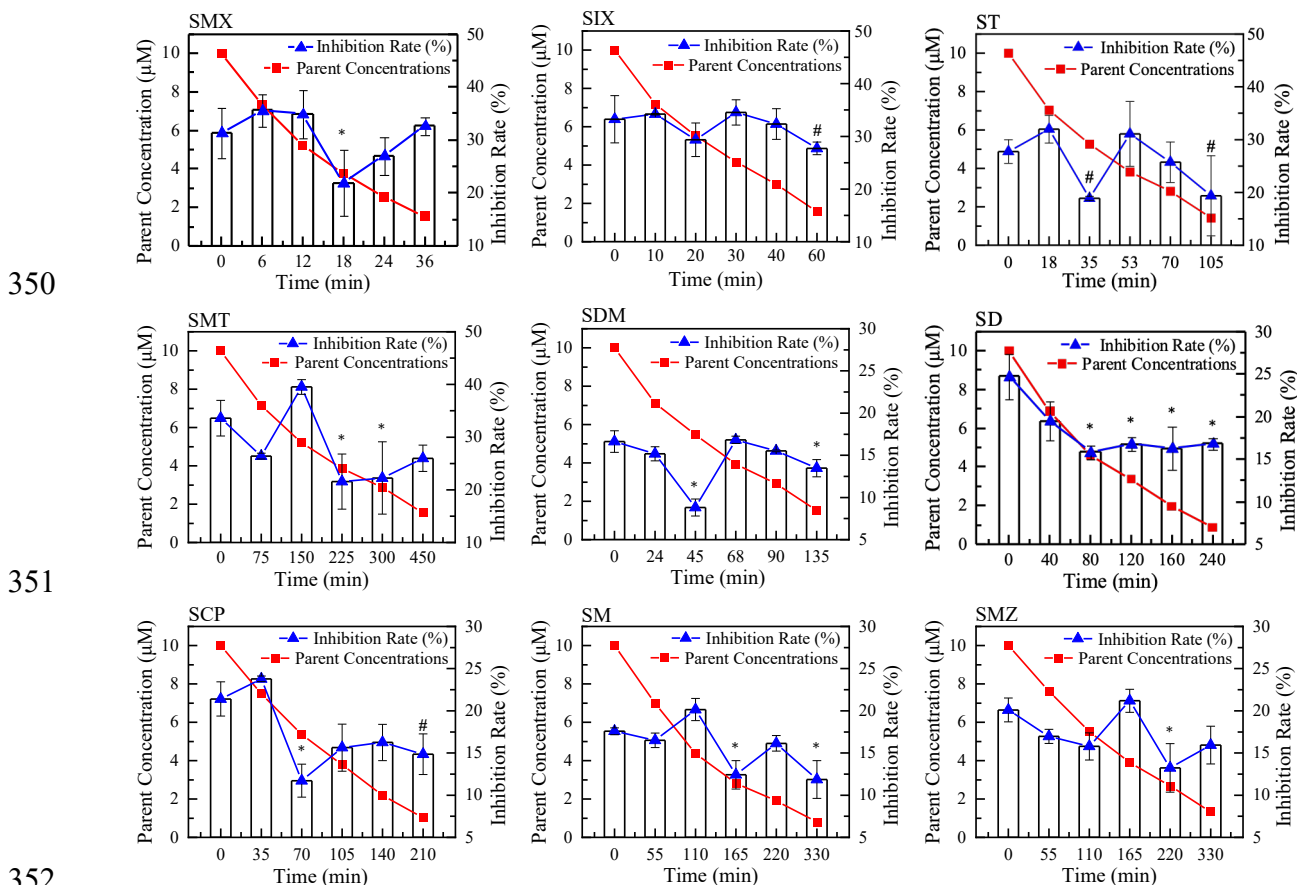
333

334 **Fig. 5.** Changes of nine SAs parent concentrations and antibacterial activity of the photo-degradation
 335 solutions to *E. coli* with irradiation time in water. The y-axis represents the agar plated bacteria-free zone
 336 diameter.

337 3.5 Photo-modified toxicities to *Vibrio fischeri*

338 The toxicities of the nine parent SAs were first assessed using *Vibrio fischeri*. As shown in
 339 Fig. 6, their luminescence inhibition rates ($I\%$) ranged from 16.64% for SDM to 33.58% for
 340 SMT. During the apparent photo-degradation, their $I\%$ values exhibited diverse evolution
 341 curves with the declining concentrations of the parent compounds. The observed toxicity
 342 fluctuations were a result of the continuous formation and degradation of intermediates with
 343 varying toxicity. In most cases, the maximal $I\%$ values of the photolytic solutions were
 344 observably greater than those of the raw SAs ($p < 0.05$), indicating the formation of more toxic
 345 photo-products. Furthermore, the toxicity of individual products was assessed by ECOSAR
 346 version 2.2 software. Based on the results (acute toxicity and chronic toxicity) shown in Table

347 S8, it can be seen that SMX and SDM photo-transformed into several intermediates with
 348 comparable or higher toxicity, such as SMX-P1 and SDM-P4. Importantly, the toxicity results
 349 agreed well with the *Vibrio fischeri* bioassay.



352
 353 **Fig. 6.** Time-dependent changes in parent SA concentrations and luminescence inhibition rates to *Vibrio*
 354 *fischeri* during photolysis.

355 According to the photo-degradation products and primary pathways (Fig. 4), a mechanism
 356 for toxicity evolvement has been proposed. The cleavage of the sulfonamide bond (S–N)
 357 increases the number of –NH₂ groups that interact with *flavin mononucleotide*, a significant
 358 process for the luminescence inhibition of *Vibrio fischeri* (Jablonski and DeLuca, 1978).
 359 Meanwhile, the reduction in molecular weight and steric hindrance from photo-cleavage can
 360 facilitate penetration into *Vibrio fischeri* cells, thus promoting the observed increase in toxicity
 361 (Park et al., 2023). Moreover, products of addition reactions such as SDM-P4 are likely to
 362 exhibit higher toxicity than their parent compounds. Consequently, given the photo-enhanced

363 potential threat of the intermediates to aquatic systems, evaluating the photo-modified toxicity
364 of emerging contaminants like SAs should be included in their environmental risk assessment
365 (Ge et al., 2025a; Zheng et al., 2025).

366 4. Conclusion

367 This research provides a comprehensive investigation of the reaction kinetics, self-sensitized
368 photo-oxidation, transformation pathways, antibacterial activity changes and photo-induced
369 toxicity during the apparent photo-degradation process of nine widely detected SAs in the
370 aqueous environment. Although their apparent photo-degradation followed pseudo-first-order
371 kinetics ($t_{1/2} = 12.2\text{--}153.3$ min), most SAs (i.e. ST, SIX, SMX) with five-membered heterocycle
372 groups photodegraded faster than all SAs (i.e., SMZ, SM, SCP, SD, SDM) with six-membered
373 heterocycle groups. This apparent photolytic difference can be ascribed to their different
374 chemical structures including diverse heterocyclic groups and associated substituents (e.g.,
375 $-\text{CH}_3$). As a representative, SMX not only underwent self-sensitized photo-oxidation via $\text{O}_2^{\cdot-}$,
376 $^1\text{O}_2$ and $\cdot\text{OH}$, but also experienced direct photo-degradation via $^3\text{P}^*$. The contribution of $\text{O}_2^{\cdot-}$
377 (40.71%–59.01%) and $^1\text{O}_2$ (29.13%) mediated self-sensitized photo-oxidation was confirmed
378 to be the highest under acidic and alkaline conditions, respectively, which was attributed to the
379 different deprotonation degrees and RS reactivity at various pHs. Through the apparent photo-
380 reactions, these SAs transformed into intermediates with different photo-reactivity. The
381 transformation pathways of five-membered heterocyclic SMX involved photo-cleavage and
382 photo-polymerization, while six-membered heterocyclic SDM underwent cleavage of the S–N
383 bond, hydroxylation, desulfonation, methylation and addition reaction at sulfonyl-N.
384 Furthermore, both the antibacterial activity changes and photo-modified toxicity evolution were
385 found to be dependent on dominant photo-degradation pathways and primary intermediates.
386 Importantly, the toxicity predictions from the ECOSAR software for the individual
387 intermediates agreed with the *Vibrio fischeri* bioassay results.

388 This study not only filled a critical gap in understanding the RS-mediated apparent photolytic
389 mechanisms of SAs in aqueous systems but also revealed their pathway-dependent antibacterial
390 activity changes and toxicity evolution during the apparent photo-degradation. These findings
391 are essential for accurately assessing the photochemical persistence, fate, and ecological risks
392 of the emerging contaminants in aquatic systems. Furthermore, the clarified transformation
393 mechanisms and photo-modified toxicity of SAs help to better understand the persistence and
394 risks posed by SAs and key products in photo-assisted advanced oxidation processes (Felis et
395 al., 2024; Duan et al., 2022; Li et al., 2025b).

396

397 **CRedit authorship contribution statement**

398 **Linke Ge:** Writing – review & editing, Conceptualization, Funding acquisition. **Jiahui**
399 **Wang:** Writing – original draft, Data curation. **Andrew Sweetman:** Writing – review & editing.
400 **Zhixuan Zheng:** Data curation. **Peng Zhang:** Writing – review & editing. **Nannan Cui:**
401 Writing – review & editing.

402

403 **Declaration of competing interest**

404 The authors declare that they have no known competing financial interests or personal
405 relationships that could have appeared to influence the work reported in this paper.

406

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414

415 **Appendix A. Supplementary data**

416 Supplementary data to this article can be found online at https://doi.org/*****/j.

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